

Name: \_\_\_\_\_

Chem 203  
December 10, 2011

Final Exam Part I  
(40 points)

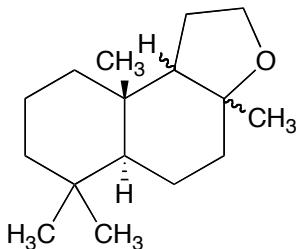
**THIS FINAL EXAM CONSISTS OF PART I  
AND TWO OUT OF THE THREE PROBLEMS FROM PART II**

**IF THREE PROBLEMS FROM PART II ARE SUBMITTED,  
ONLY THE FIRST TWO (Part II, PROBLEMS 1-2) WILL BE GRADED**

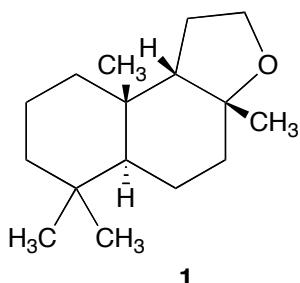
If you wish to unstaple the pages, please initial each page.

Books, notes, lecture videos, calculators, rulers, and laptop computers are permitted as is wireless (or wired) internet access and appropriate software (e.g., PyMOL, Maestro/MacroModel, Excel, ChemDoodle, Chemdraw, ElComp, MolE, etc.). Communication with other students by e-mail, text, or in person is not permitted. Catalogs of molecular structures (e.g., the Aldrich catalog, the Merck Index, etc.) or databases of molecular structures (such as wireless access to SciFinder Scholar, the Sigma-Aldrich website, etc.) are NOT PERMITTED. INAPPROPRIATE COMMUNICATION OR USE OF SUCH ITEMS CONSTITUTES ACADEMIC DISHONESTY, WILL RESULT IN A FAILING GRADE (F) IN THE CLASS, AND MAY RESULT IN EXPULSION FROM THE Ph.D. PROGRAM.

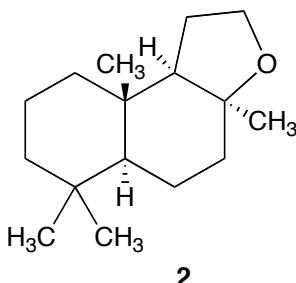
**1.** The following spectral data are provided for a tricyclic compound: 500.22 MHz  $^1\text{H}$  NMR, 125.79 MHz  $^{13}\text{C}$  NMR, DEPT, COSY, TOCSY, HMQC, HMBC, NOESY, and HSQC-TOCSY spectra with 5-, 10-, 20-, and 100-ms mixing times. All NMR spectra were measured in  $\text{C}_6\text{D}_6$  solution.



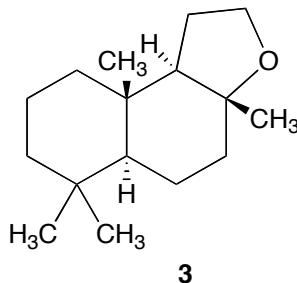
Using these data, determine the stereochemistry and assign all of the  $^1\text{H}$  and  $^{13}\text{C}$  resonances to their respective atoms in the structure. Specifically, assign the stereochemistry of the two stereocenters shown with squiggly lines, and hence which of the four possible diastereomers (**1**, **2**, **3**, or **4**) is consistent with these data.



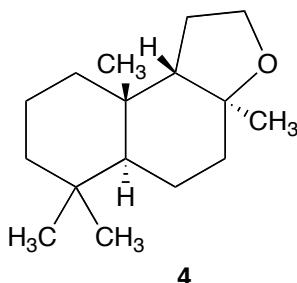
**1**



**2**



**3**



**4**

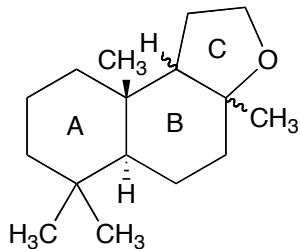
MAKE SURE TO COMPLETELY ANSWER THE QUESTIONS **a-g** ON PAGES 2-6.

**a.** Build energy-minimized molecular models of diastereomers **1**, **2**, **3**, and **4** using PyMOL and the "clean" function. Rotate each molecule into an appropriate orientation. Use the side menu (S) to show each molecule as sticks (Show-sticks). Use the pulldown menu to display each molecule in maximum quality with (Display-Quality-Maximum Quality). Save the .pse files as diastereomer1.pse, diastereomer2.pse, diastereomer3.pse, and diastereomer4.pse.

Feel free to use the *trans*-decalin template .pse file or .pdb file on the course web page "Simple Conformational Analysis of Cyclic and Bicyclic Compounds" which is linked to the "Assignments" and "Class Materials" web pages. You are also welcome to just make your own *trans*-decalin.

NOTE: For diastereomer **4** you will have to build the B ring in a boat conformation, because the B–C ring junction does not allow the B ring to adopt a chair conformation. After minimization, the B ring should be in a twist-boat conformation. The A–B ring junction will look like that of the other diasteromers, but the B–C ring junction will be enforcing a twist-boat conformation of the B ring.

If you would like to review the conformations of cyclohexane, please see my Chem H52A course web page "Cyclohexanes" at: <https://eee.uci.edu/10f/40600/cyclohexane.html>. The page contains links to .pdb files of boat and twist-boat cyclohexane, which can be opened in PyMOL. If you would like to see an example of a related exercise in which ring fusion induces a twist boat conformation in a cyclohexane ring, please see my course web page "Perhydroanthracenes" and the associated .pdb files at: <https://eee.uci.edu/10f/40600/perhydroanthracenes.html>.



E-mail the .pse files to me (jsnowick@uci.edu).

- b.** Examine the  $^1\text{H}$  NMR spectra and familiarize yourself with the resonances, which have been lettered *a*–*l* for you. Examine the  $^{13}\text{C}$  NMR spectra and number the sixteen unique resonances associated with the molecule 1–16.

Examine the DEPT spectra and identify the quaternary (C), methine (CH), methylene (CH<sub>2</sub>), and methyl (CH<sub>3</sub>) peaks. NOTE: Although the DEPT 90 contains small resonances associated with the methyl and methylene groups, it is easy to identify the methine groups.

Examine the HMQC spectra and correlate the numbers of  $^{13}\text{C}$  resonances with the letters of the  $^1\text{H}$  resonances.

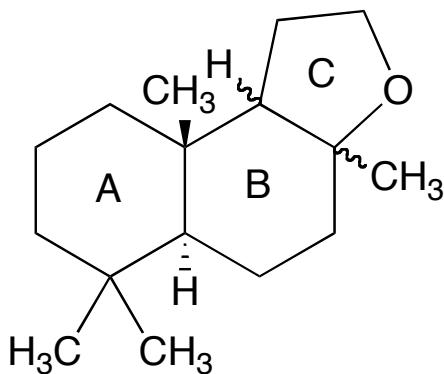
Finally, examine the HSQC-TOCSY spectrum with 100-ms mixing time and identify which methine and methylene resonances are associated with the three main spin systems in the molecule, which we will call the A-ring spin system, the B-ring spin system, and the C-ring spin system:

Numbers associated with the  $^{13}\text{C}$  resonances associated with the A-ring spin system: \_\_\_, \_\_\_, \_\_\_.

Numbers associated with the  $^{13}\text{C}$  resonances associated with the B-ring spin system: \_\_\_, \_\_\_, \_\_\_.

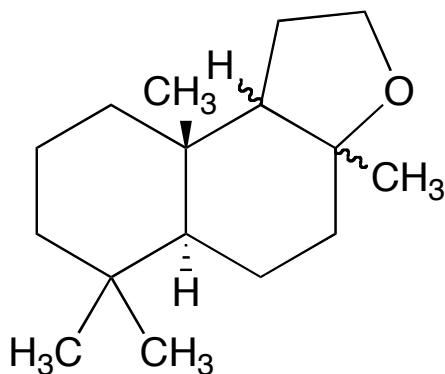
Numbers associated with the  $^{13}\text{C}$  resonances associated with the C-ring spin system: \_\_\_, \_\_\_, \_\_\_.

- c.** Examine the HSQC-TOCSY spectra with 5, 10, 20, and 100-ms mixing times. Assign the six methine and methylene  $^{13}\text{C}$  resonances from the B- and C-ring spin systems, to the corresponding atoms in the structure, below. That is, write the number next to the atom in the structure below.



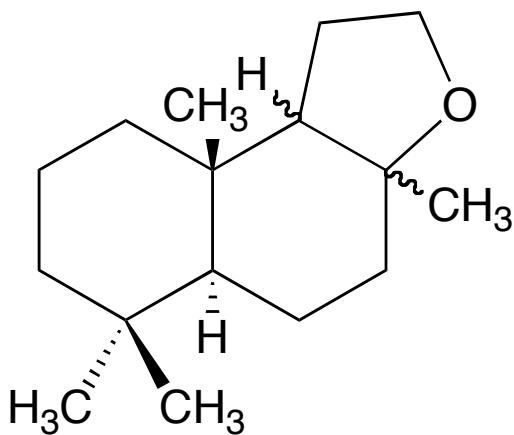
NOTE: Work the problem the way we did in class and is described in the assigned reading in "Two-Dimensional NMR Spectroscopy: Applications for Chemists and Biochemists." Start with the track that grows in under  $^1\text{H}$  resonances *a* and *b* and assign one spin system. Then work with the track that grows in under  $^1\text{H}$  resonance *c* and assign another spin system.

**d.** Use the HMBC spectrum and particularly the expansion that gives the higher-level contours of the isolated methyl groups to assign the remaining  $^{13}\text{C}$  resonances to the corresponding atoms in the structure, below. That is, write the number next to the atom in the structure below.



**e.** Use the NOESY spectrum to stereospecifically assign the diastereotopic geminal dimethyl  $^{13}\text{C}$  resonances to the corresponding atoms in the structure, below. That is, write the number next to the atom in the structure below. HINT: Consider the proximity of the methyl groups to each other; use the models that you have built in Part **a** for insights.

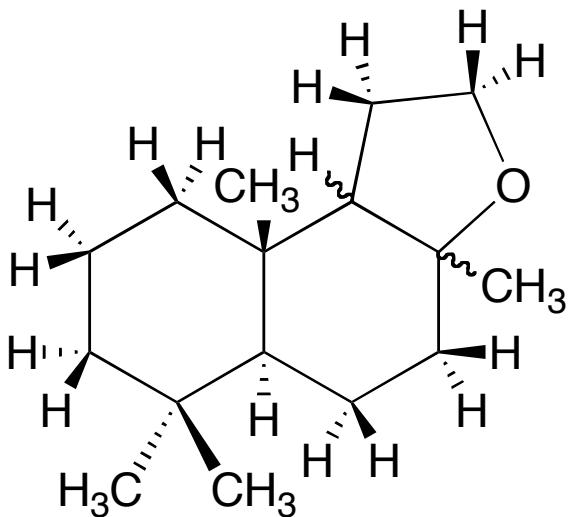
Transcribe your other numbers from Parts **c** and **d**, so that every carbon is numbered. Write the letters of the corresponding  $^1\text{H}$  resonances next to the numbers.



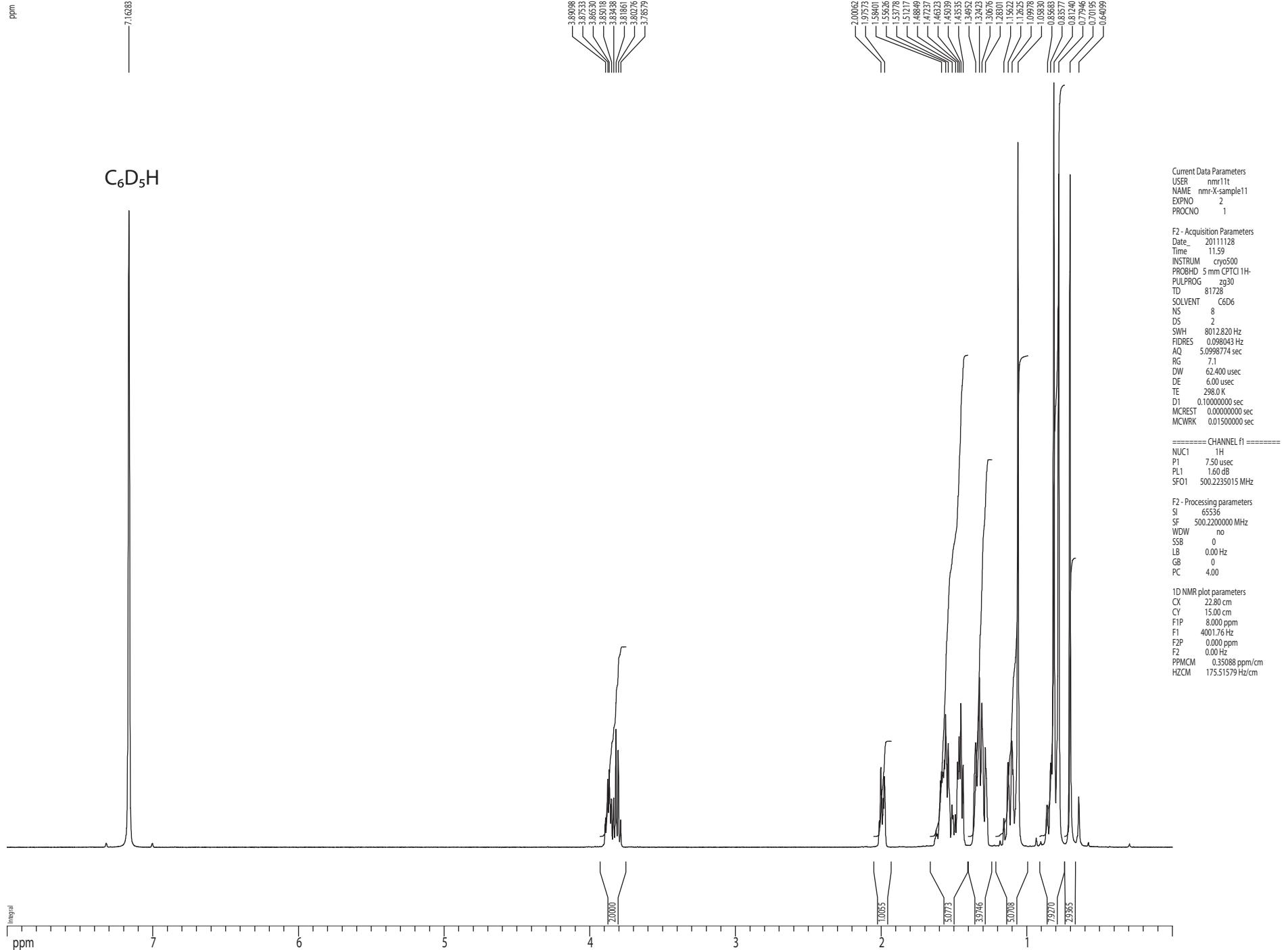
f. Determine the stereochemistry of the molecule. Which diastereomer is it? \_\_\_\_\_

Explain how you determined the stereochemistry of the molecule. Make a conformationally realistic drawing of the molecule to help aid in your explanation. If there are any aspects of the stereochemistry you are uncertain about, please include this in your explanation.

g. Insofar as possible, assign the  $^1\text{H}$  resonances to the corresponding atoms in the structure, below. That is, write the letter (a–l) next to the atom in the structure below. You will not likely be able to assign them all. Explain briefly about those that you are uncertain.



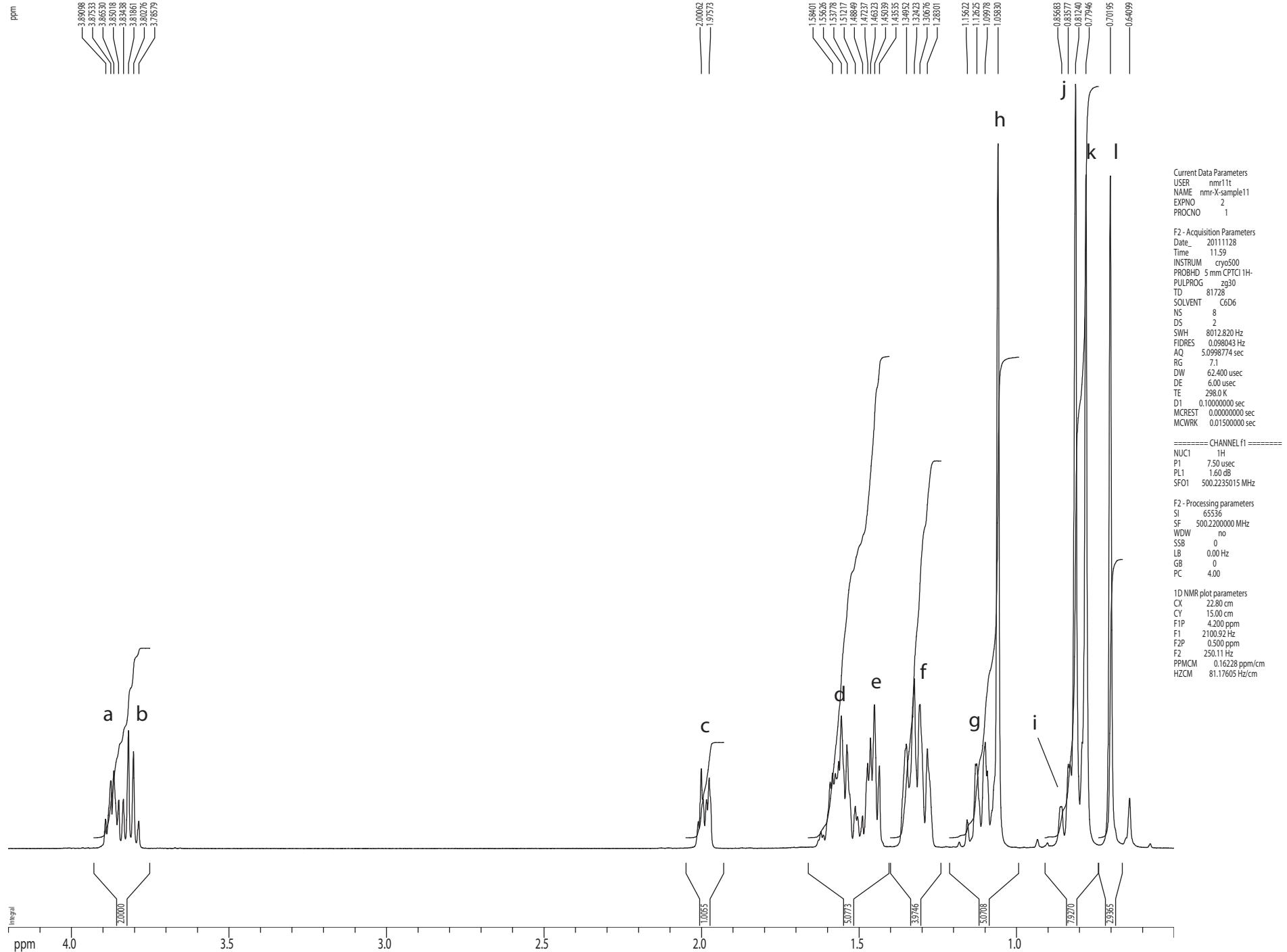
<sup>1</sup>H spectrum at 500 MHz in C<sub>6</sub>D<sub>6</sub>



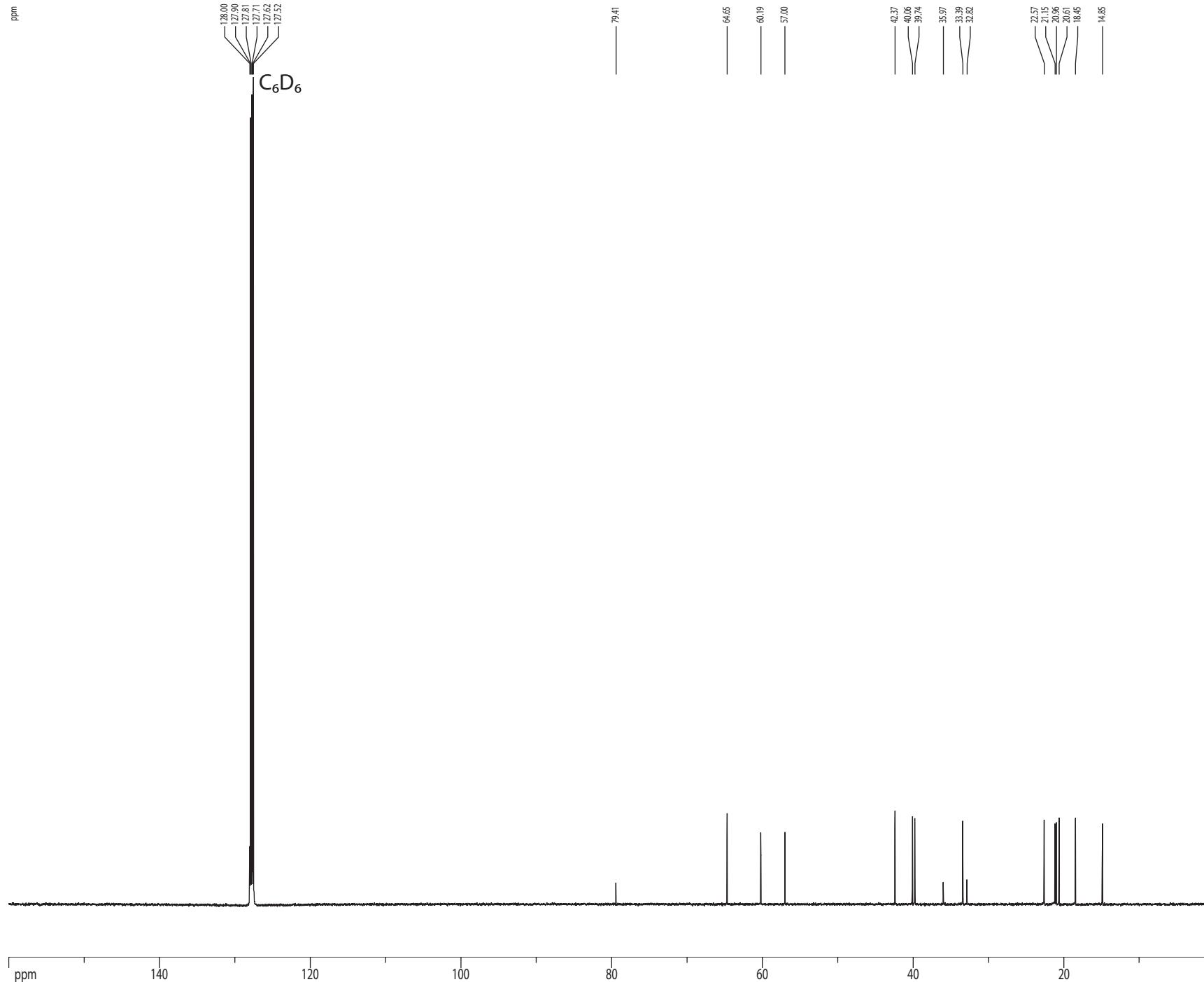
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3	45592.8	1933.502	3.8653	1.56
4	45654.7	1925.939	3.8502	0.99
5	45719.3	1918.036	3.8344	1.01
6	45783.9	1910.146	3.8186	2.35
7	45848.7	1902.217	3.8028	1.94
8	45918.1	1893.729	3.7858	0.58
9	53221.7	1000.748	2.0006	1.60
10	53323.6	988.298	1.9757	1.43
11	54926.2	792.353	1.5840	1.52
12	55039.7	778.472	1.5563	2.64
13	55115.3	769.226	1.5378	2.07
14	55220.1	756.417	1.5122	0.87
15	55317.0	744.573	1.4885	0.68
16	55382.9	736.512	1.4724	1.71
17	55420.3	731.938	1.4632	2.20
18	55472.8	725.515	1.4504	2.86
19	55534.3	717.993	1.4354	1.65
20	55885.5	675.057	1.3495	2.09
21	55989.0	662.406	1.3242	3.36
22	56060.4	653.669	1.3068	2.86
23	56157.6	641.786	1.2830	1.99
24	56676.4	578.362	1.1562	0.61
25	56799.0	563.371	1.1262	1.70
26	56907.3	550.131	1.0998	2.12
27	57077.0	529.382	1.0583	13.83
28	57901.2	428.605	0.8568	0.86
29	57987.4	418.071	0.8358	1.69
30	58083.0	406.376	0.8124	15.00
31	58217.8	389.900	0.7795	13.22
32	58534.9	351.131	0.7020	13.21
33	58784.3	320.638	0.6410	1.03

<sup>1</sup>H spectrum at 500 MHz in C<sub>6</sub>D<sub>6</sub>



Z-restored spin-echo  $^{13}\text{C}$  spectrum with  $^1\text{H}$  decoupling at 125 MHz in  $\text{C}_6\text{D}_6$



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 d11 0.0300000 sec  
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 MCWRK 0.0150000 sec  
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 P11 500.00 usec  
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 PL1 -1.00 dB  
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 SP1 3.20 dB  
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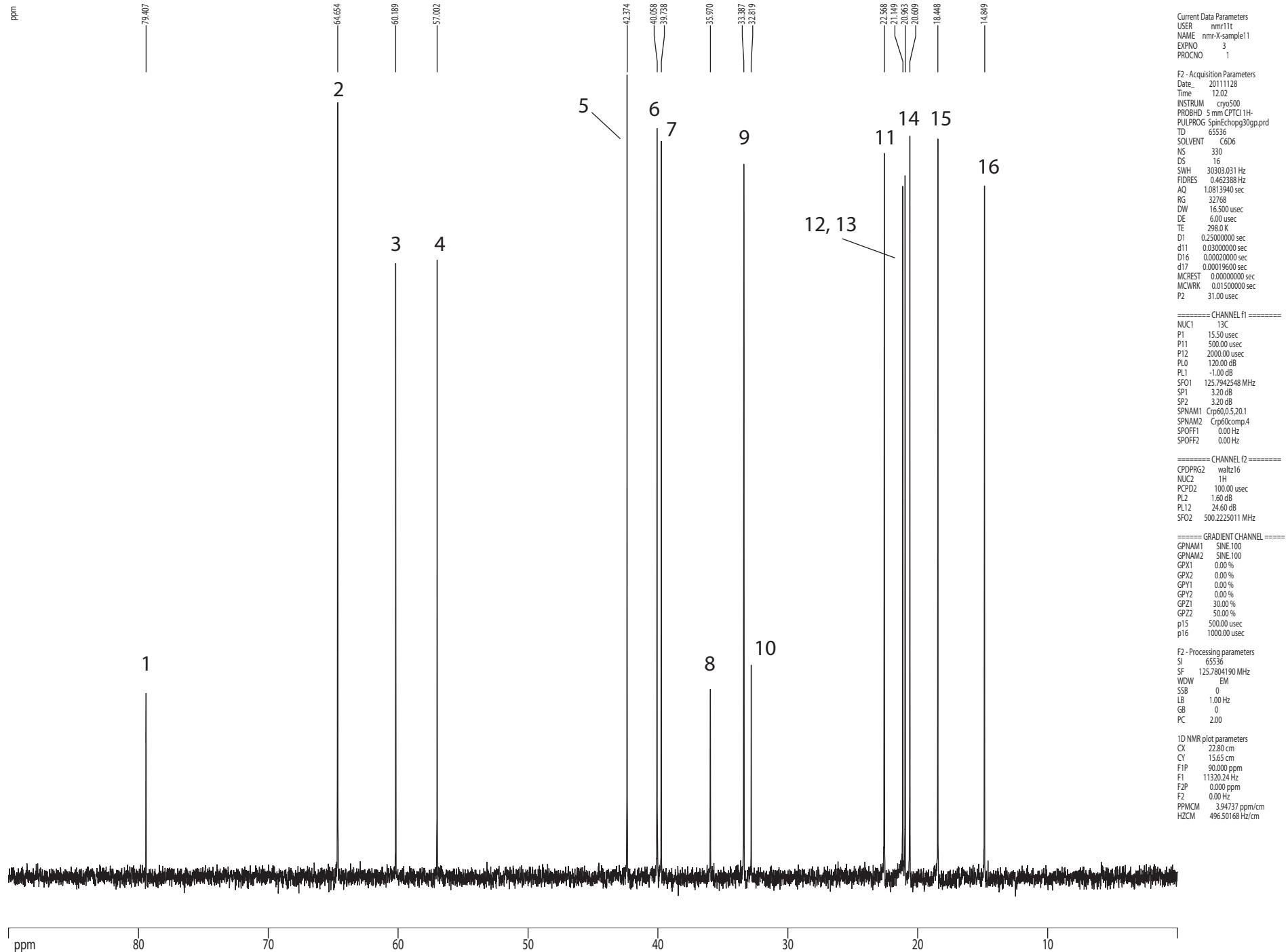
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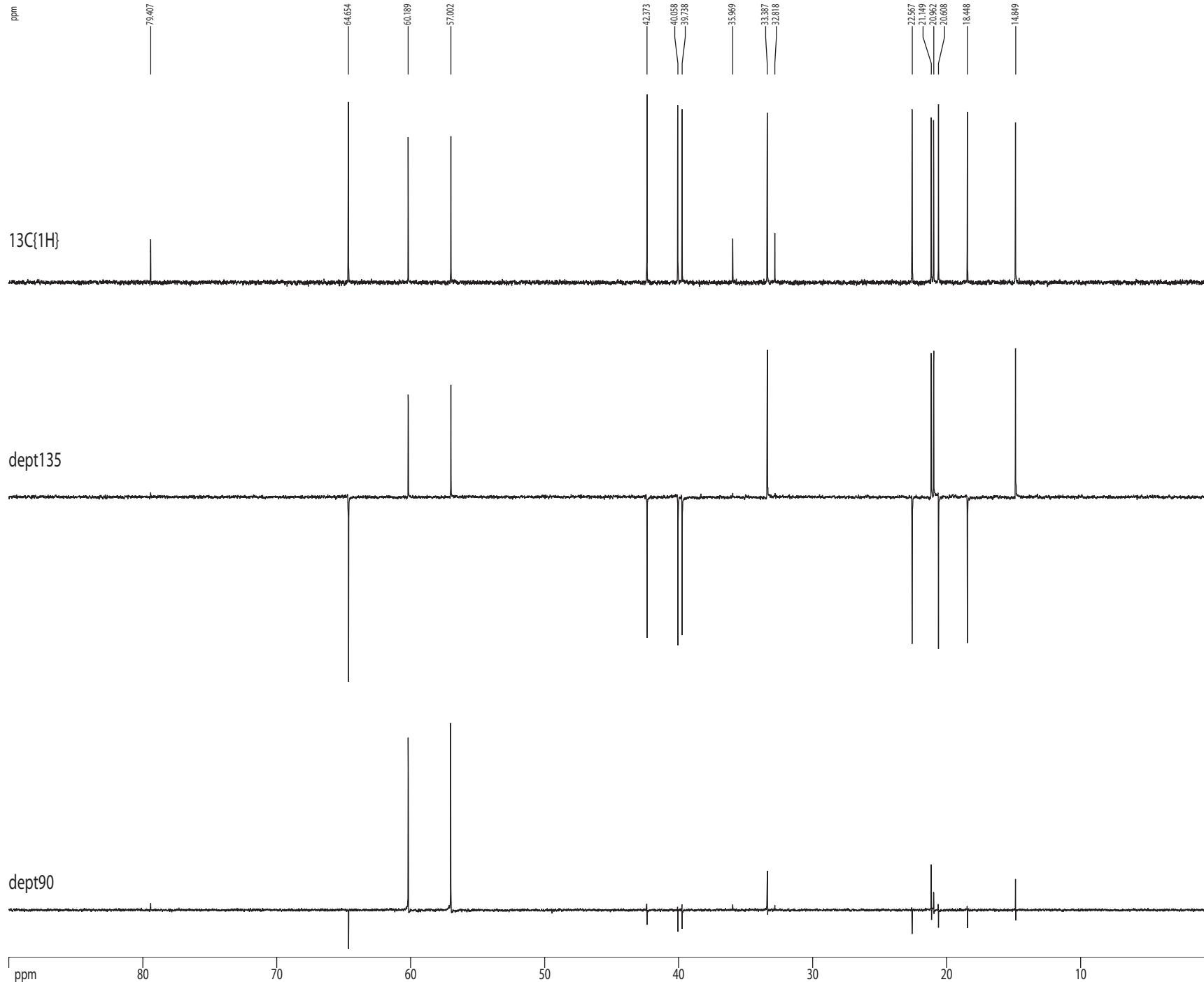
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 F2 0.00 Hz  
 PPMCM 7.01754 ppm/cm  
 HZCM 882.66962 Hz/cm

Z-restored spin-echo  $^{13}\text{C}$  spectrum with  $^1\text{H}$  decoupling at 125 MHz in  $\text{C}_6\text{D}_6$



Z-restored spin-echo  $^{13}\text{C}$  spectrum with  $^1\text{H}$  decoupling at 125 MHz in  $\text{C}_6\text{D}_6$



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===== CHANNEL f1 =====  
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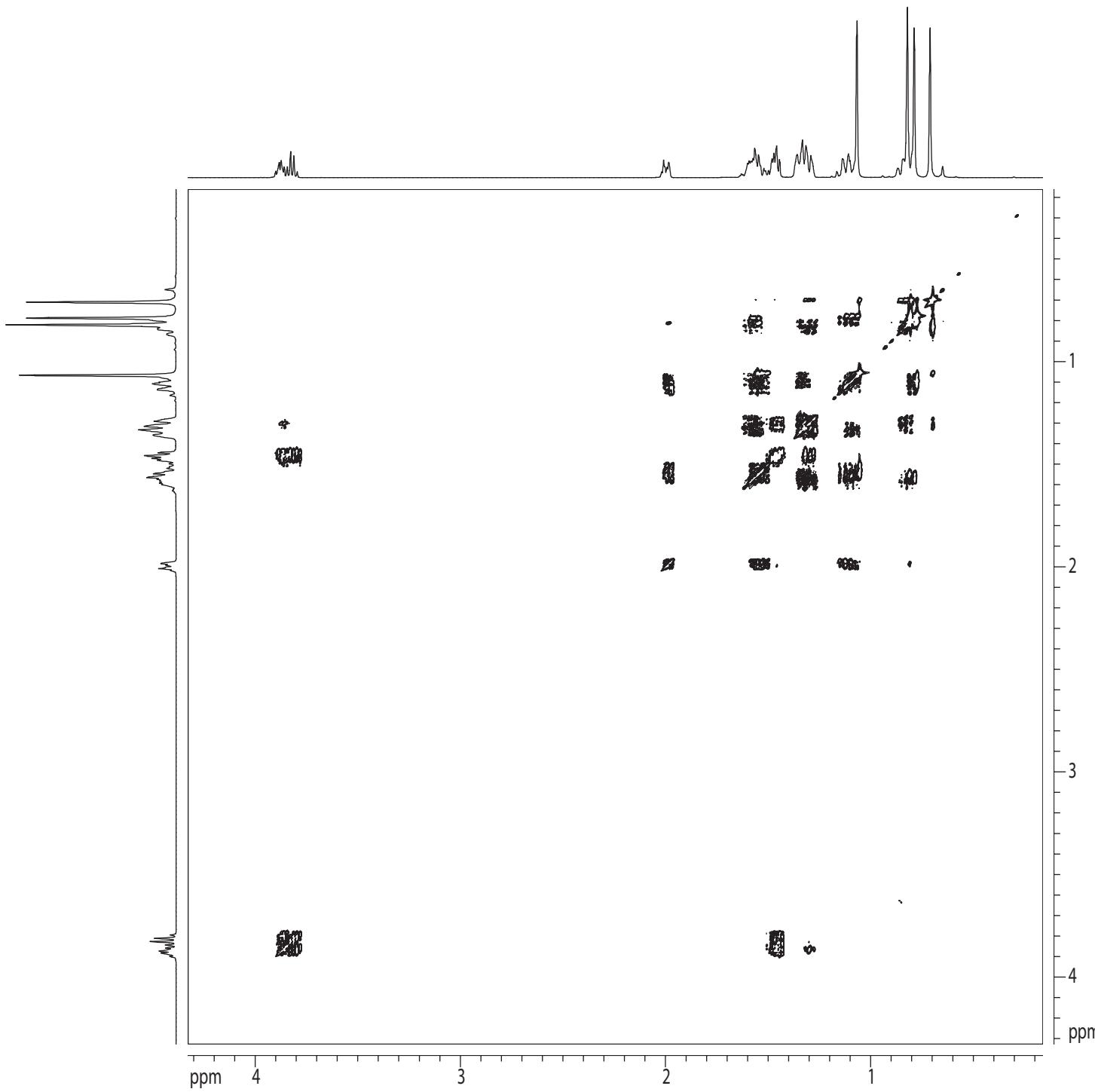
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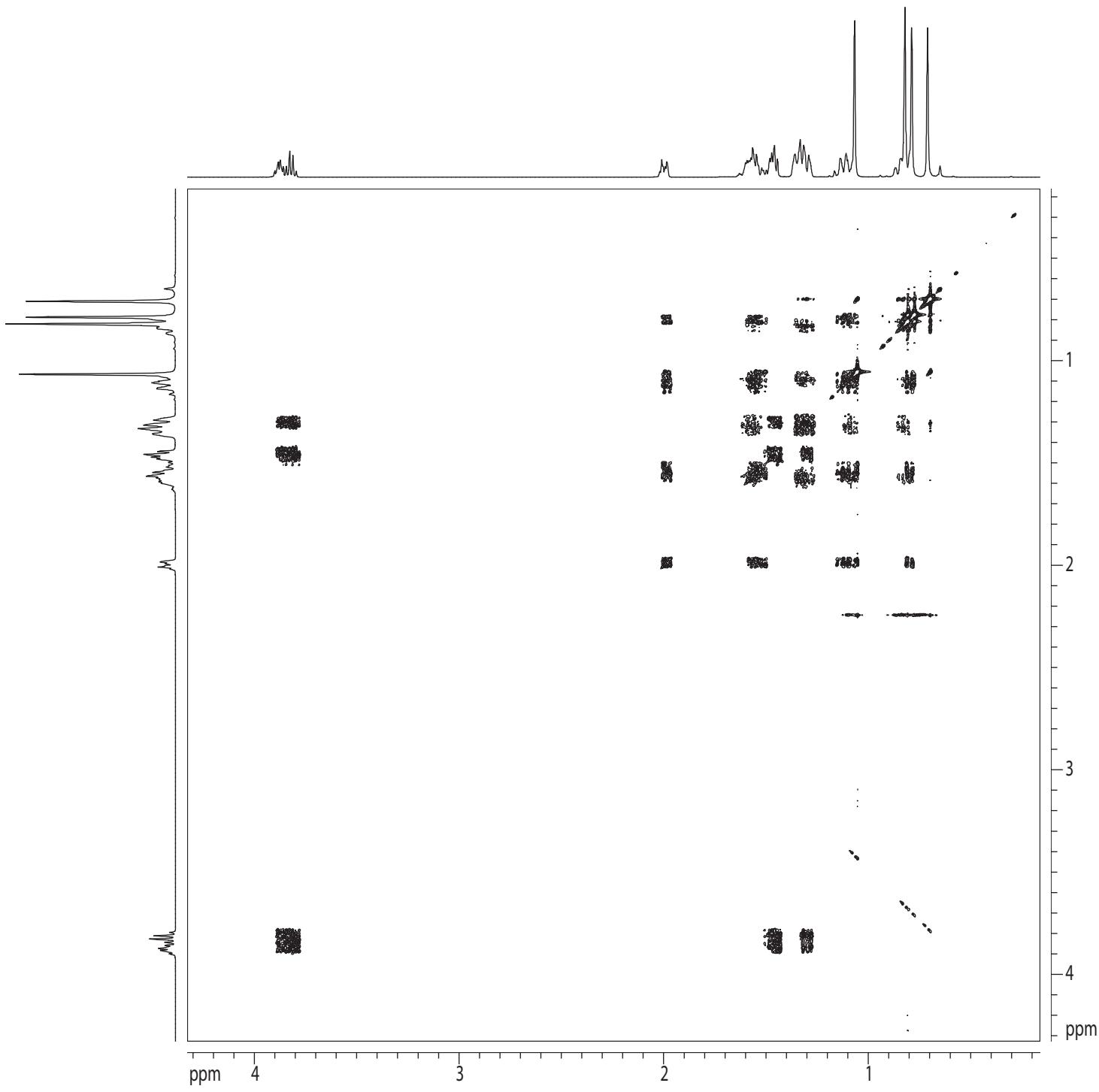
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F2HI 80.79 Hz  
F1PLO 4.326 ppm  
F1LO 2164.12 Hz  
F1PHI 0.162 ppm  
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F2PPMCM 0.27766 ppm/cm  
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F1PPMCM 0.27766 ppm/cm  
F1HZCM 138.88889 Hz/cm

gcosy60



gtoctsy



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D1 2.000000 sec  
D9 0.060000 sec  
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FACTOR1 4  
INO 0.00048000 sec  
I1 24  
SCALEF 6

===== CHANNEL f1 =====  
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p5 23.34 usec  
P6 35.00 usec  
p7 70.00 usec  
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GPZ2 10.00 %  
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F1 - Processing parameters  
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2D NMR plot parameters  
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CX1 15.00 cm  
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F2LO 2164.12 Hz  
F2PHI 0.162 ppm  
F2HI 80.79 Hz  
F1PLO 4.326 ppm  
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F1HI 80.79 Hz  
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d2 0.0344828 sec  
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d13 0.0000300 sec  
D16 0.0002000 sec  
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IN0 0.00004960 sec

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PL2 -1.00 dB  
PL12 11.30 dB  
SF02 125.7865620 MHz

===== GRADIENT CHANNEL =====  
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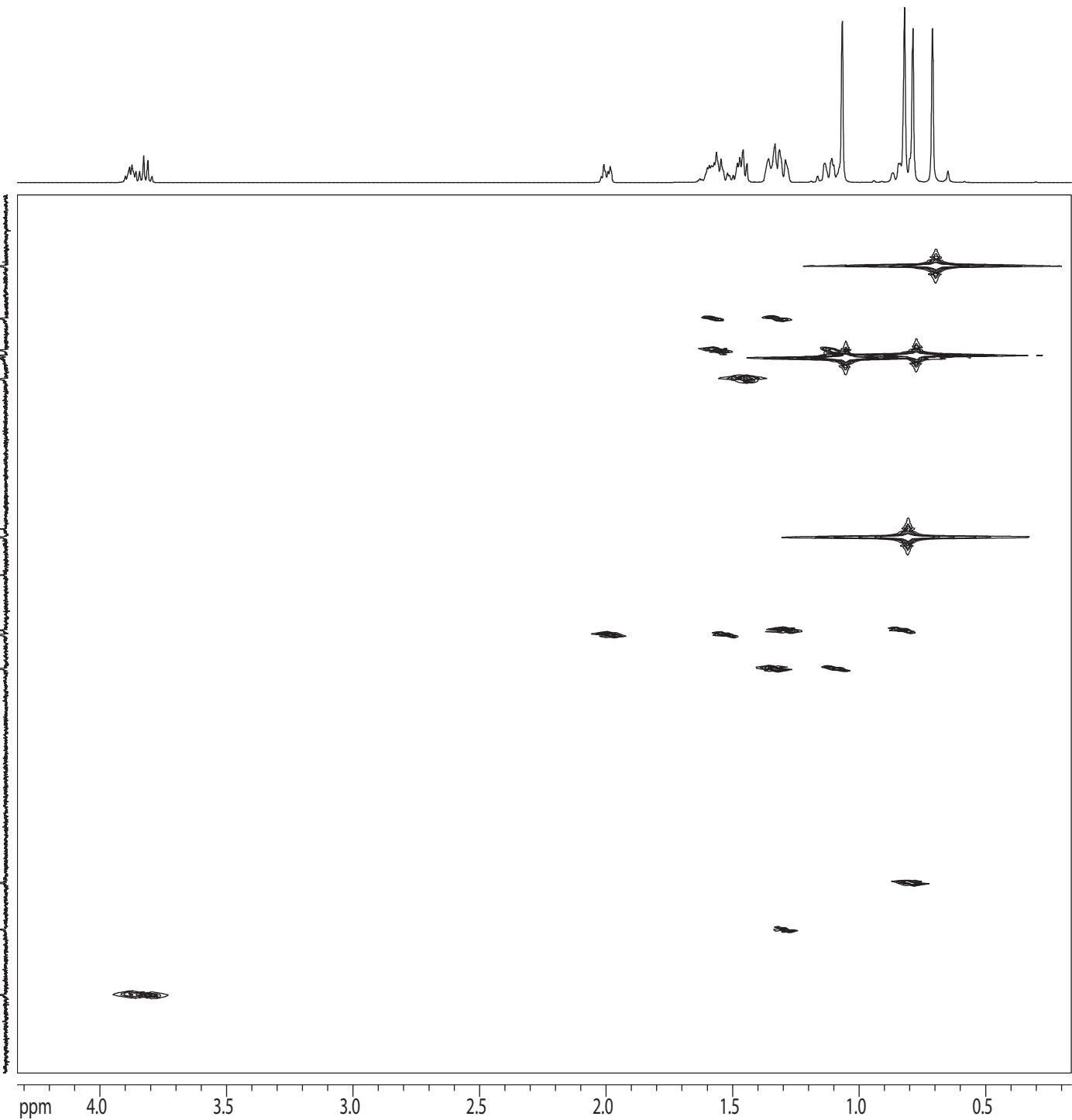
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SF01 125.7866 MHz  
FIDRES 19.688761 Hz  
SW 80.141 ppm  
FnMODE undefined

F2 - Processing parameters  
SI 1024  
SF 500.2200000 MHz  
WDW EM  
SSB 0  
LB 5.00 Hz  
GB 0  
PC 1.40

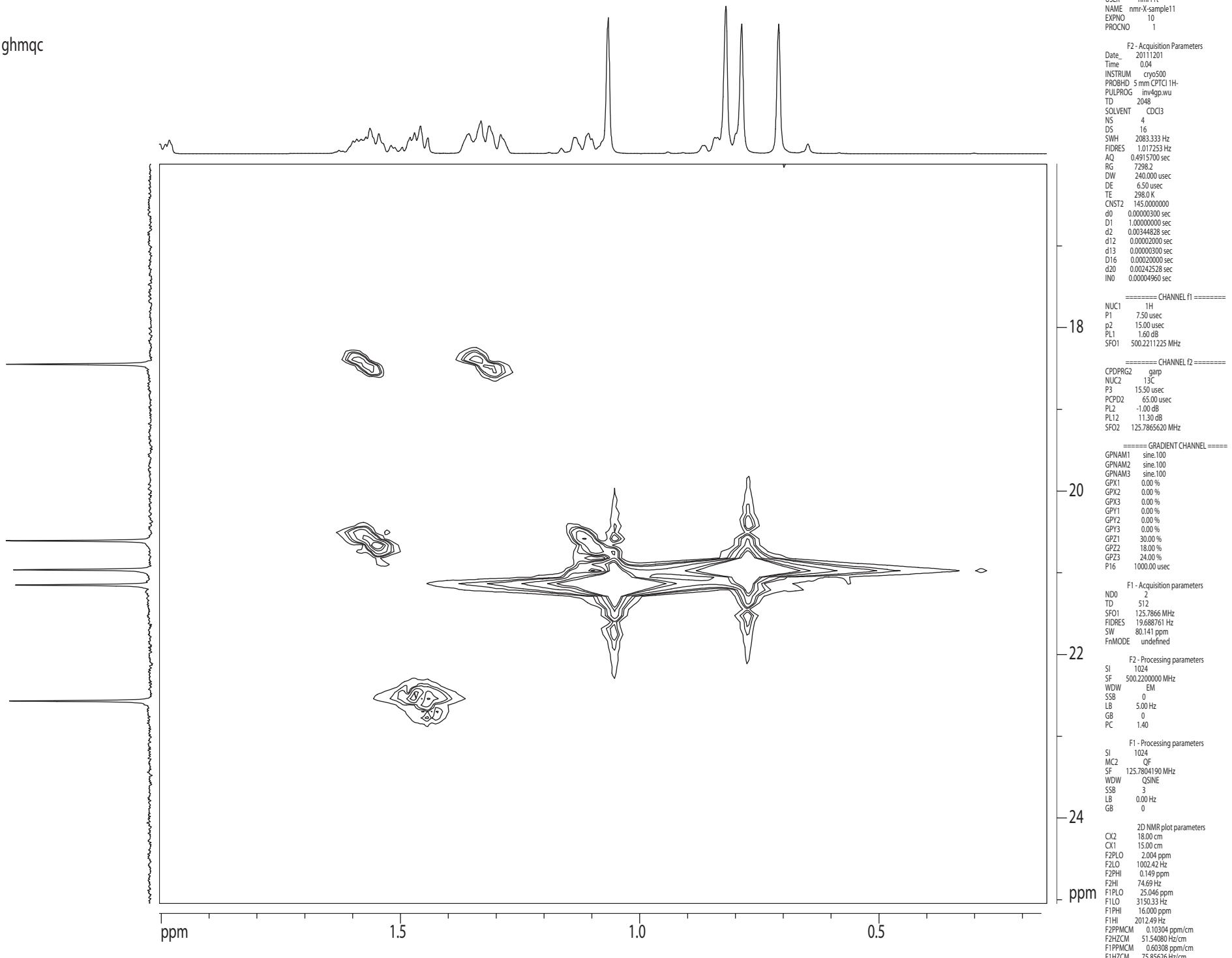
F1 - Processing parameters  
SI 1024  
MC2 QF  
SF 125.7804190 MHz  
WDW QSINE  
SSB 3  
LB 0.00 Hz  
GB 0

2D NMR plot parameters  
CX2 18.00 cm  
CX1 15.00 cm  
F2PLO 4.326 ppm  
F2LO 2164.12 Hz  
F2PHI 0.162 ppm  
F2HI 80.79 Hz  
F1PLO 70.000 ppm  
F1LO 880.63 Hz  
F1PHI 10.000 ppm  
F1HI 1257.80 Hz  
F2PPMCM 0.23138 ppm/cm  
F2H2CM 115.4074 Hz/cm  
F1PPMCM 4.00000 ppm/cm  
F1H2CM 503.12167 Hz/cm

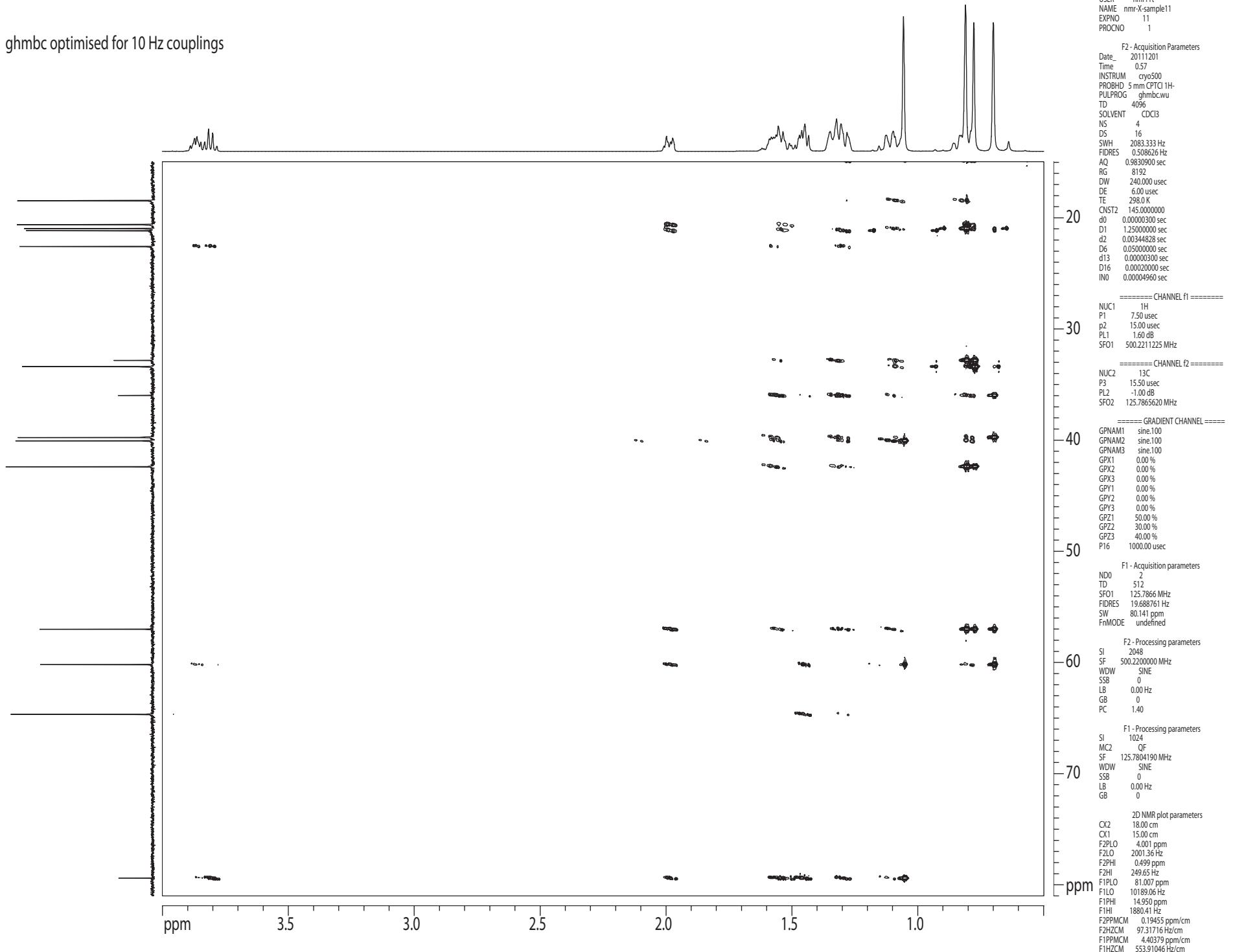
ghmqc



ghmqc



ghmbc optimised for 10 Hz couplings



Current Data Parameters  
USER nmr11t  
NAME nmr-X-sample11  
EXPNO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20111201  
Time 0.57  
INSTRUM cryo500  
PROBHD 5 mm CPTCI 1H-  
PULPROG ghmbc.wu  
TD 4096  
SOLVENT CDCl3  
NS 4  
DS 16  
SWH 2083.333 Hz  
FIDRES 0.508626 Hz  
AQ 0.983090 sec  
RG 8192  
DW 240.000 usec  
DE 6.00 usec  
TE 298.0 K  
CNST2 145.000000  
d0 0.0000300 sec  
D1 1.2500000 sec  
d2 0.0034482 sec  
D6 0.0500000 sec  
d13 0.0000300 sec  
D16 0.0002000 sec  
IN0 0.00004960 sec

===== CHANNEL f1 ======  
NUC1 1H  
P1 7.50 usec  
p2 15.00 usec  
PL1 1.60 dB  
SF01 500.2211225 MHz

===== CHANNEL f2 ======  
NUC2 13C  
P3 15.50 usec  
PL2 -1.00 dB  
SF02 125.7865620 MHz

===== GRADIENT CHANNEL =====  
GPNAME1 sine.100  
GPNAME2 sine.100  
GPNAME3 sine.100  
GPX1 0.00 %  
GPX2 0.00 %  
GPX3 0.00 %  
GPy1 0.00 %  
GPy2 0.00 %  
GPy3 0.00 %  
GPZ1 50.00 %  
GPZ2 30.00 %  
GPZ3 40.00 %  
P16 1000.00 usec

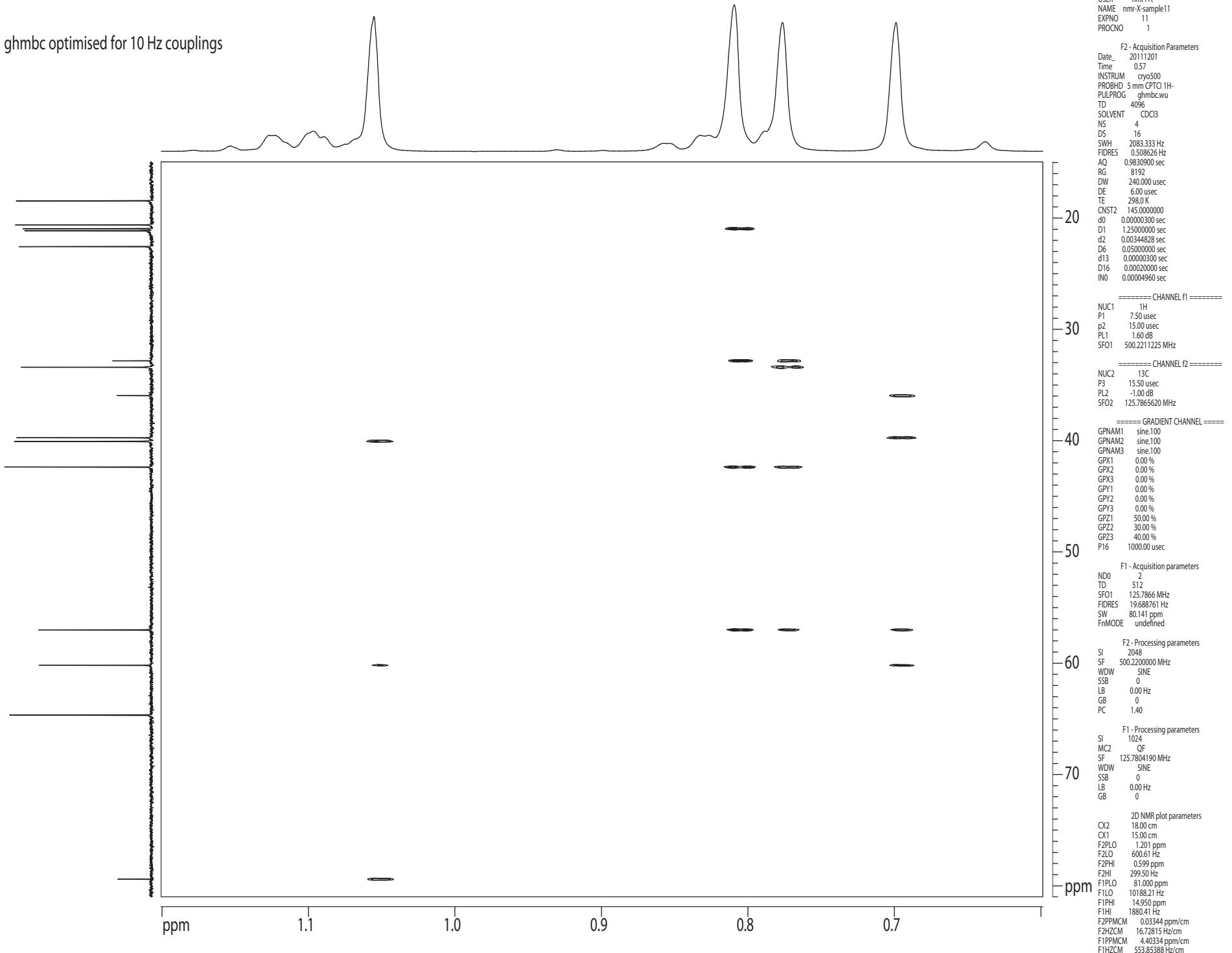
F1 - Acquisition parameters  
ND0 2  
TD 512  
SF01 125.7866 MHz  
FIDRES 19.688761 Hz  
SW 80.141 ppm  
FnMODE undefined

F2 - Processing parameters  
SI 2048  
SF 500.2200000 MHz  
WDW SINE  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.40

F1 - Processing parameters  
SI 1024  
MC2 QF  
SF 125.7804190 MHz  
WDW SINE  
SSB 0  
LB 0.00 Hz  
GB 0

2D NMR plot parameters  
CX2 18.00 cm  
CX1 15.00 cm  
F2PLO 4.001 ppm  
F2LO 2001.36 Hz  
F2PHI 0.499 ppm  
F2HI 249.65 Hz  
F1PLO 81.007 ppm  
F1LO 10189.06 Hz  
F1PHI 14.950 ppm  
F1HI 1880.41 Hz  
F2PPMCM 0.19455 ppm/cm  
F2HZCM 97.31716 Hz/cm  
F1PPMCM 4.40379 ppm/cm  
F1HZCM 553.91046 Hz/cm

ghmbc optimised for 10 Hz couplings



Current Data Parameters  
USER nmr11t  
NAME nmr-X-sample11  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20111201  
Time 2.17  
INSTRUM cryo500  
PROBHD 5 mm CPTCI 1H-  
PULPROG noesygptp  
TD 2048  
SOLVENT CDCl3  
NS 4  
DS 16  
SWH 2083.333 Hz  
FIDRES 1.017253 Hz  
AQ 0.4915700 sec  
RG 25.4  
DW 240.000 usec  
DE 6.00 usec  
TE 298.0 K  
D0 0.00000300 sec  
D1 2.0000000 sec  
D8 0.8000001 sec  
D16 0.0020000 sec  
d20 0.3988002 sec  
IN0 0.00024000 sec

===== CHANNEL f1 =====  
NUC1 1H  
P1 7.50 usec  
P2 15.00 usec  
PL1 1.60 dB  
SF01 500.2211225 MHz

===== GRADIENT CHANNEL =====  
GPNAME1 sine.100  
GPNAME2 sine.100  
GPX1 0.00 %  
GPX2 0.00 %  
GPY1 0.00 %  
GPY2 0.00 %  
GPZ1 40.00 %  
GPZ2 -40.00 %  
P16 1000.00 usec

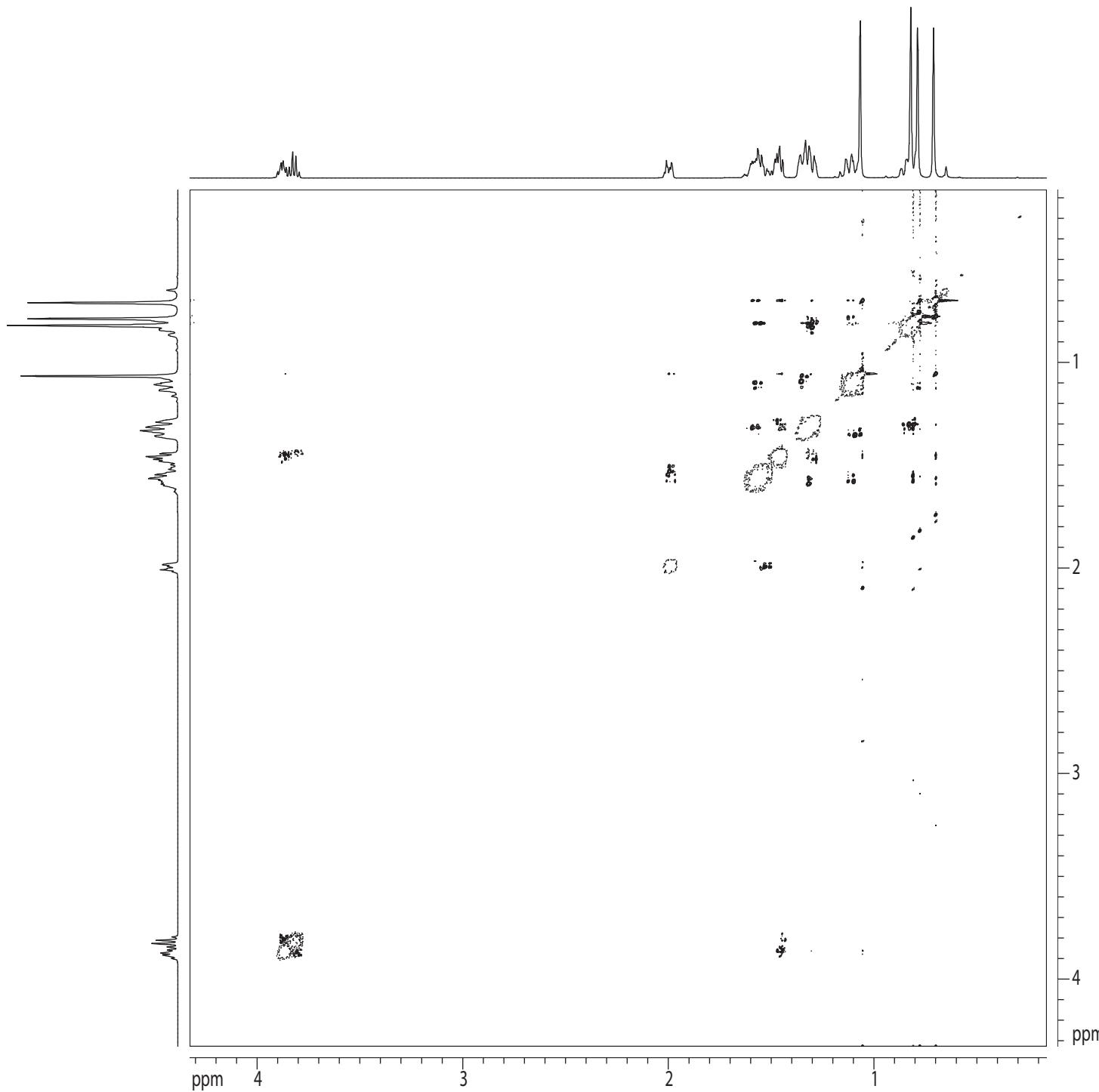
F1 - Acquisition parameters  
ND0 2  
TD 512  
SF01 500.2211 MHz  
FIDRES 4.069010 Hz  
SW 4.165 ppm  
FnMODE undefined

F2 - Processing parameters  
SI 1024  
SF 500.2200000 MHz  
WDW QSINE  
SSB 2  
LB 0.00 Hz  
GB 0  
PC 1.40

F1 - Processing parameters  
SI 1024  
MC2 TPII  
SF 500.2200000 MHz  
WDW QSINE  
SSB 2  
LB 0.00 Hz  
GB 0

2D NMR plot parameters  
CX2 15.00 cm  
CX1 15.00 cm  
F2PLO 4.326 ppm  
F2PHI 0.162 ppm  
F2HI 80.79 Hz  
F1PLO 4.326 ppm  
F1LO 2164.12 Hz  
F1PHI 0.162 ppm  
F1HI 80.79 Hz  
F2PPCM 0.27766 ppm/cm  
F2HZCM 138.88889 Hz/cm  
F1PPCM 0.27766 ppm/cm  
F1HZCM 138.88889 Hz/cm

gnnoesy



Current Data Parameters  
 USER nmr1lt  
 NAME nmr-X-sample11  
 EXPNO 12  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20111201  
 Time 2.17  
 INSTRUM cryo500  
 PROBHD 5 mm CPTCI 1H-  
 PULPROG noeipygtp  
 TD 2048  
 SOLVENT CDCl3  
 NS 4  
 DS 16  
 SWH 2083.333 Hz  
 FIDRES 1.017253 Hz  
 AQ 0.4915700 sec  
 RG 25.4  
 DW 240.000 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D0 0.0000300 sec  
 D1 2.000000 sec  
 D8 0.8000001 sec  
 D16 0.0002000 sec  
 d20 0.3988002 sec  
 IN0 0.0002400 sec

===== CHANNEL f1 ======  
 NUC1 1H  
 P1 7.50 usec  
 P2 15.00 usec  
 PL1 1.60 dB  
 SFO1 500.2211225 MHz

===== GRADIENT CHANNEL =====  
 GPNAM1 sine.100  
 GPNAM2 sine.100  
 GPX1 0.00 %  
 GPX2 0.00 %  
 GPY1 0.00 %  
 GPY2 0.00 %  
 GPZ1 40.00 %  
 GPZ2 -40.00 %  
 P16 1000.00 usec

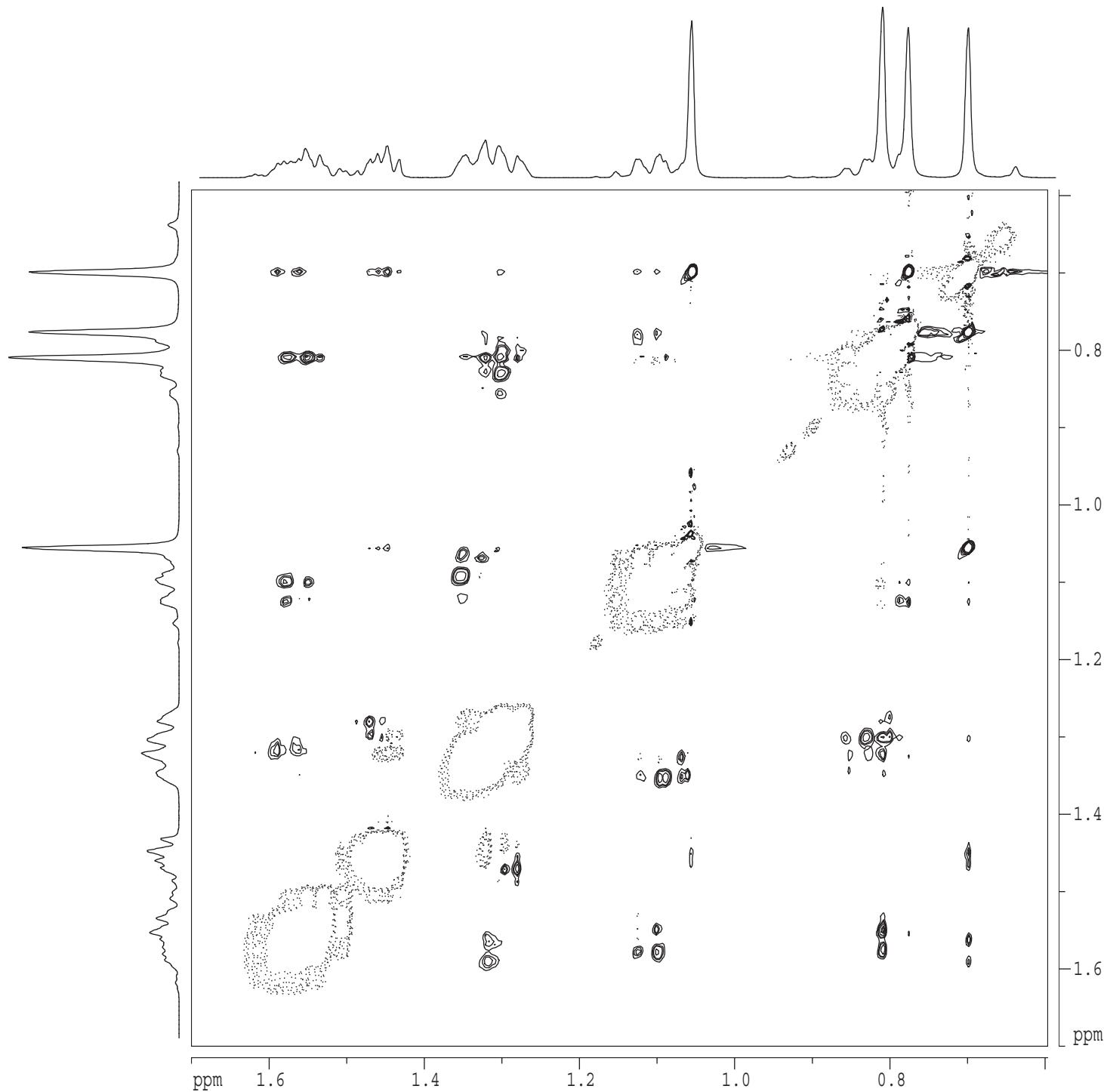
F1 - Acquisition parameters  
 ND0 2  
 TD 512  
 SFO1 500.2211 MHz  
 FIDRES 4.069010 Hz  
 SW 4.165 ppm  
 FnMODE undefined

F2 - Processing parameters  
 SI 1024  
 SF 500.2200000 MHz  
 WDW QSINE  
 SSB 2  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

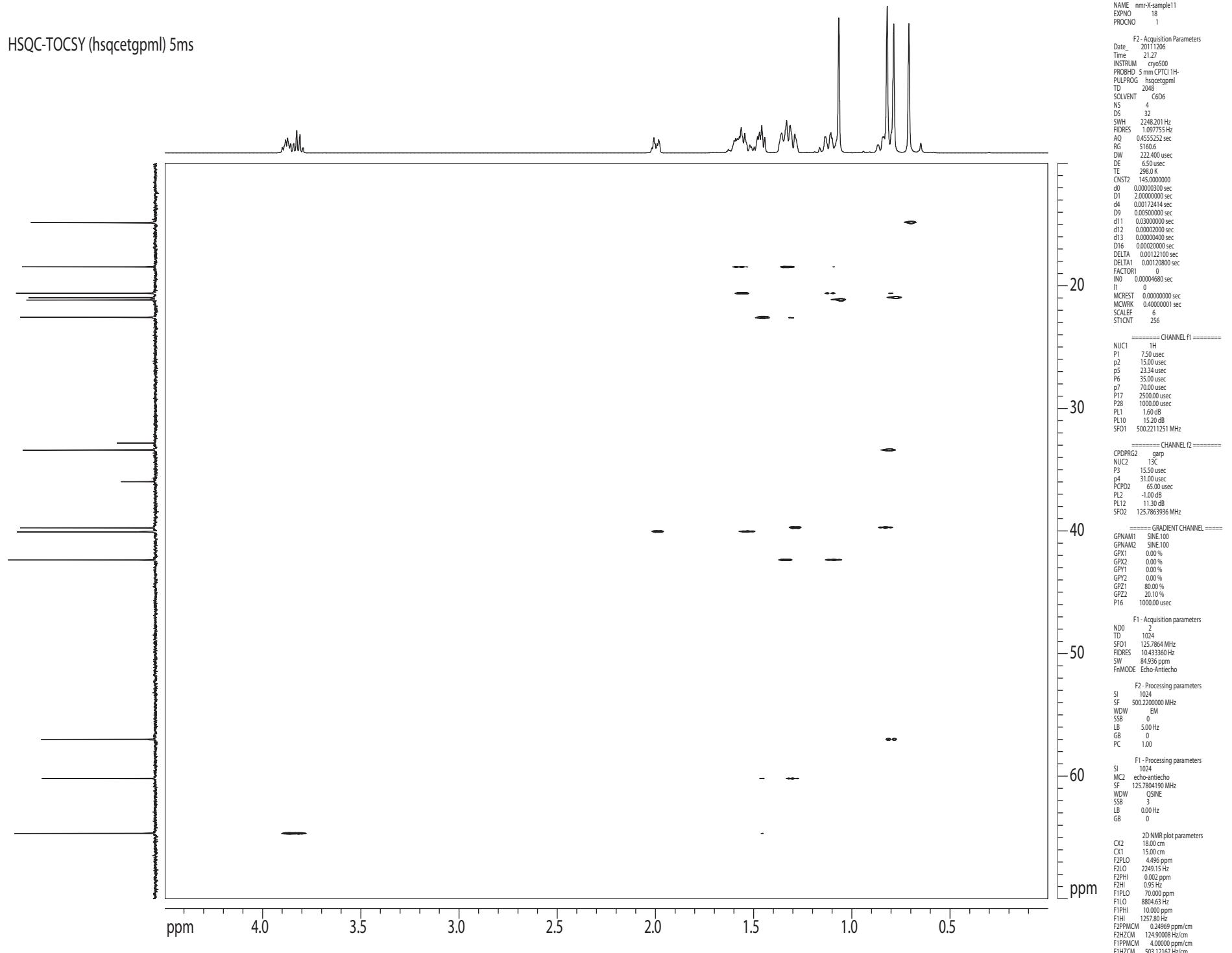
F1 - Processing parameters  
 SI 1024  
 MC2 TPPI  
 SF 500.2200000 MHz  
 WDW QSINE  
 SSB 2  
 LB 0.00 Hz  
 GB 0

2D NMR plot parameters  
 CX2 15.00 cm  
 CX1 15.00 cm  
 F2PLO 1.700 ppm  
 F2LO 850.37 Hz  
 F2PHI 0.597 ppm  
 F2HI 298.48 Hz  
 F1PLO 1.700 ppm  
 F1LO 850.37 Hz  
 F1PHI 0.593 ppm  
 F1HI 296.45 Hz  
 F2PPMCM 0.07355 ppm/cm  
 F2HZCM 36.79276 Hz/cm  
 F1PPMCM 0.07382 ppm/cm  
 F1HZCM 36.92840 Hz/cm

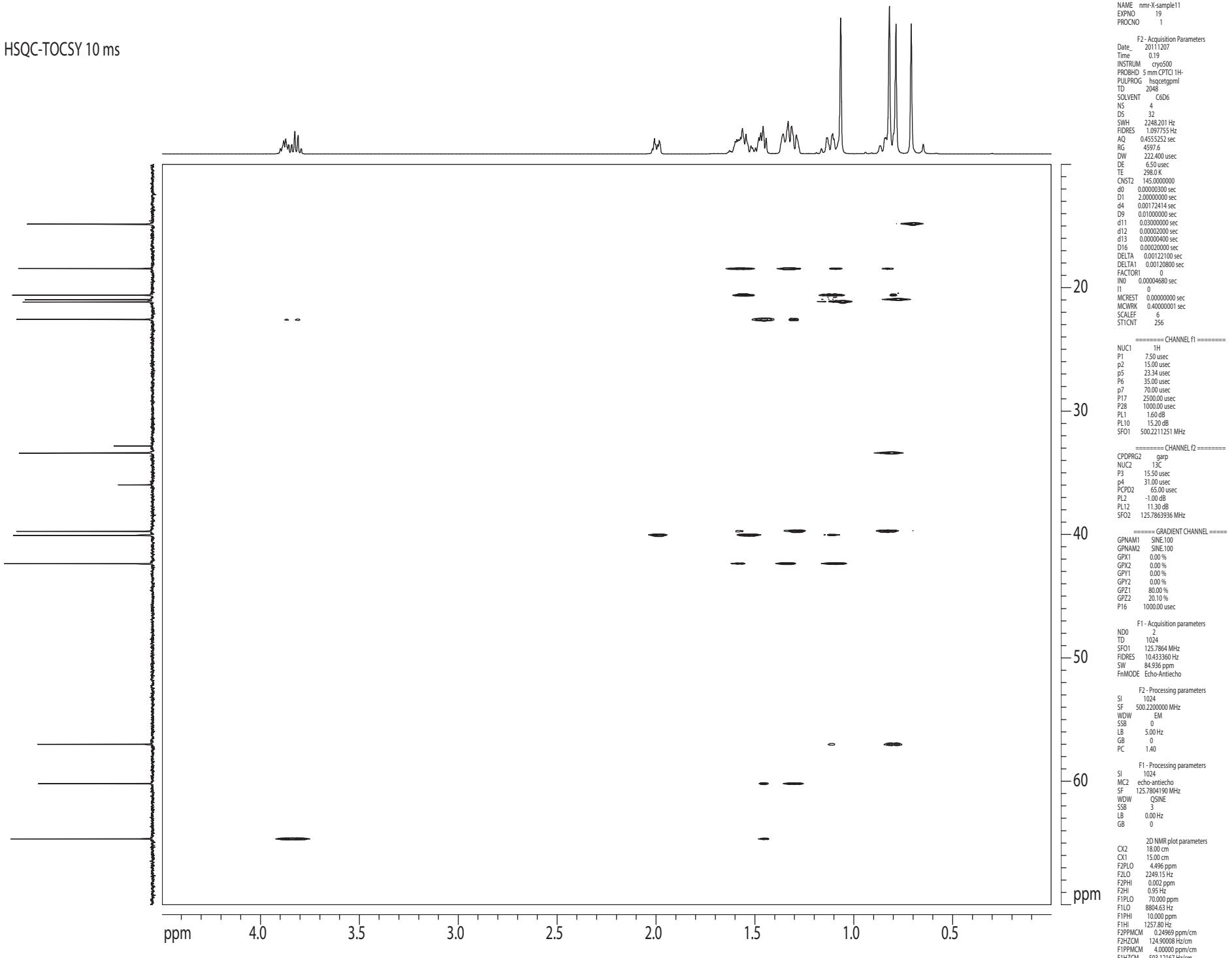
gnoesy



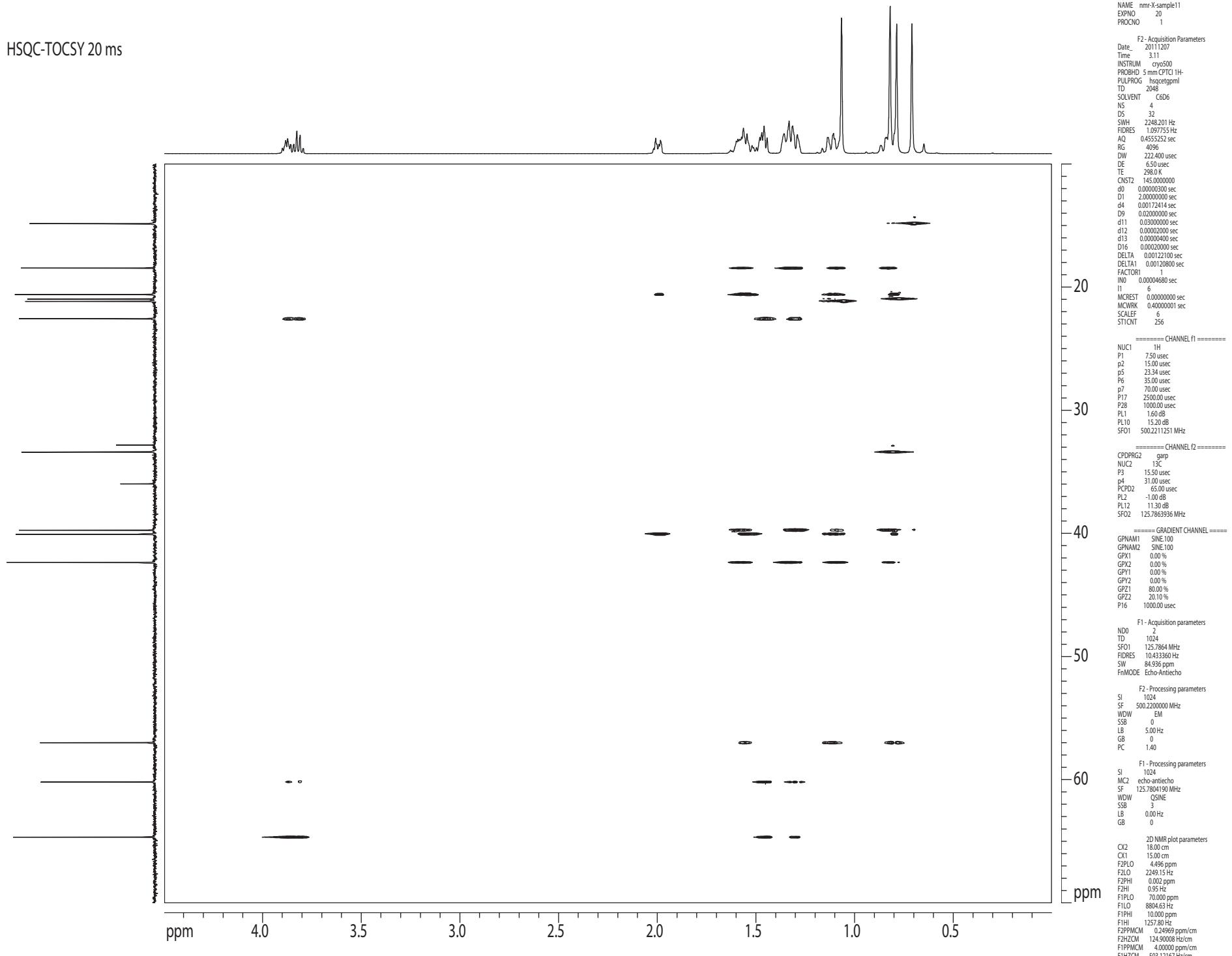
HSQC-TOCSY (hsqcetgpm) 5ms



## HSQC-TOCSY 10 ms



## HSQC-TOCSY 20 ms



## HSQC-TOCSY 100 ms

