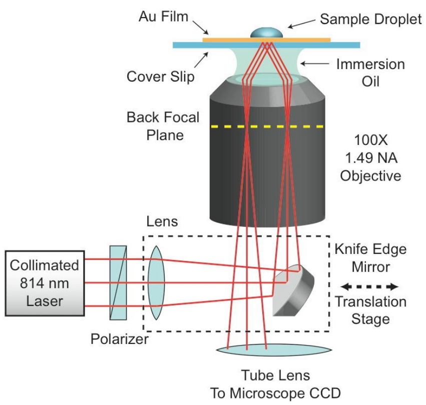
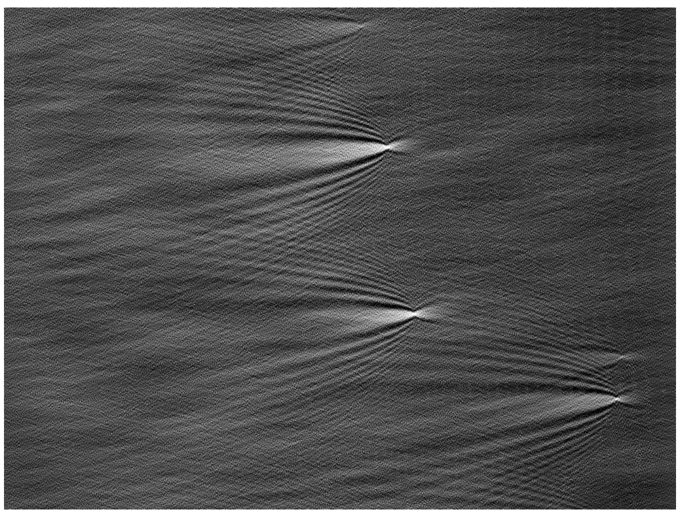
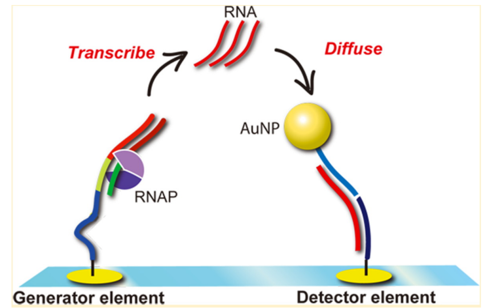
Single Nanoparticle Biosensing with Surface Plasmon Resonance Imaging

Single Nanoparticle SPRI

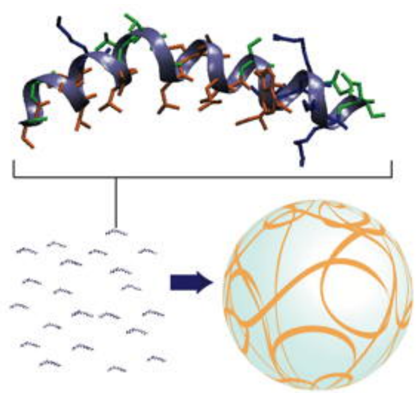


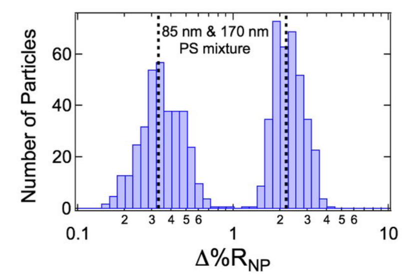


Surface Enzyme Chemistries

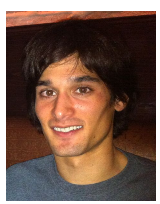
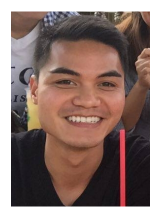
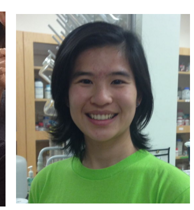
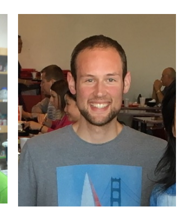


Bioaffinity Uptake into Single Nanoparticles





Robert M. Corn
UCI Department of Chemistry

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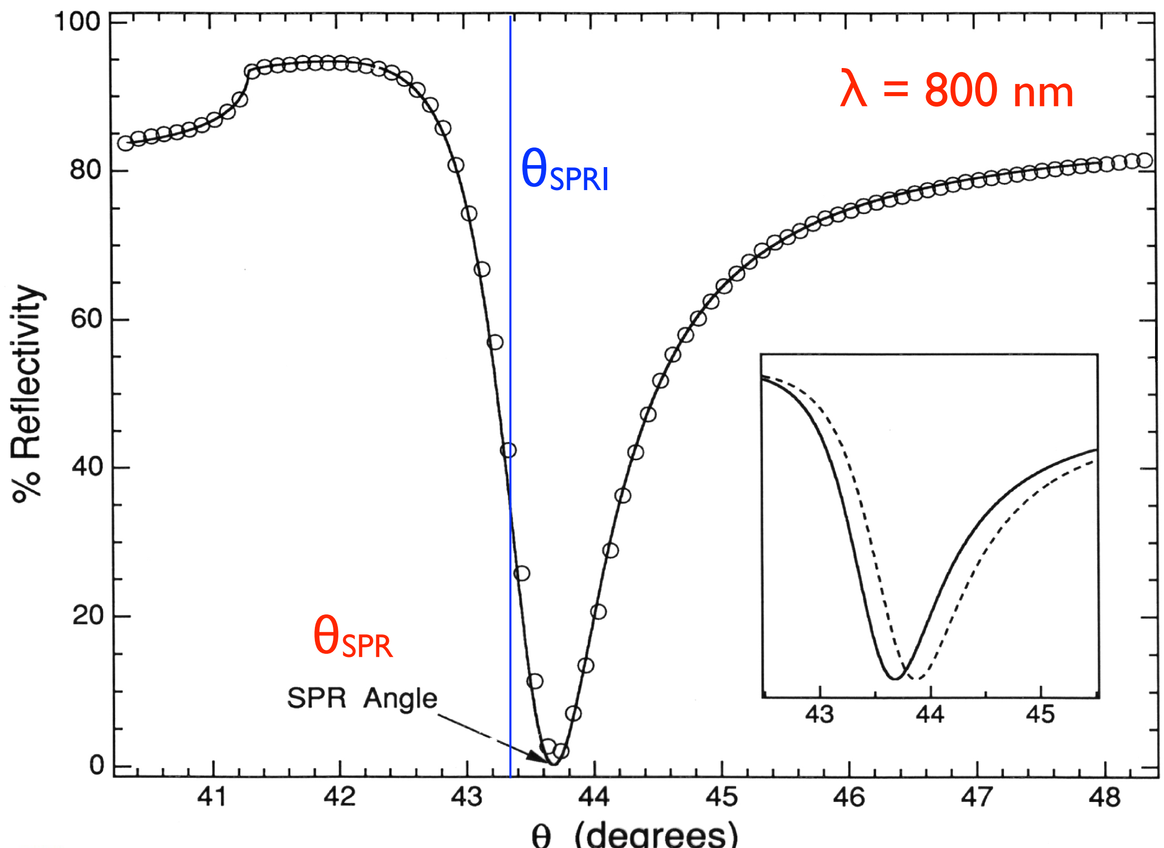
A quick note on Surface Plasmon Polaritons and SPRI

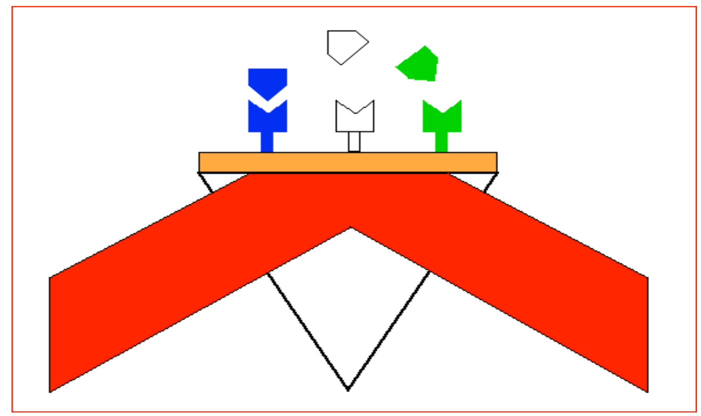
Surface plasmon polaritons (SPPs) are localized propagating electromagnetic waves that can be created on a thin gold film (45 nm) attached to a prism surface.

Surface Plasmon Resonance (SPR)

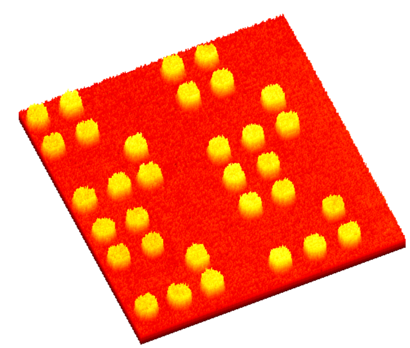
Changes in the resonant SPR angle (θ_{SPR}) can be used to study adsorption onto the gold surface.



NIR SPR Imaging (SPRI)



SPR imaging (SPRI) measures the change in reflectivity from a surface at the θ_{SPRI} upon adsorption.



JM Brockman, AG Frutos, BP Nelson and RM Corn, Analytical Chem., 71 3928-3934 (1999).

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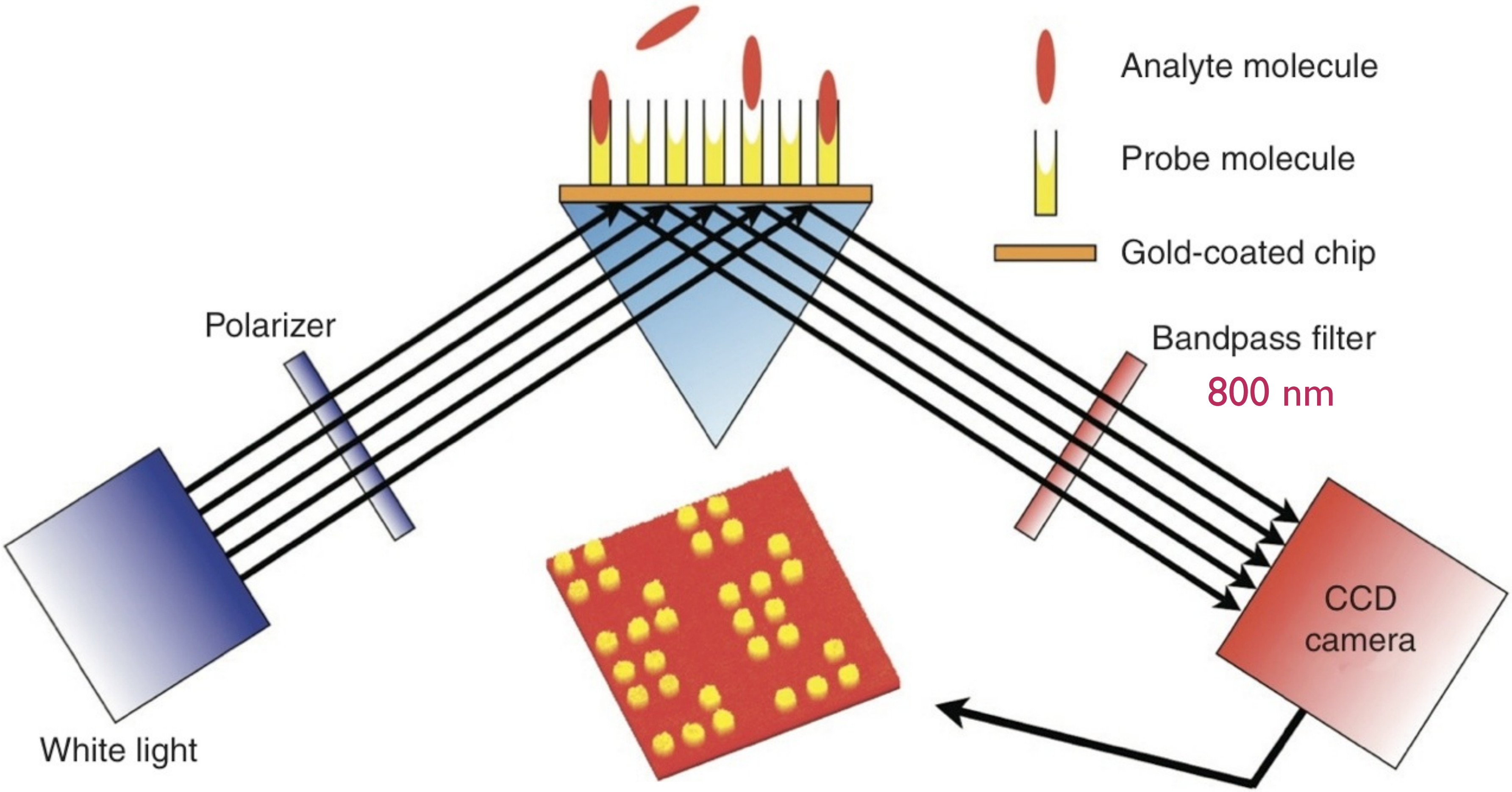
NIR Surface Plasmon Resonance Imaging (SPRI)

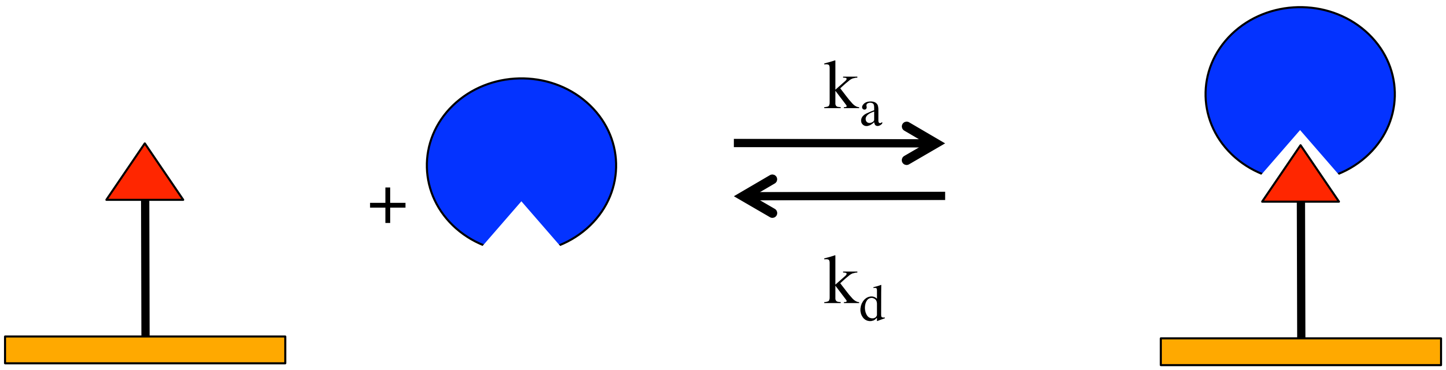


DNA microarray

Multiplexed Refractive Index Surface Bioaffinity Sensing

*B. Rothenäusler and W. Knoll, Nature **332**, 615-617 (1988).*
JM Brockman, AG Frutos, BP Nelson and RM Corn, Analytical Chem., 71 3928-3934 (1999).

Langmuir Surface Adsorption Kinetics

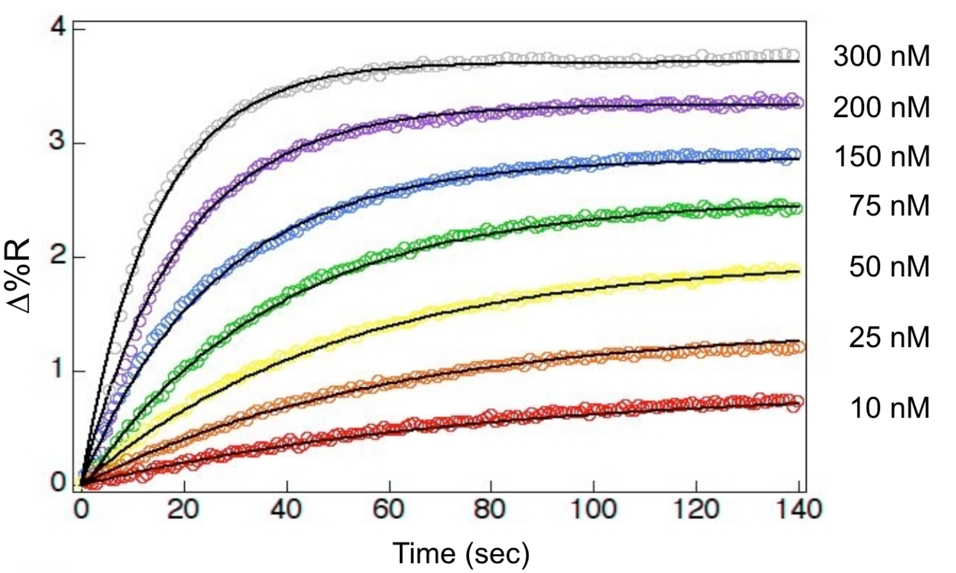


$$K_{ads} = \frac{k_a}{k_d}$$

$$\theta = \Gamma / \Gamma_{tot}$$

S protein - S peptide

$$\theta(t) = \theta^{eq}\left(1 - \exp(-(k_a C + k_d)t)\right)$$



$k_a = 1.9 \times 10^5\ M^{-1}\ s^{-1}$

$k_d = 1.1 \times 10^{-2}\ s^{-1}$

$K_{ads} = 1.7 \times 10^7\ M^{-1}$

Greta Wegner

G. J. Wegner et al. Analytical Chem., 76 5667-5684 (2004).

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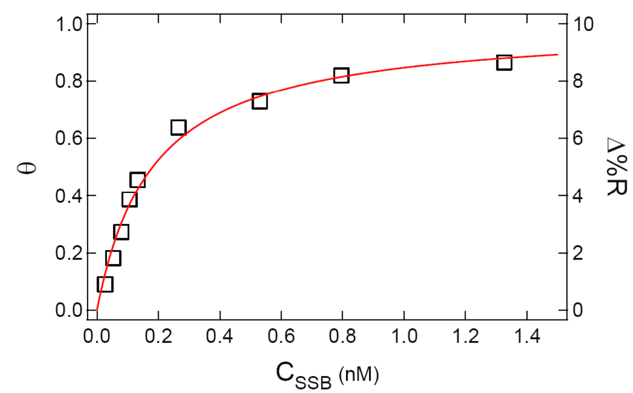
Langmuir Surface Adsorption Numbers: Femtomolar Detection

Stoichiometry: 1fM x 10μL = 10^{-20}moles = 6000 molecules

$$K_{ads} = \frac{k_a}{k_d} = \frac{\theta^{eq}}{C(1-\theta^{eq})}$$



Dr. Yulin Chen

$$\theta^{eq} = \Gamma/_{\Gamma^{tot}} \approx K_{ads}C = (10^9 M^{-1})(10^{-15} M) = 10^{-6}$$

$$\Gamma^{tot} \approx 10^{12} molecules\ cm^{-2}$$

A = 100 μm x 100 μm

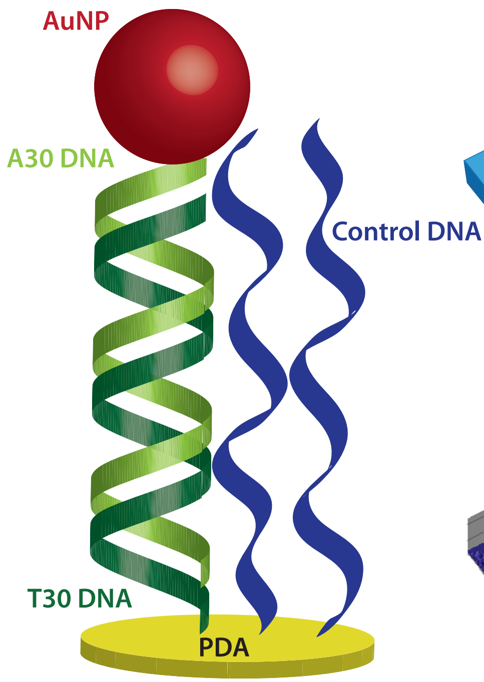
$$\Gamma A \approx (10^6 molecules\ cm^{-2})(10^{-4} cm^2) = 100\ molecules$$

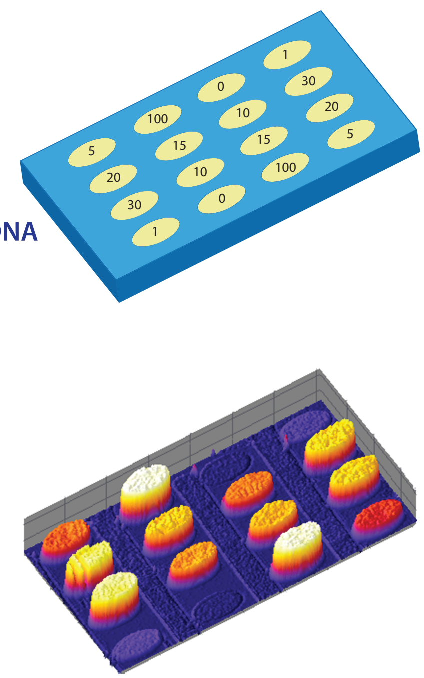
Y. Chen et al., Langmuir, ***25*** *5054-5060 (2009).*

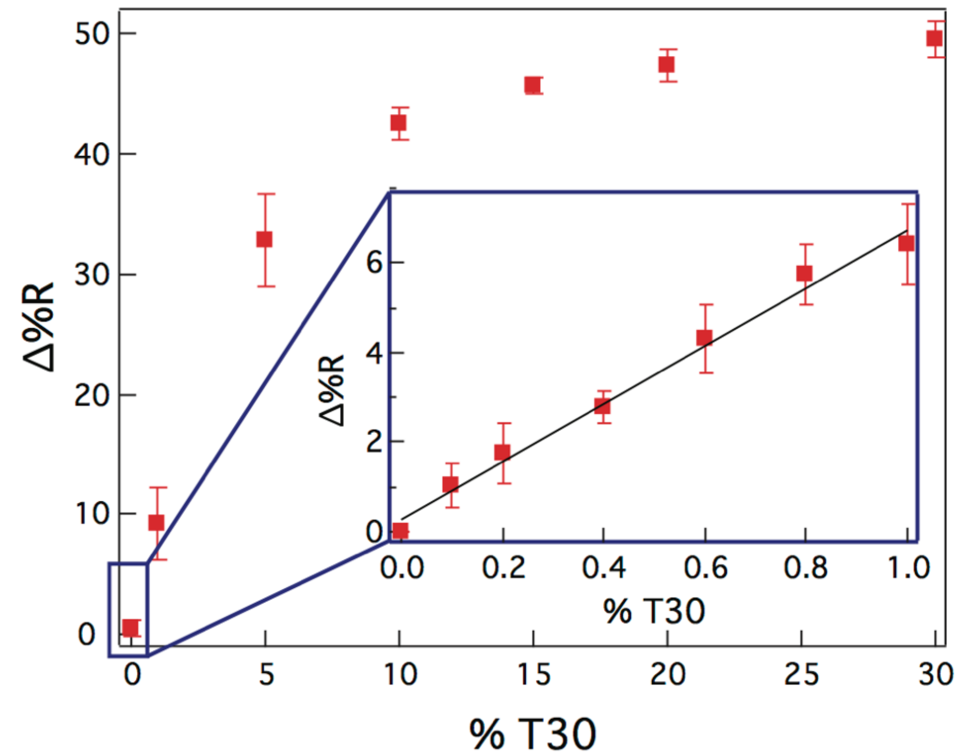
Nanoparticle-Enhanced SPRI Detection of Surface DNA

polydopamine + amino-functionalized ssDNA

$\Gamma_{max} = 10^{12}$ *molecules* cm^{-2}







0.1% ssDNA Detection in mixed ssDNA monolayers

Jennifer Wood Fasoli

J.Wood et al., Langmuir, ***29*** *10868-10873 (2013).*

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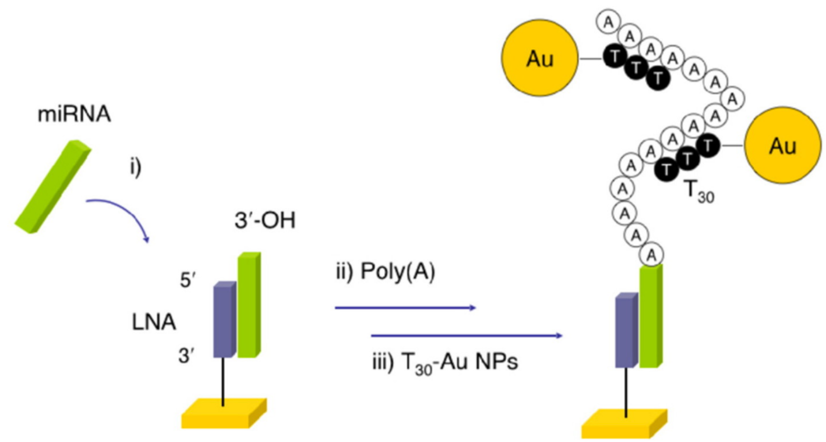
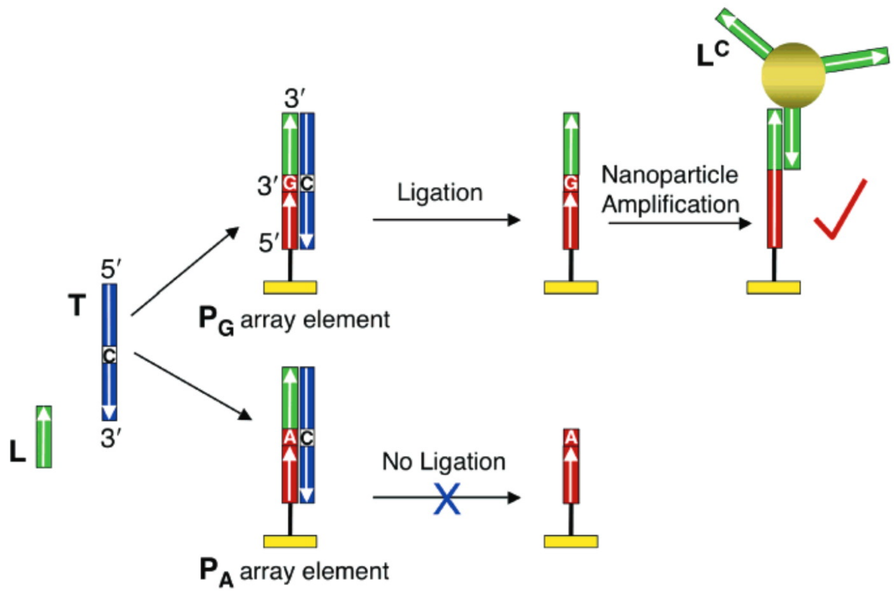
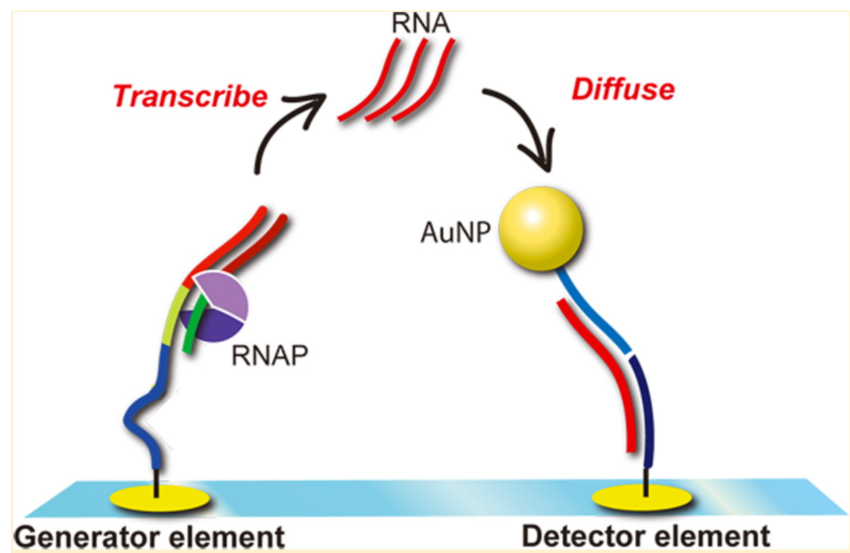
SPRI Measurements of Surface Enzyme Chemistries

- Surface Ligation Chemistry
- Surface Nuclease Chemistry
- Surface Polymerase Chemistry
- Surface DNAzyme Chemistry







Surface Enzyme Chemistry + Nanoparticle-Enhanced SPRI
Enhanced Detection

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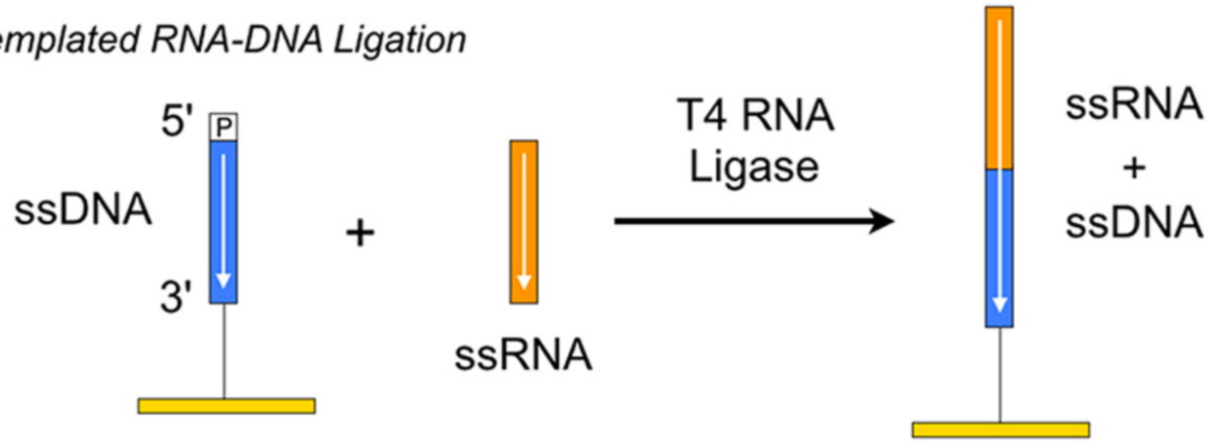
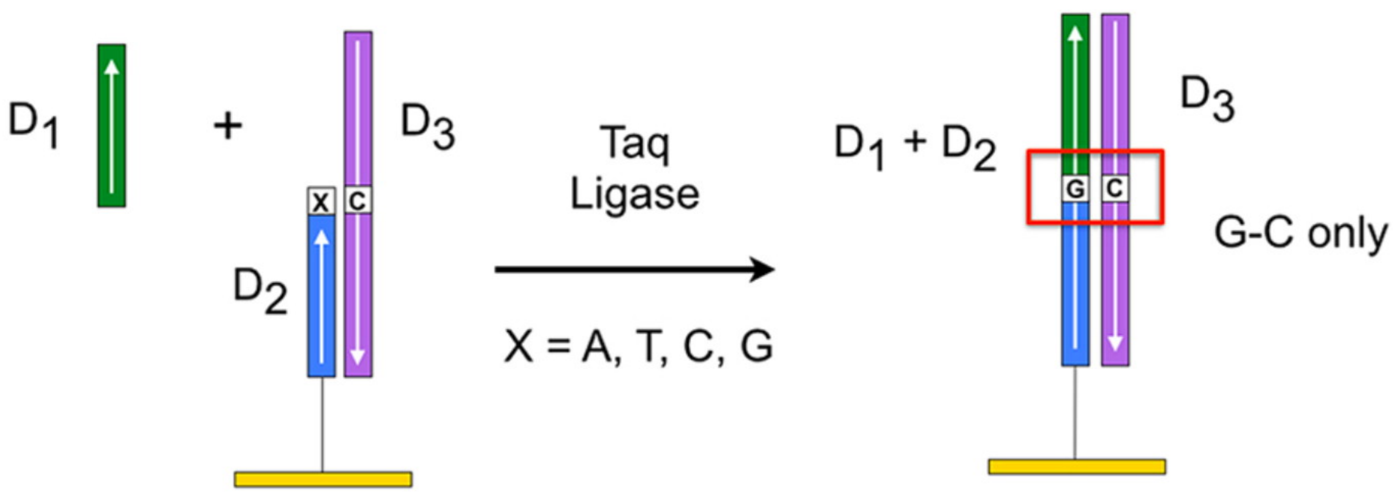
Surface Ligation Chemistries

a) *Untemplated RNA-DNA Ligation*



microRNA detection

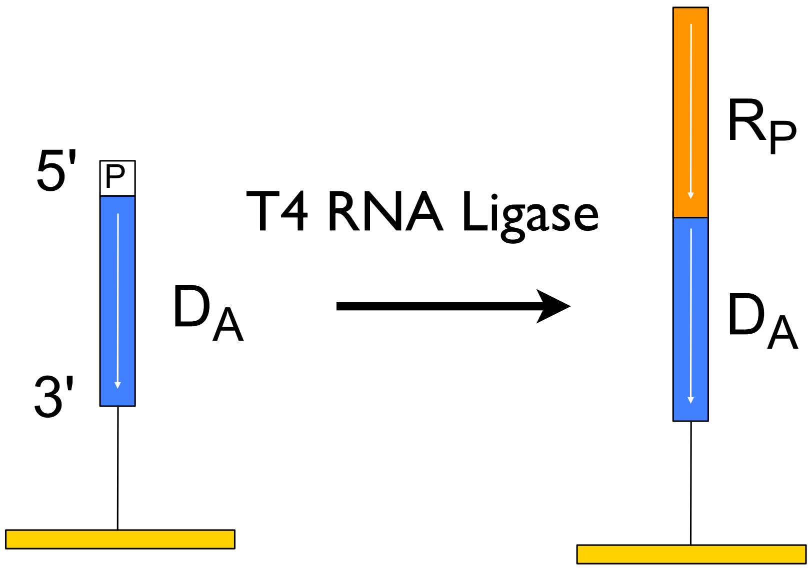
b) *Templated DNA-DNA Ligation*



SNP detection

Surface Attachment Chemistry: Enzymatic Ligation of ssRNA to ssDNA

T4 RNA Ligase: No template required.
Previously used to make RNA Aptamer arrays:



Ting Nico Hu Seefeld

Surface Enzyme Chemistries:
Attachment, Amplification, Selectivity

T. H. Seefeld et al., Langmuir, **27** 6534-6540 (2011).

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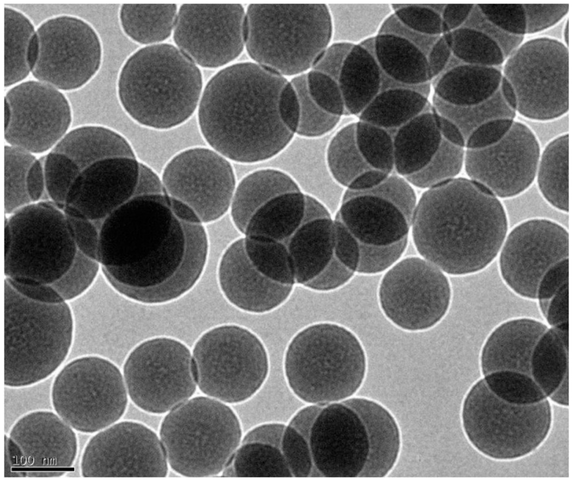
DNA-Functionalized Nanoparticles

100 nm silica nanoparticles (SiNPs)



TEM

DNA-functionalized:

- *Silica Nanoparticles*
- *Gold Nanoparticles*
- *Polystyrene Nanoparticles*
- *Magnetite Nanoparticles*

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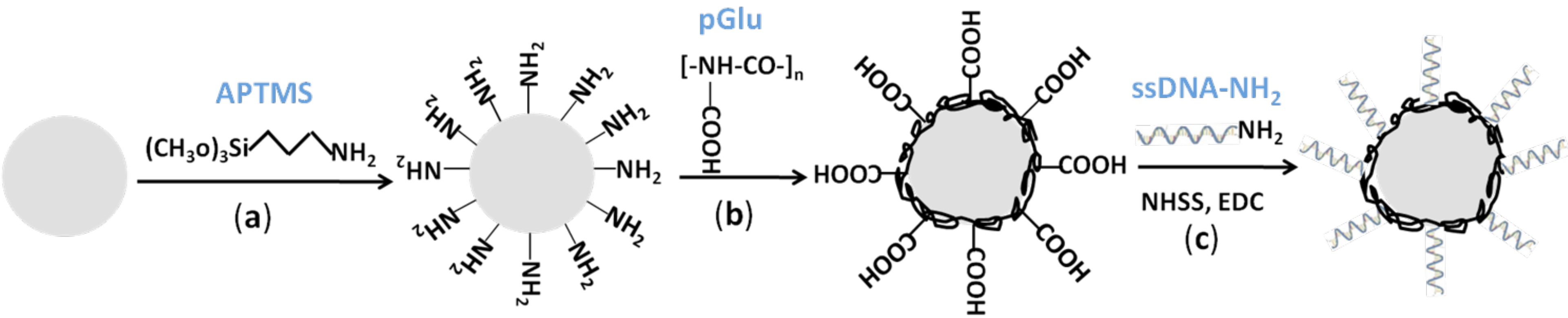
pGlu DNA Attachment Chemistry on Silica Nanoparticles (SiNPs)



W. J. Zhou et al., *Analytical Chemistry*, **83** 3897-3902 (2011).

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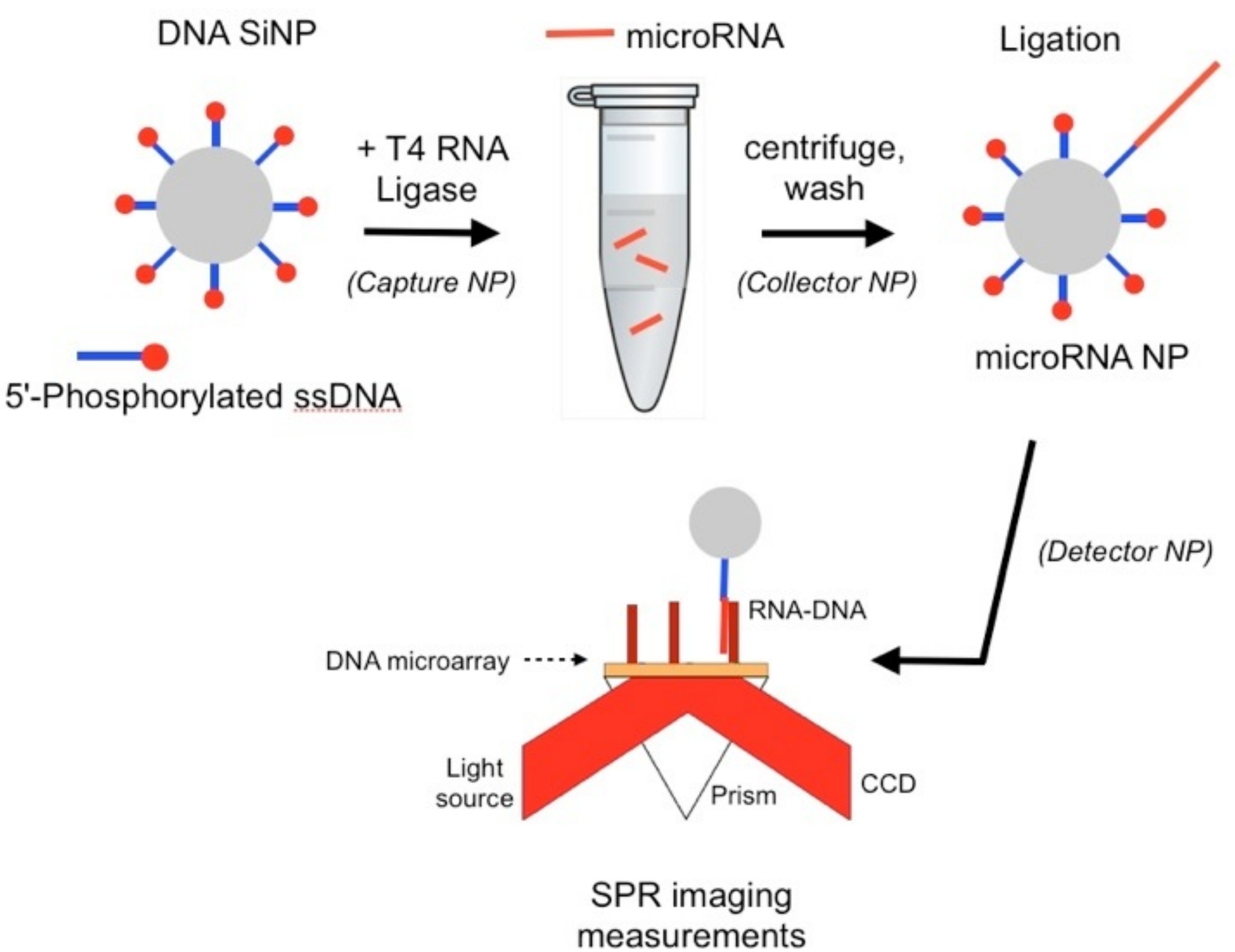
Surface Enzyme Chemistry + Nanoparticle-Enhanced SPRI

- *MicroRNA Detection with Enzymatic Nanoparticles: Ligation Capture*



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Dr. WenJuan Zhou

T4 RNA Ligation for miRNA capture

W. J. Zhou et al., Analytical Chemistry, ***83*** *3897-3902 (2011).*

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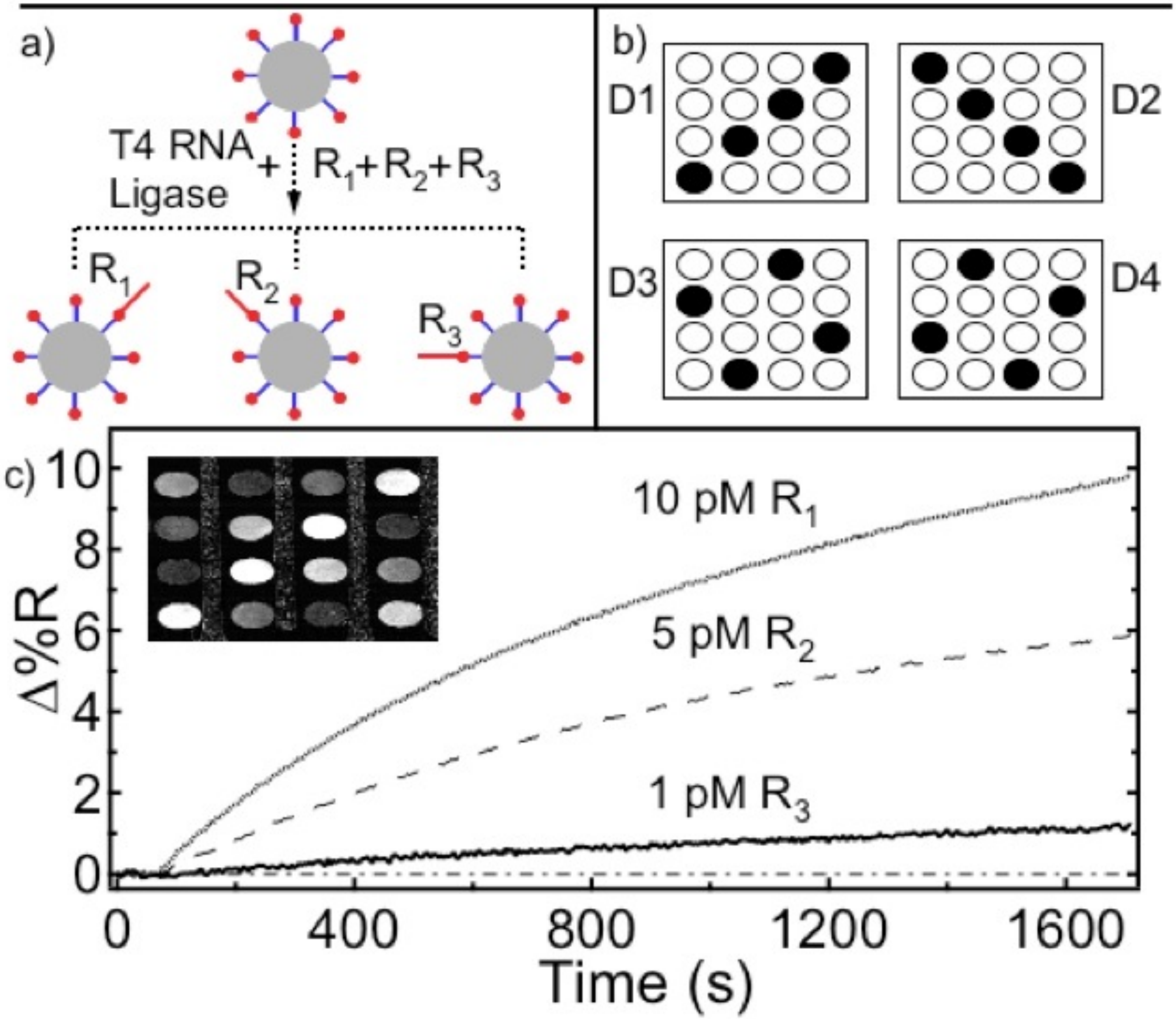
MicroRNA Detection with Enzymatic Ligation Capture Nanoparticles



Multiplex MicroRNA Detection!

*W. J. Zhou et al., Analytical Chemistry, **83** 3897-3902 (2011).*

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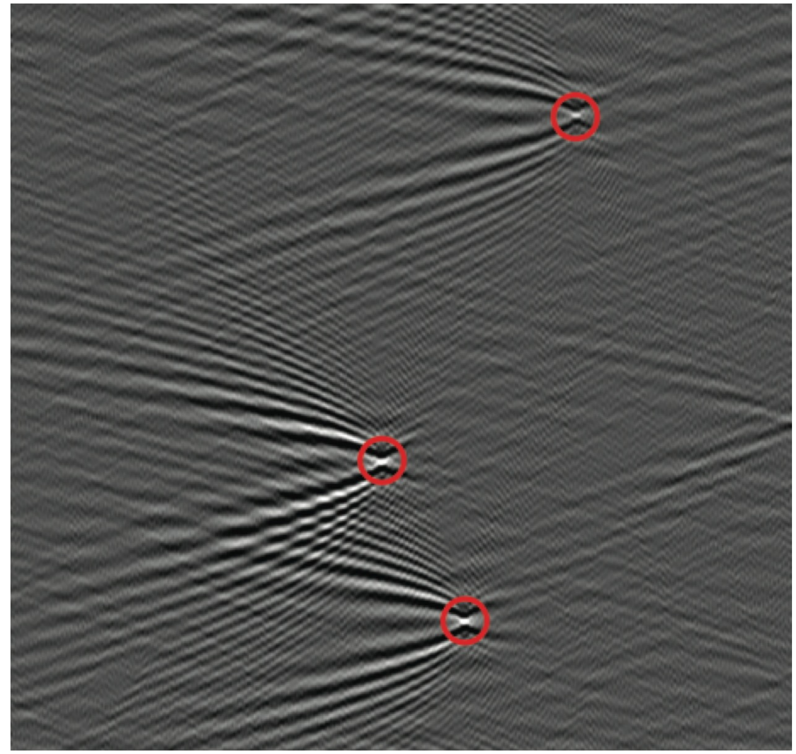
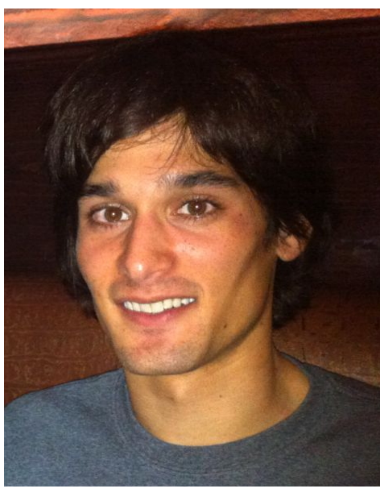
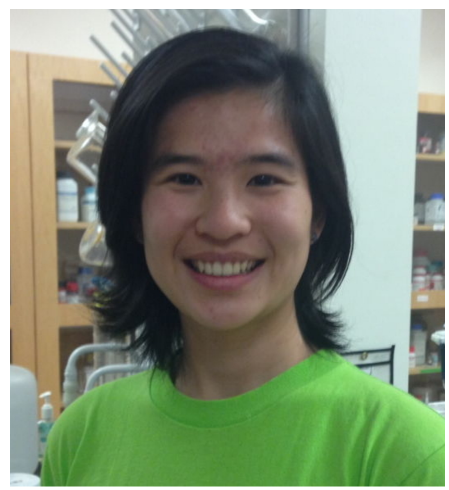
Single Nanoparticle SPRI Microscopy





Dr. Aaron Halpern

Adam Maley



Millie Fung

Robert M. Corn
UC Irvine

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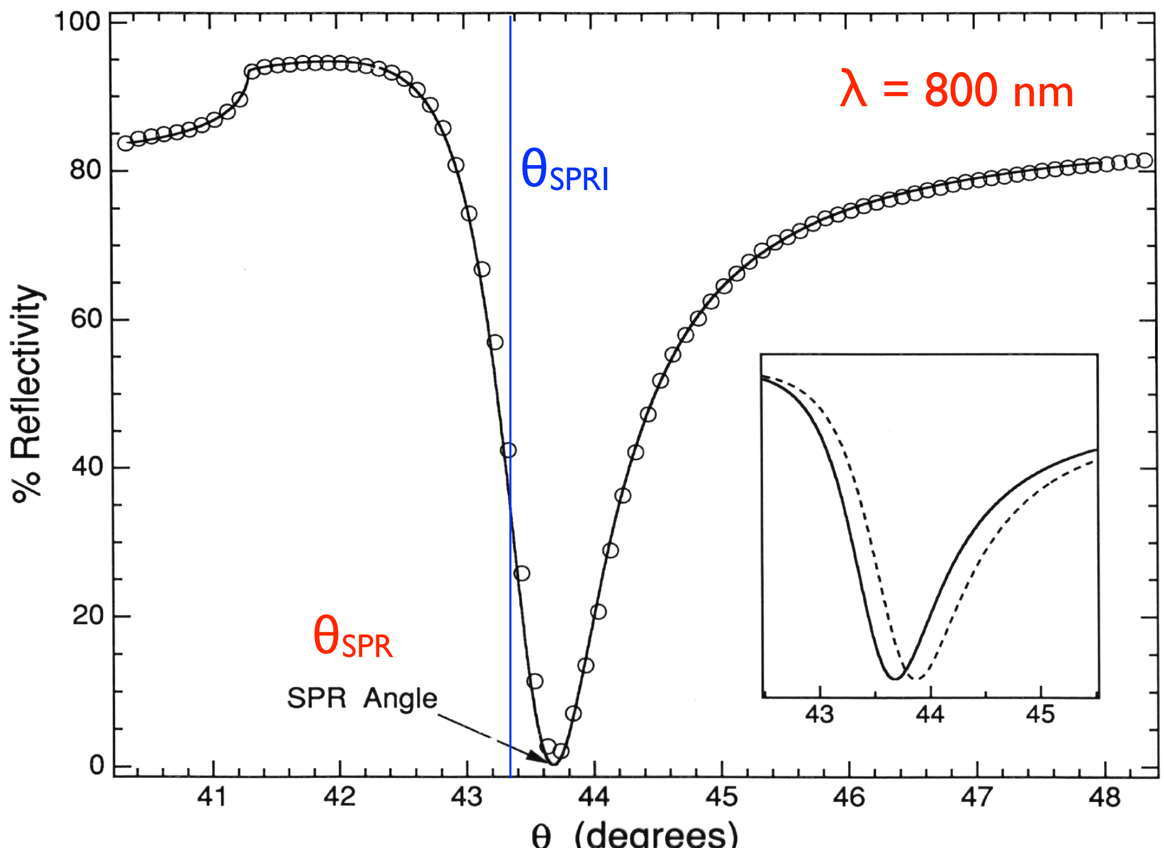
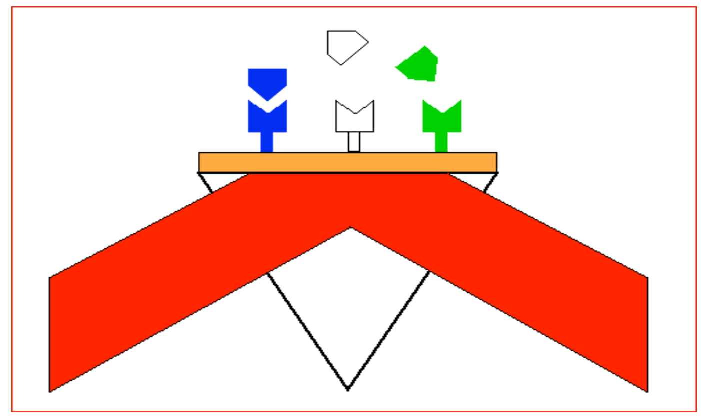


NIR SPR Imaging (SPRI)



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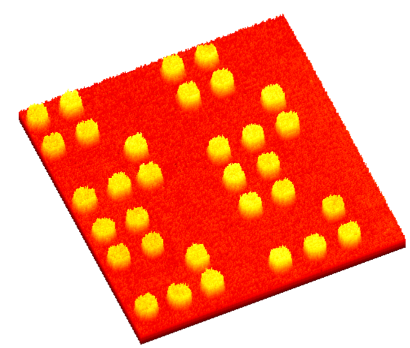

JM Brockman, AG Frutos, BP Nelson and RM Corn, Analytical Chem., 71 3928-3934 (1999).

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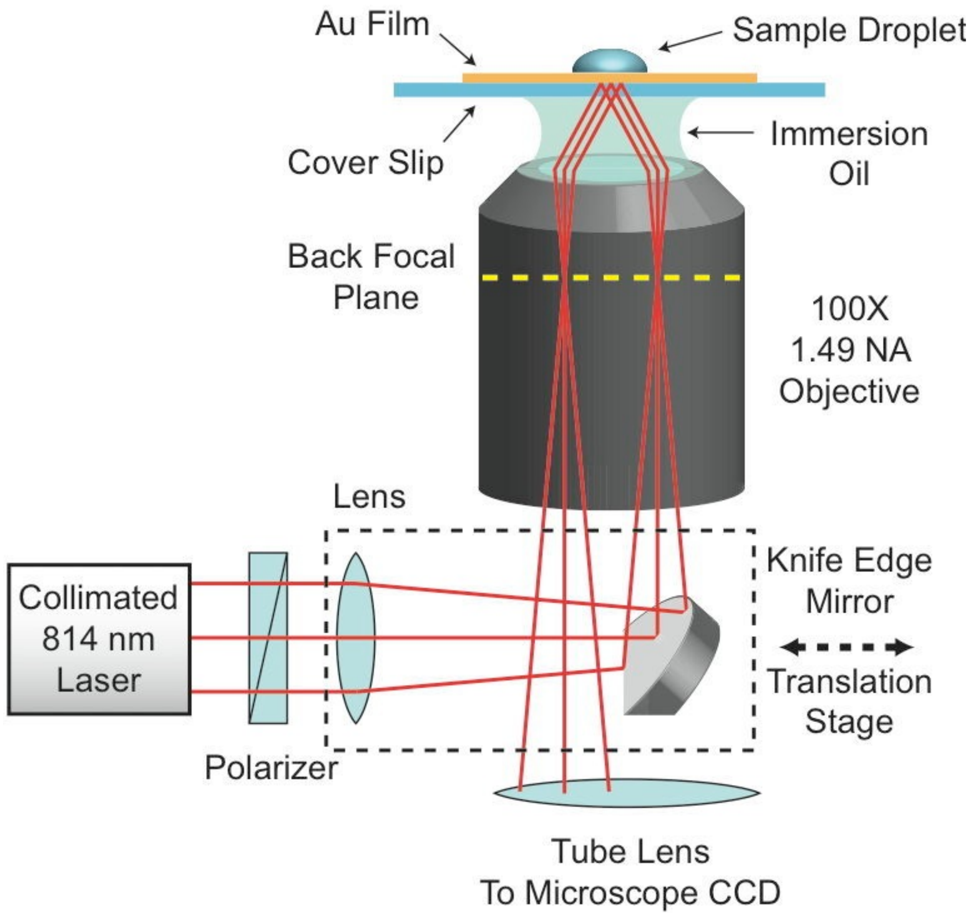
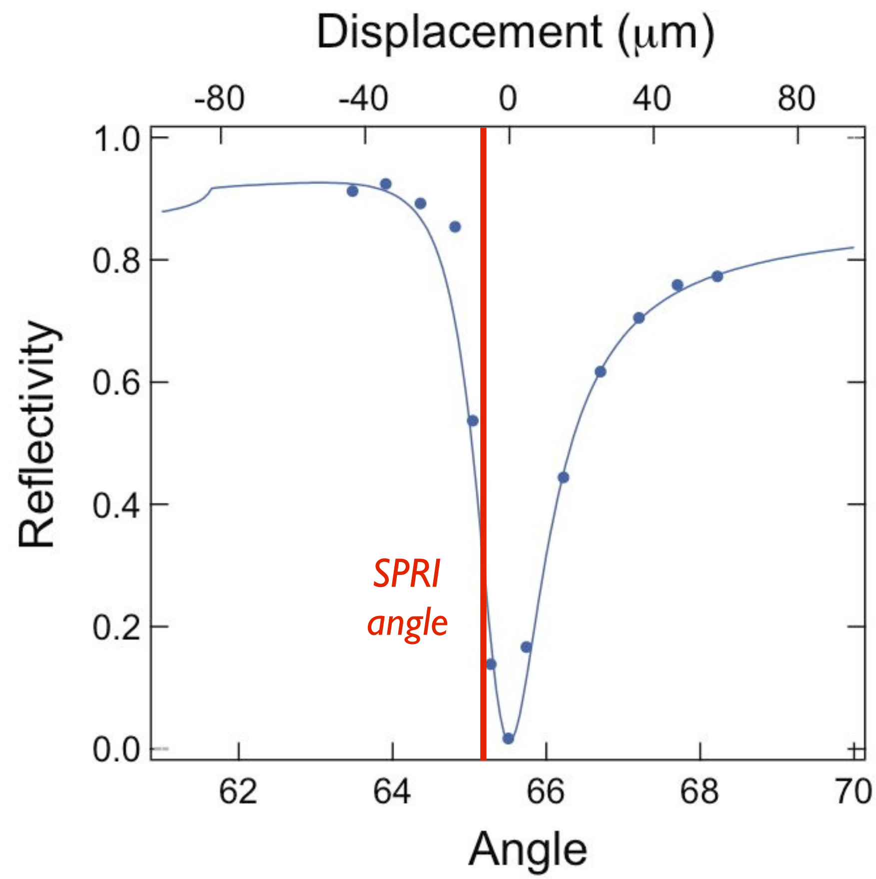
Near Infrared TIR SPR microscope: 814 nm





100x Objective
N.A. = 1.49
90 μm x 70 μm FOV

Control angle of incidence by varying d

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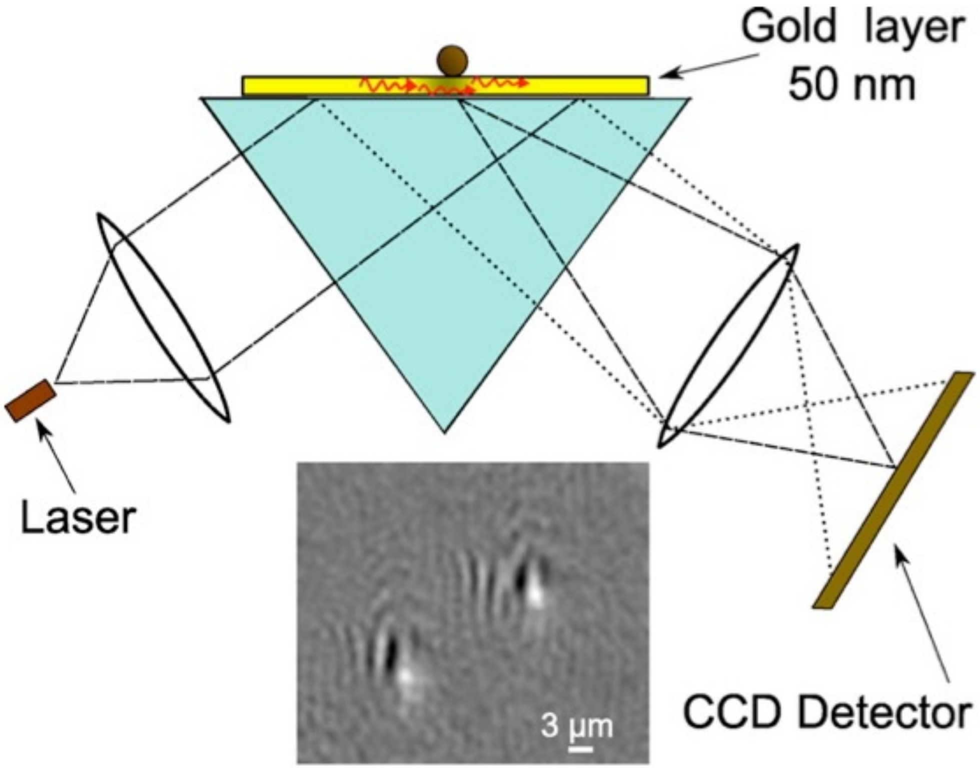
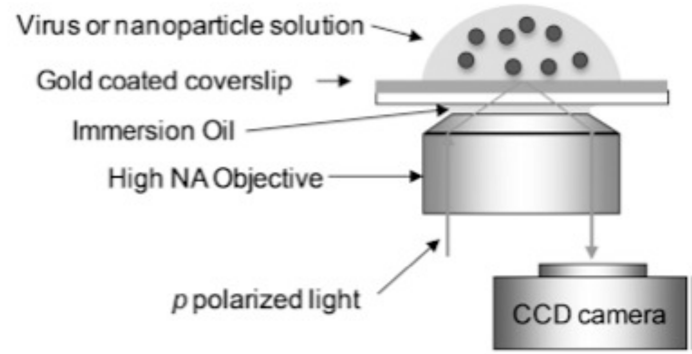
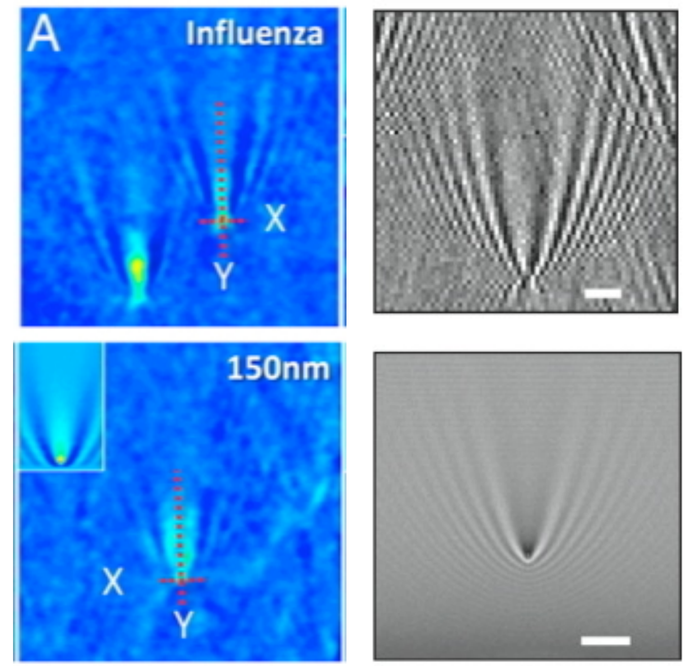
SPR microscopy of nanoparticles



Viruses and polystyrene NPs (40-200 nm)

A. Zybin et al.,
*Plasmonics **5**, 31–35 (2010).*
*Sens. Actuators B, **151** 281–290 (2010).*





Silica NP *80 nm Pt NP*

N. Tao et al.,
*Proc. Natl. Acad. Sci. **107**, 16028−16032 (2010).*
*Nature Nanotech. **134** 668-672(2012)*

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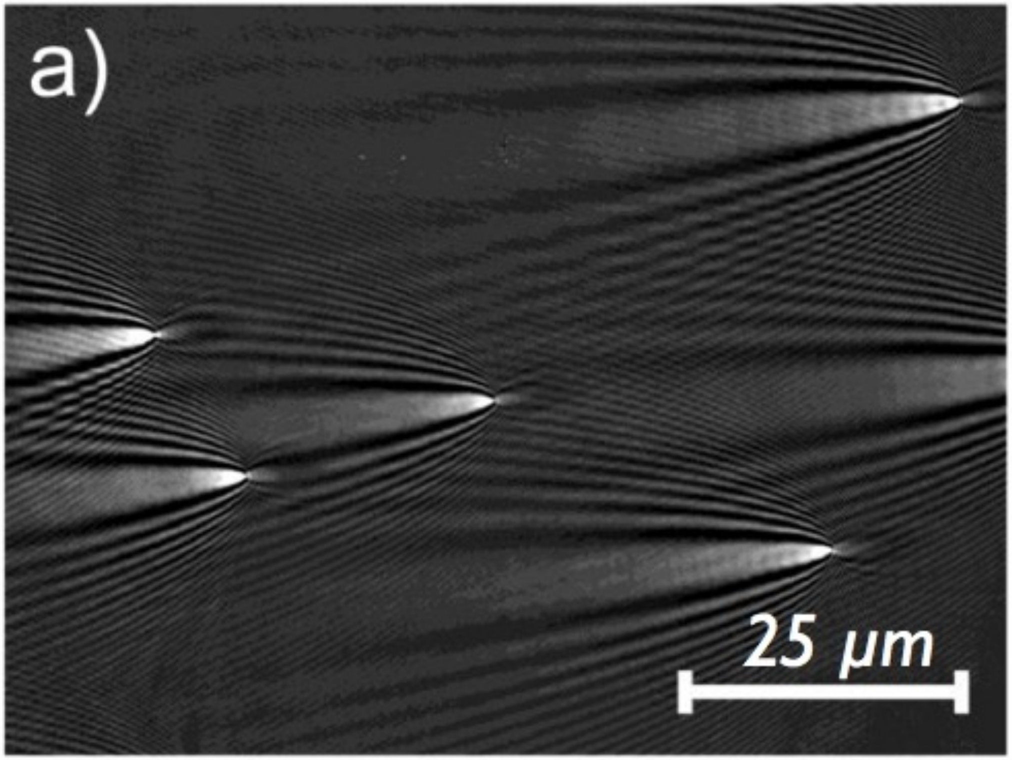
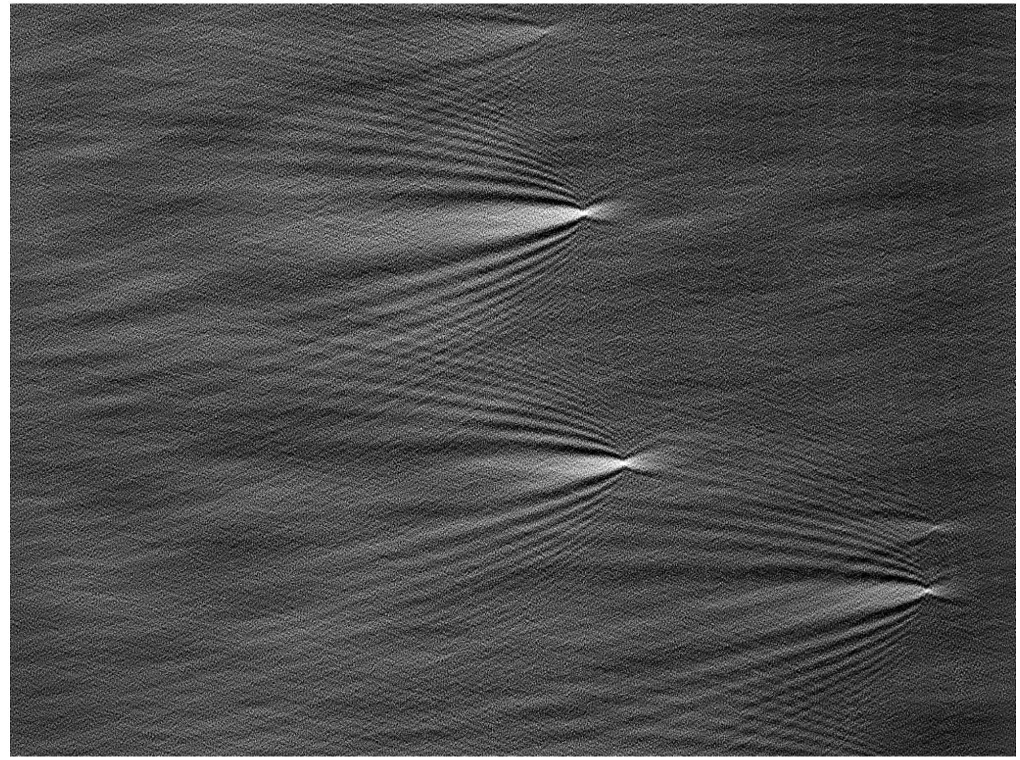
Near Infrared TIR SPRI microscope data:



200 nm polystyrene NPs



40 nm gold NPs

90 μm x 70 μm FOV

814 nm SPP excitation

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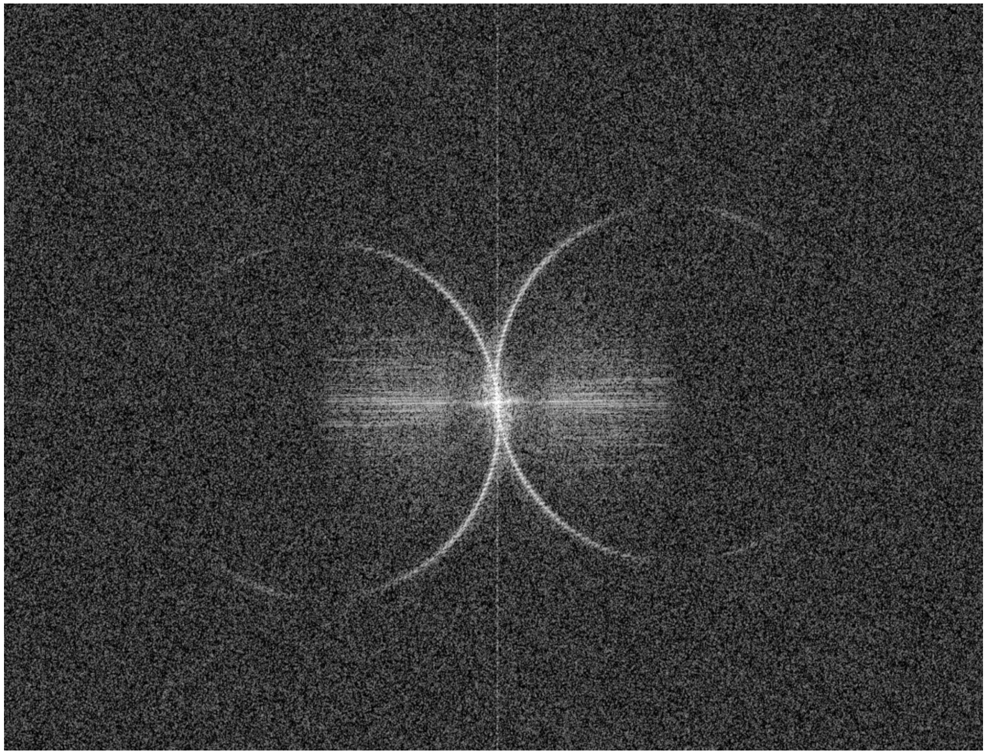
40 nm Gold Nanoparticles



2D FFT of CCD image

2D Helmholtz Equation
for wave propagation

$$(\nabla^2 + k_{sp}^2)u(x,y) = 0$$

$$(-\omega_x^2 + -\omega_y^2 + k_{sp}^2)u(\omega_x, \omega_y) = 0$$

Fourier Transform

$$\omega_x^2 + \omega_y^2 = k_{sp}^2$$

Resonance Condition

2D diffraction pattern of the surface plasmon polaritons off of the nanoparticle results in circles in the FFT of the image.

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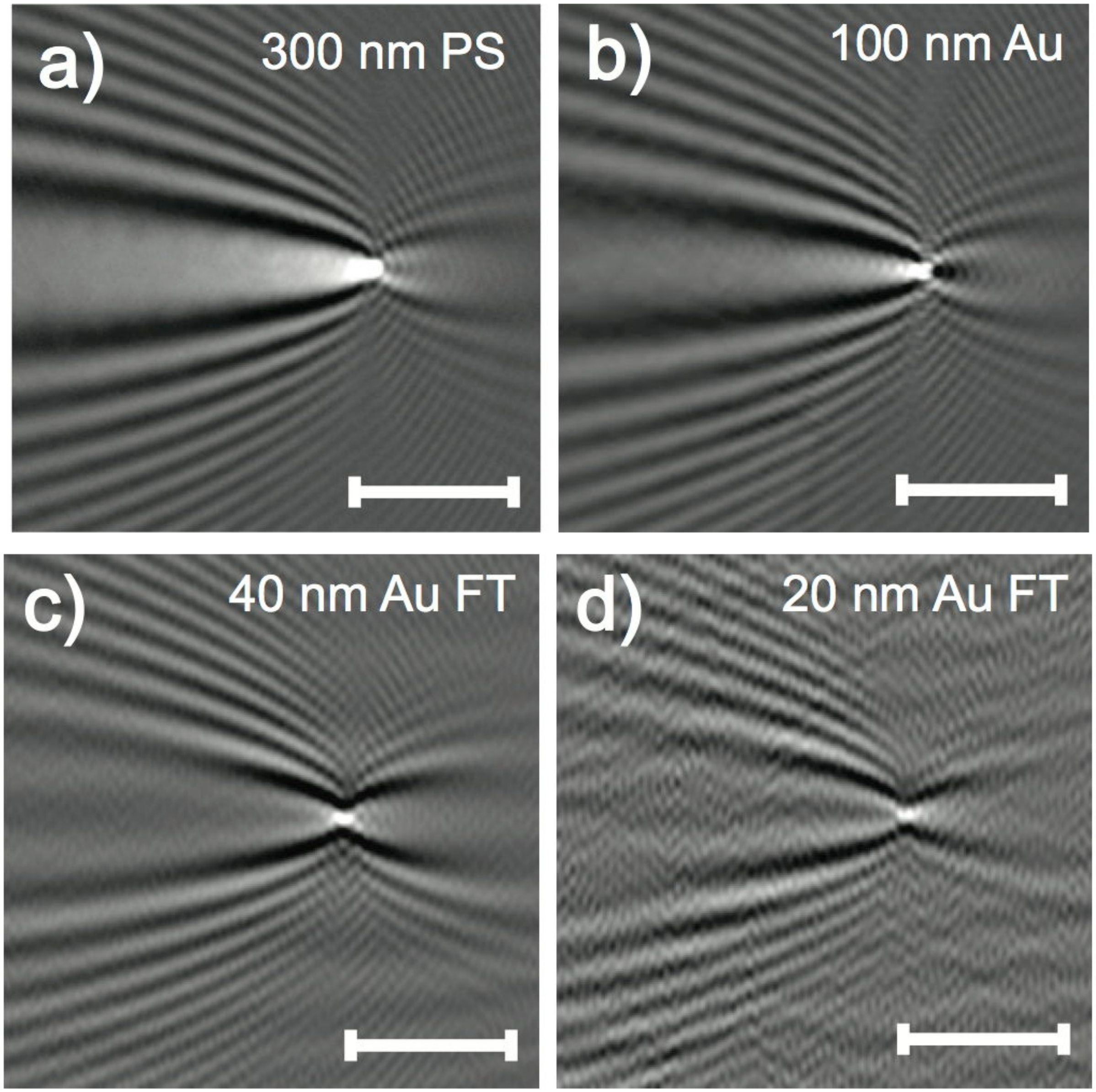
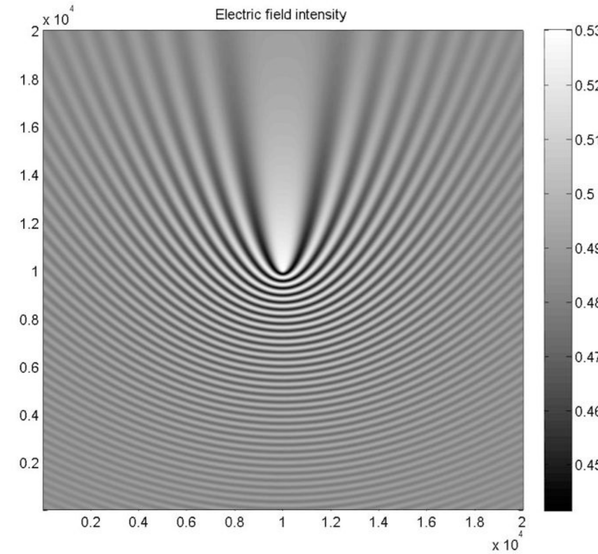
Near Infrared TIR SPR microscope: 814 nm SPP diffraction





Calculations from Prof. D. Kim at Yonsei University

All particles have similar functional form

Tail periodicity due to SP propagation length at 814 nm

Difference Image in Aqueous Solution
3 sec integration time

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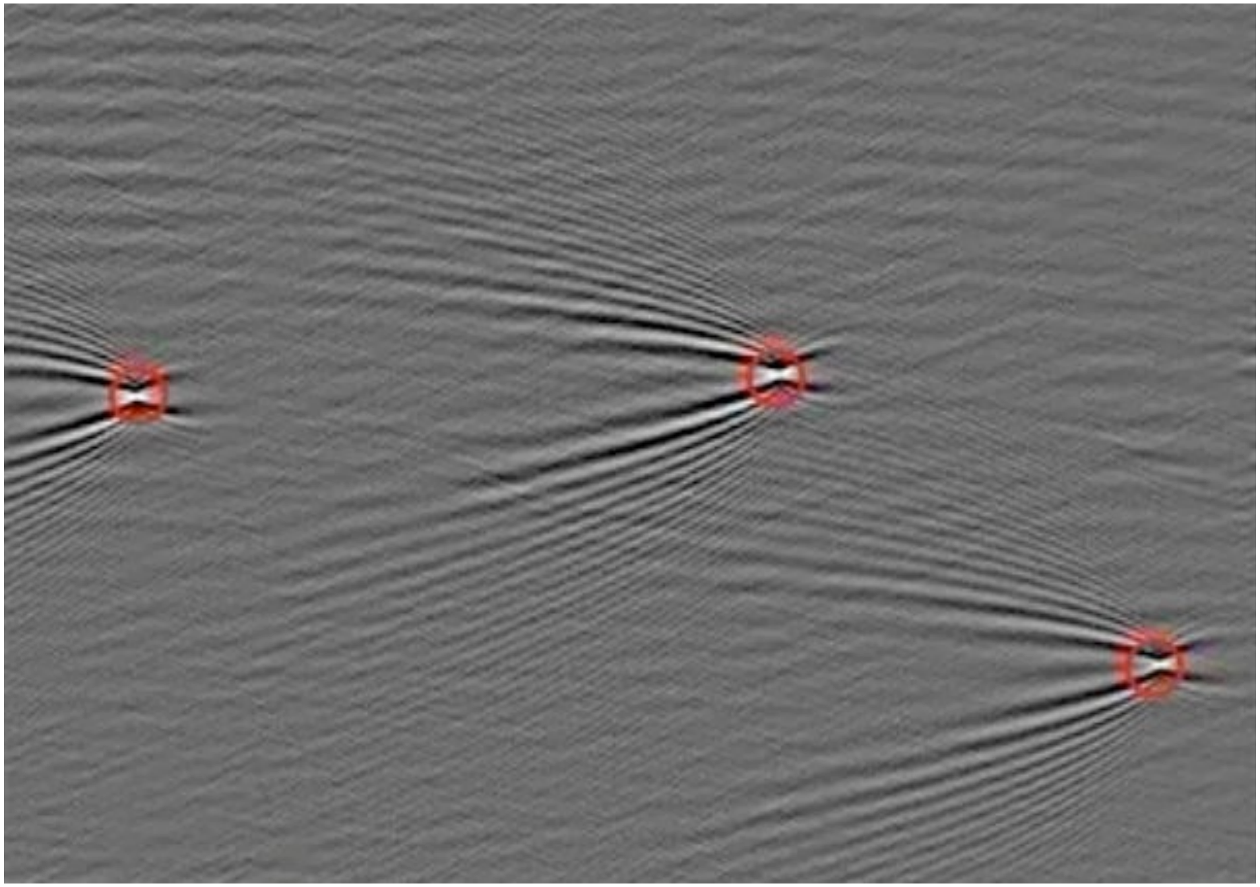
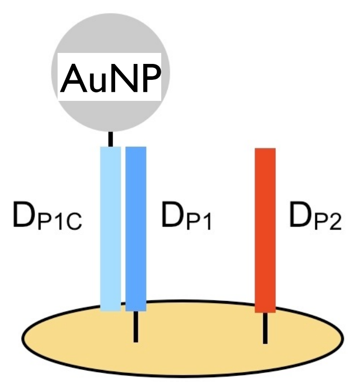
Bioaffinity Adsorption of 40 nm DNA-modified Au Nanoparticles



A 3 second frame image from a 1 pM solution with three Au NPs

100% complementary surface



Initial solution: 3 µL

Add 3 µL of AuNPs

Record SPR images every 3 seconds

Create difference image movie

Count NPs in each frame

Create Adsorption Kinetics Plot

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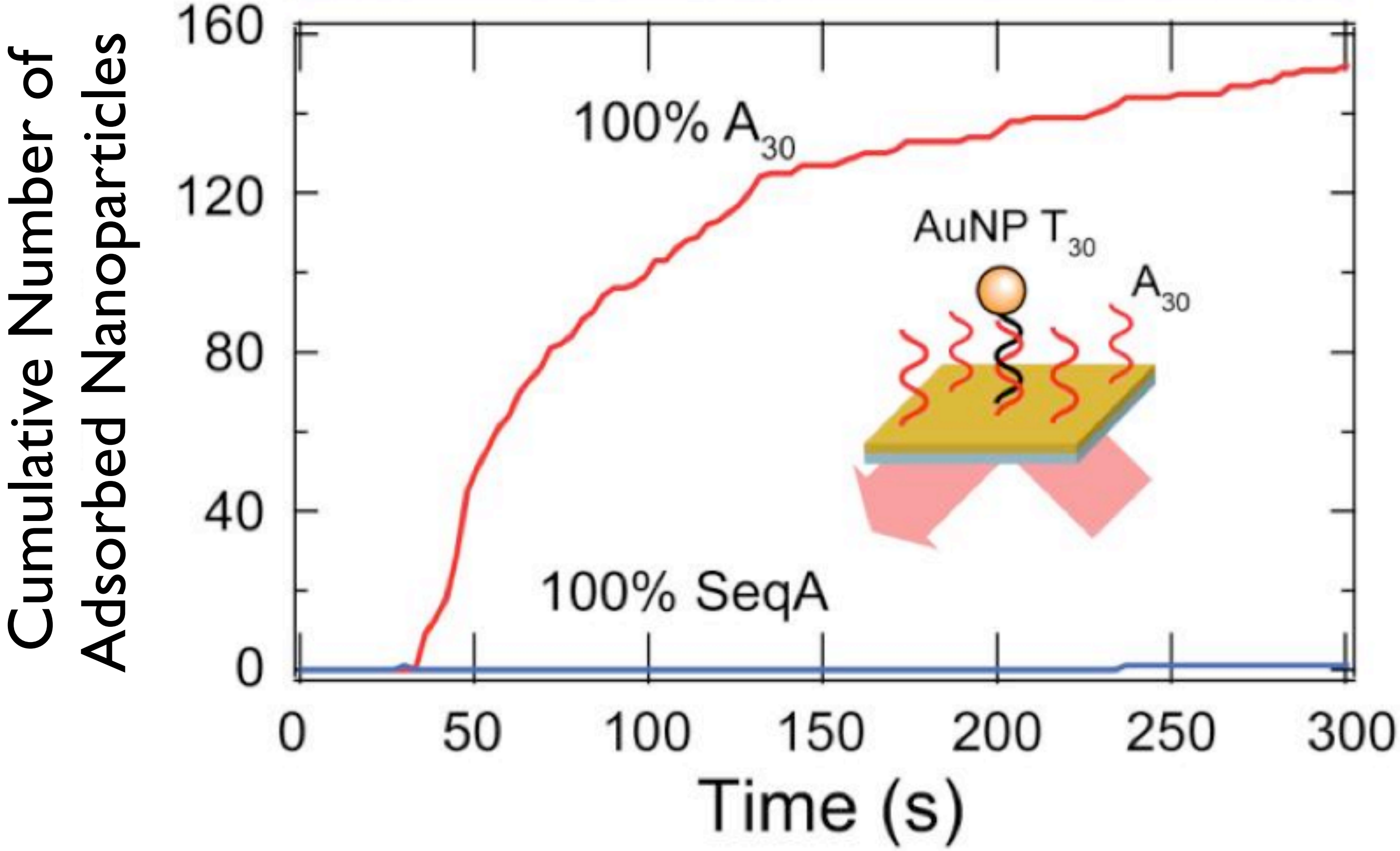
Bioaffinity Adsorption of 40 nm DNA-modified Au Nanoparticles



1 pM solution of 40 nm DNA-modified AuNPs

Noncomplementary DNA-modified surface shows negligble nanoparticle binding

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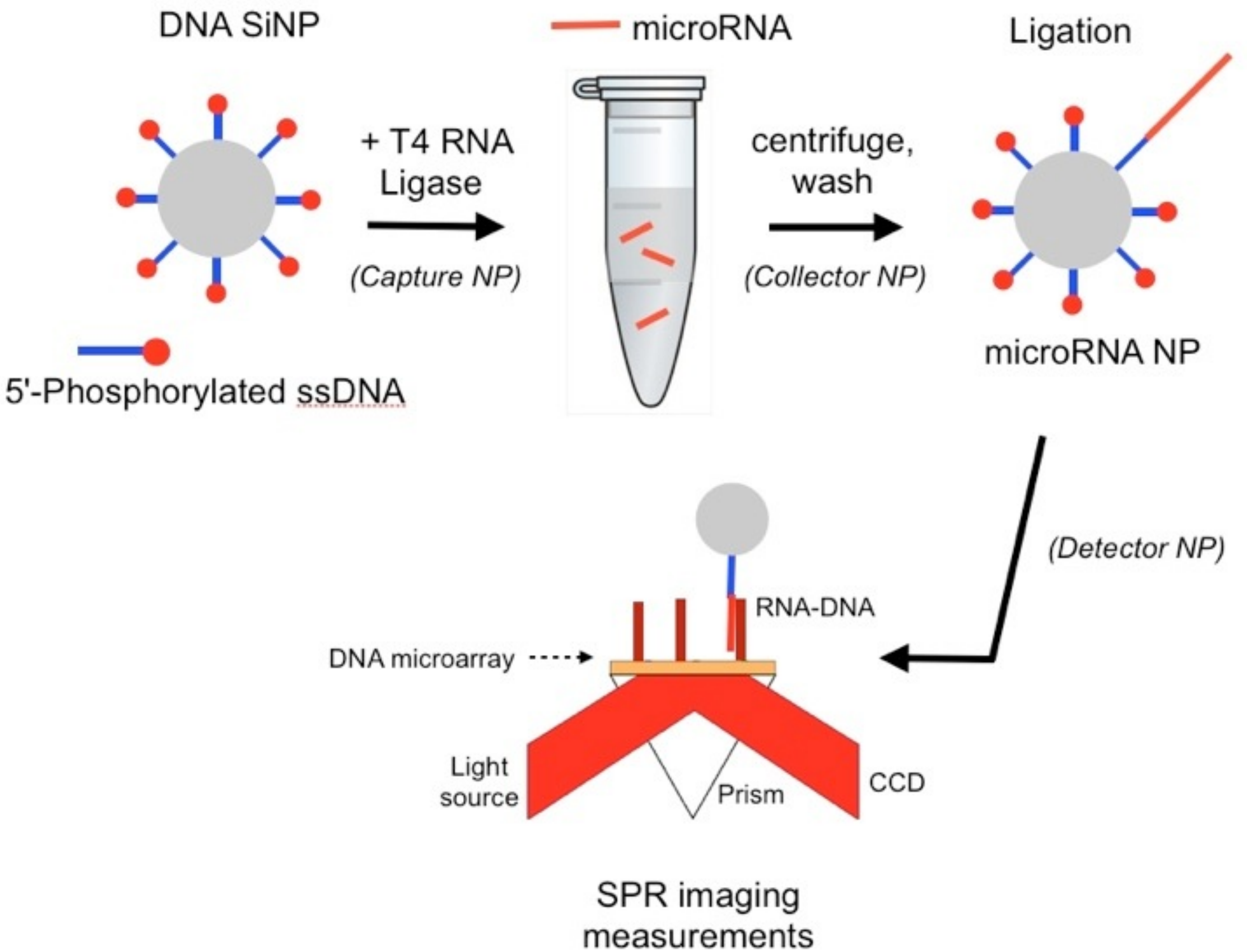
Surface Enzyme Chemistry + Nanoparticle-Enhanced SPRI

- MicroRNA Detection with Enzymatic Nanoparticles: Ligation Capture



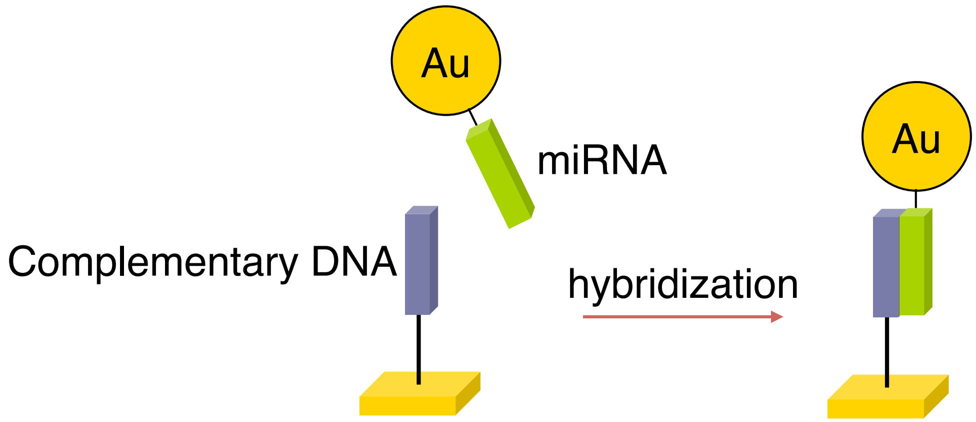
Dr. Yulin Chen

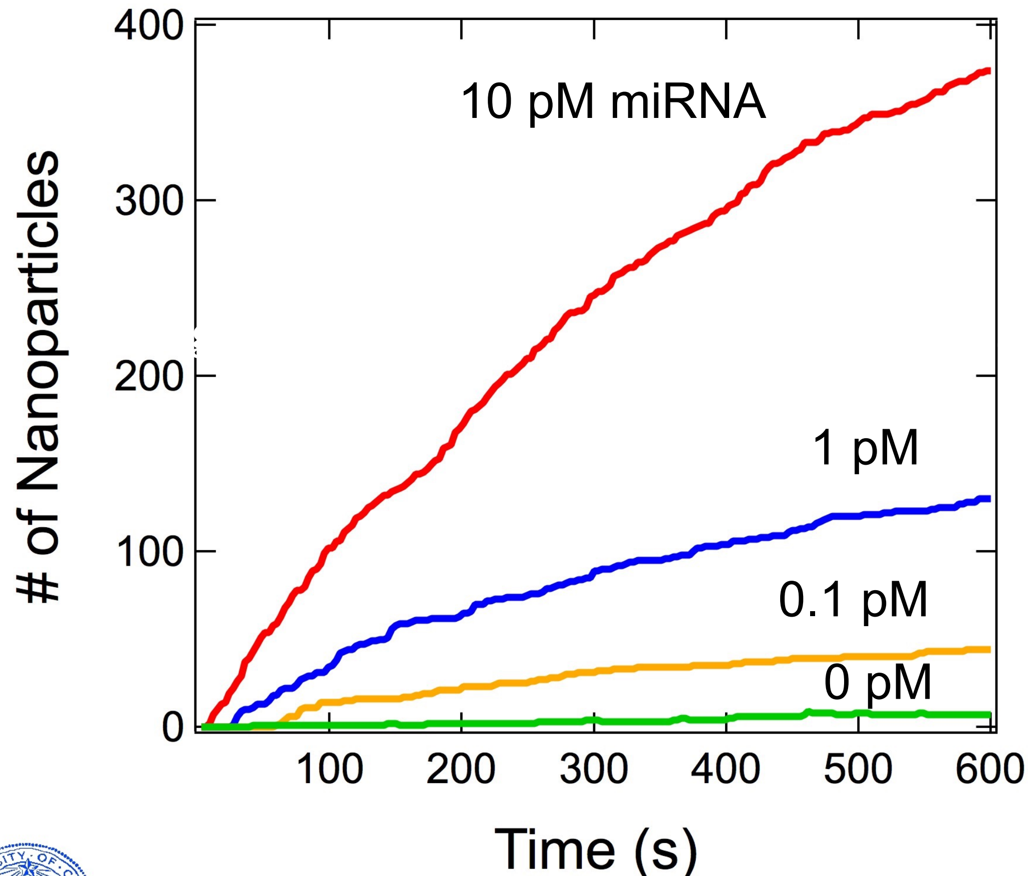
Dr. WenJuan Zhou

T4 RNA Ligation for miRNA capture

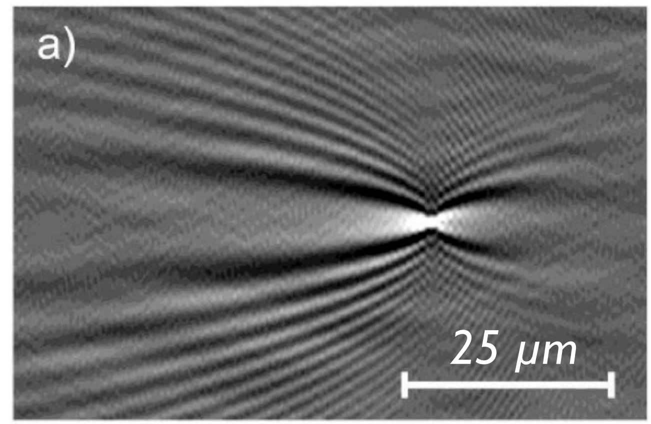
W. J. Zhou et al., Analytical Chemistry, ***83*** *3897-3902 (2011).*

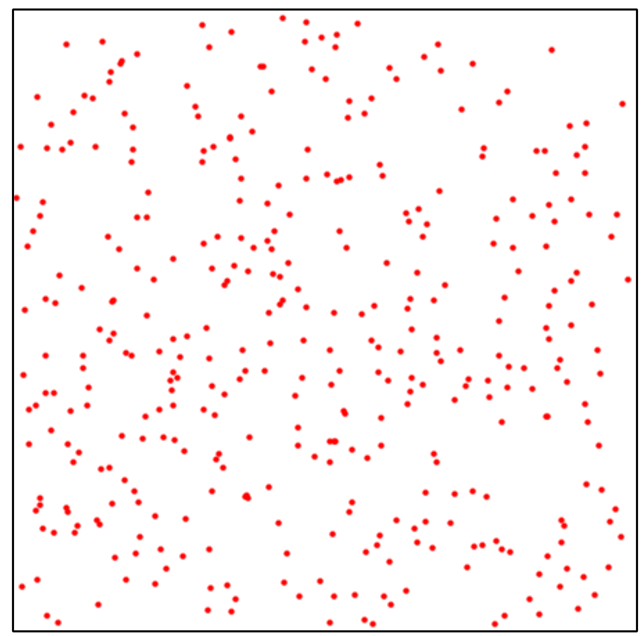
*Single NP Detection of miRNA-146a**





Single NP adsorption event





NP adsorption map (10 pM)

Total Concentration of AuNPs: 25 pM

** UGAGAACUGAAUUCCAUAGGC*

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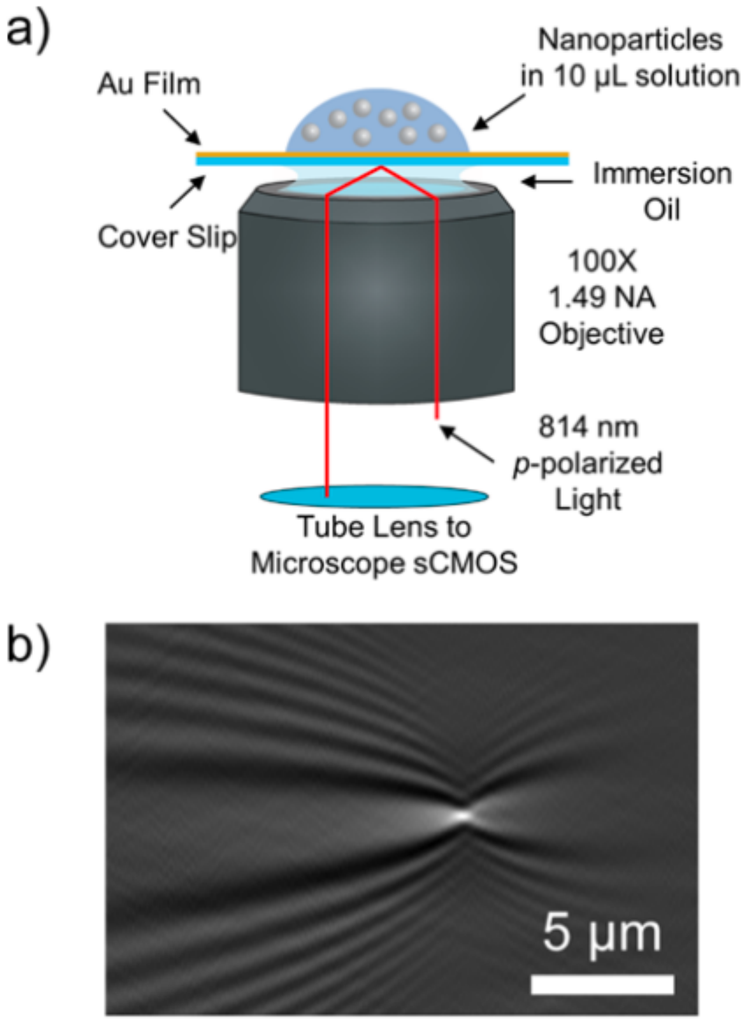
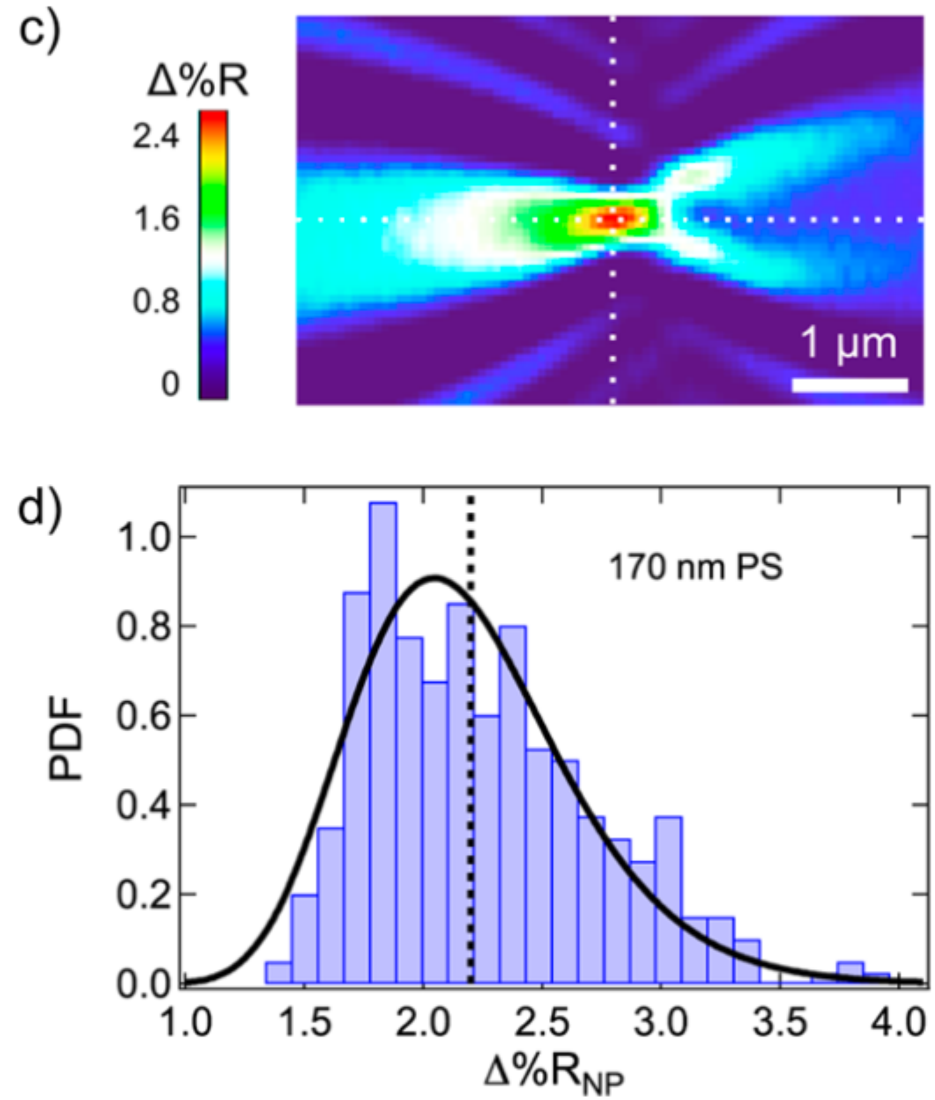
Single Nanoparticle Distribution Measurements





$\Delta\%R_{NP}$ *"Single Nanoparticle SPRI Reflectivity Change"*

A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, ***11*** *7447-7456 (2017).*

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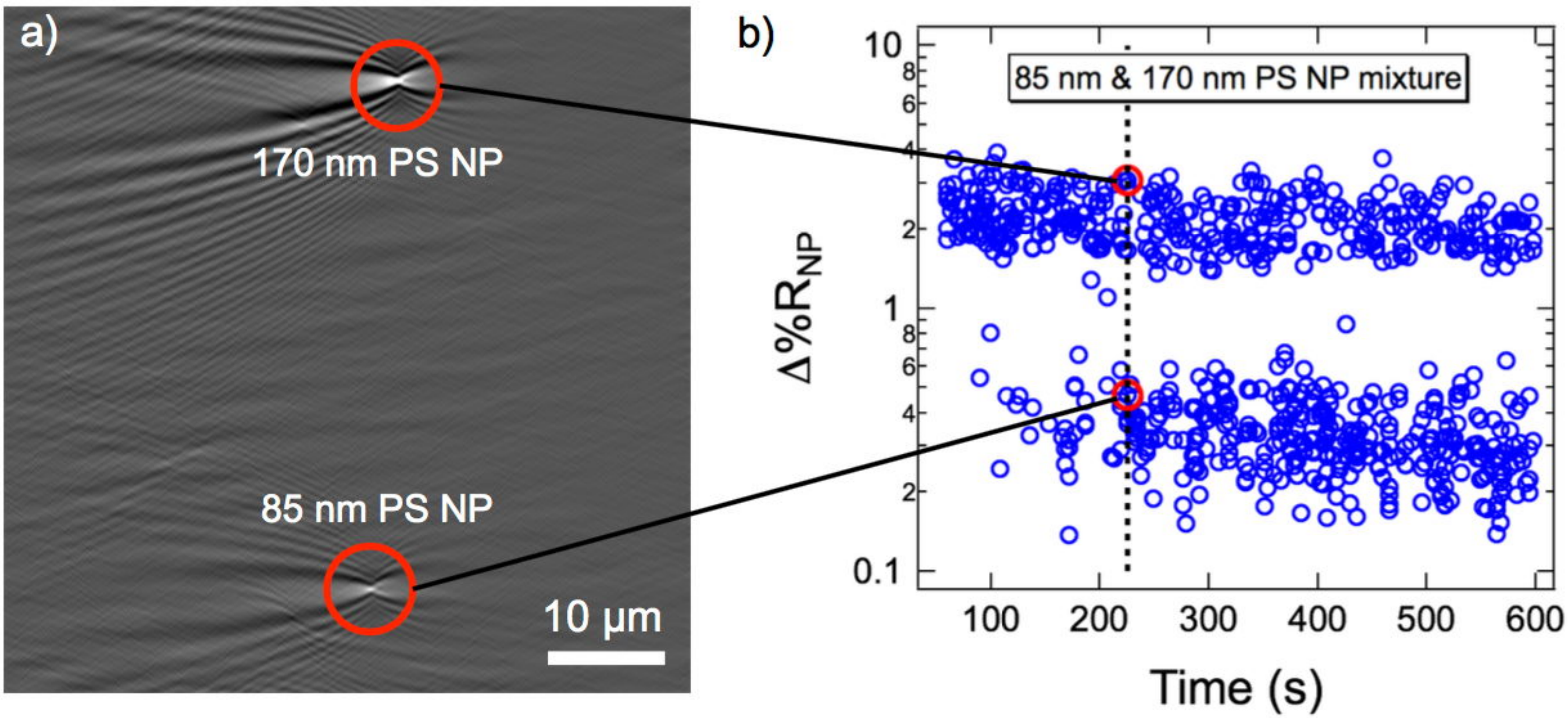
Single Nanoparticle Distribution Measurements

1:1 Mixture of 85 nm and 170 nm PS nanoparticles



Irreversible electrostatic adsorption of carboxy-modified PS NPs onto an amino-terminated alkanethiol monolayer (MUAM).

*A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, **11** 7447-7456 (2017).*

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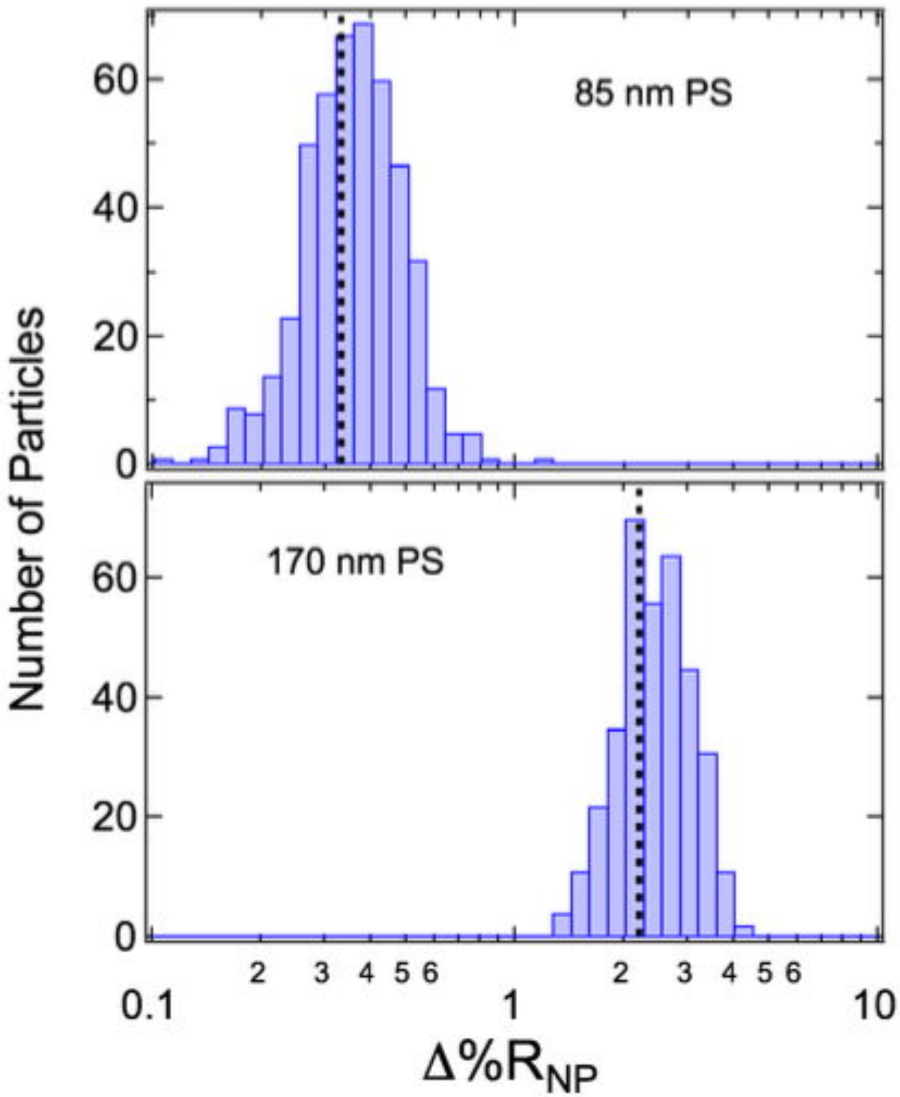
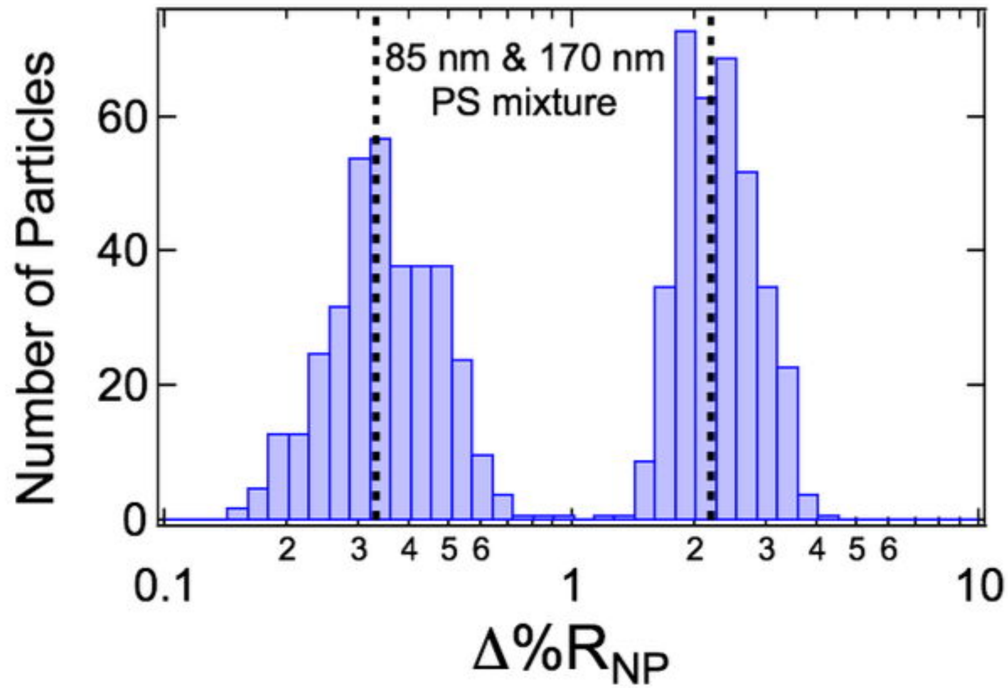
Single Nanoparticle Distribution Measurements



Mixture of Two Polystyrene NPs



A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, ***11*** *7447-7456 (2017).*

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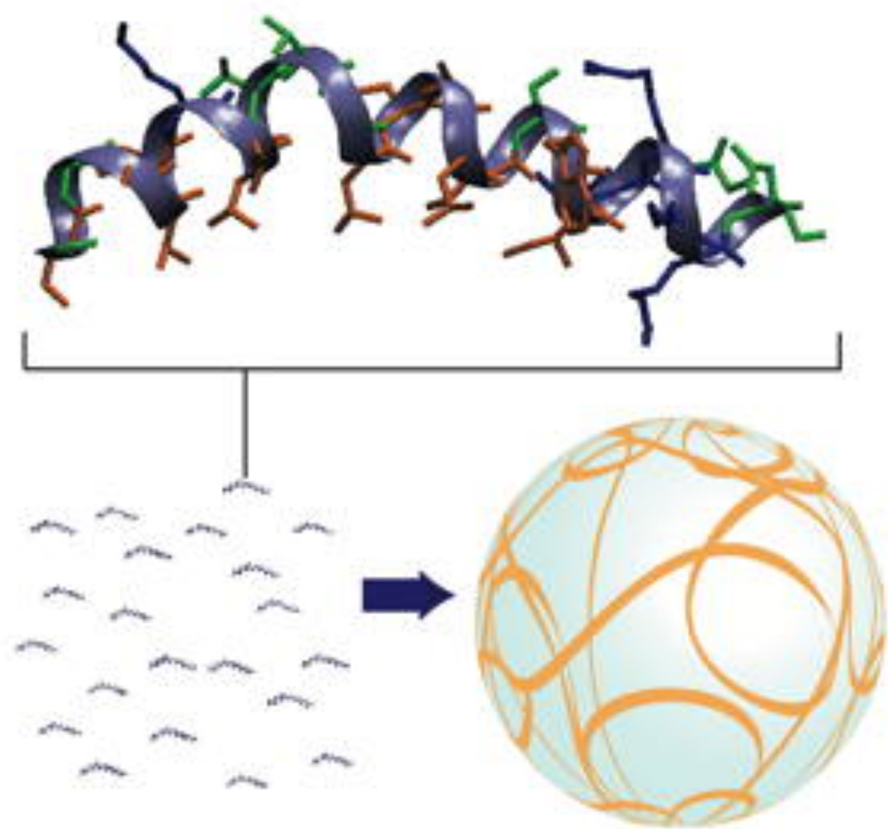
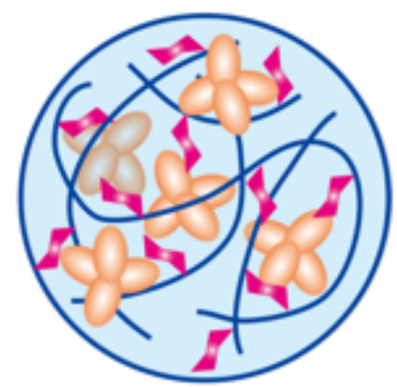
Bioaffinity Uptake in Nanoparticles



Noncovalent bioaffinity interactions can also be used to absorb biomolecules (peptides, proteins, metabolites) into porous hydrogel, silica and liposomal nanoparticles.

Melittin peptide uptake into hydrogel nanoparticles (HNP) for drug delivery

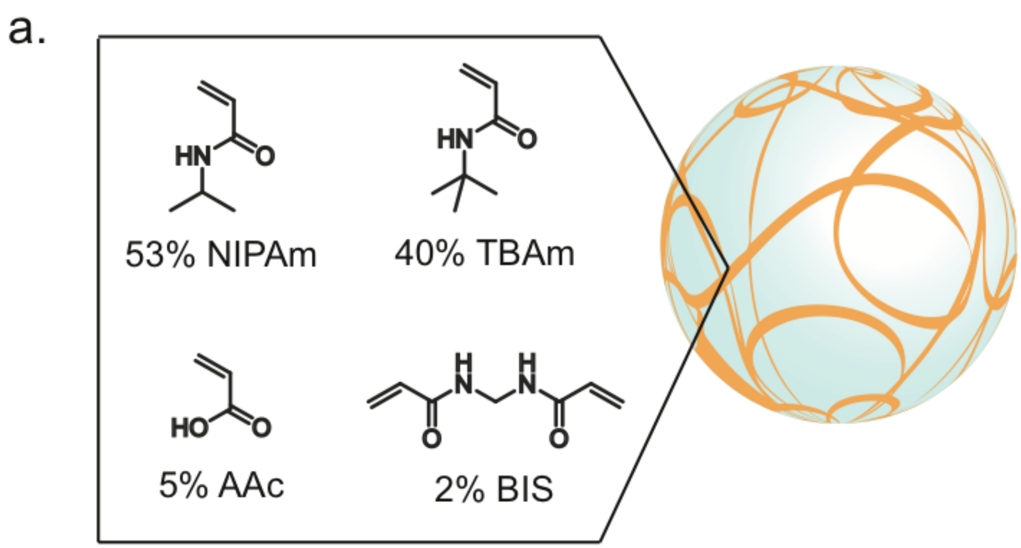


ConA and mannose (HNPs)

Melittin: GIGAVLKVLTTGLPALISWIKRKRQQ

NIPam-based Hydrogel Nanoparticles

220 nm HNPs

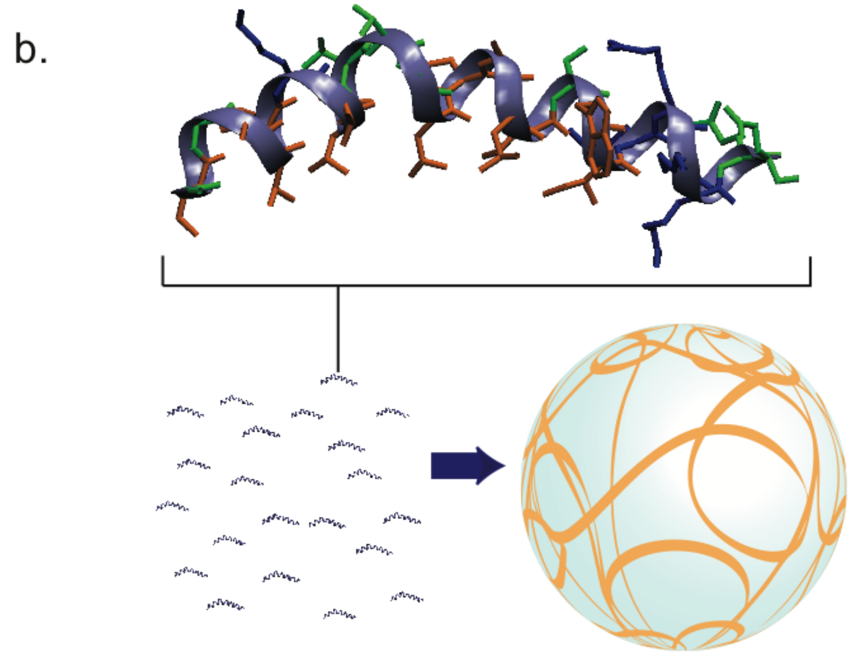


220 nm diameter (from DLS)

1.24 E09 Mol.Wt. (from MALS)

approx. 65% solvent (PBS) by volume

Melittin



26 residue peptide
from bee venom

GIGAVLKVLTTGLPALISWIKRKRQQ

NIPam: N-isopropylacrylamide

Collaborator: Prof. Ken Shea

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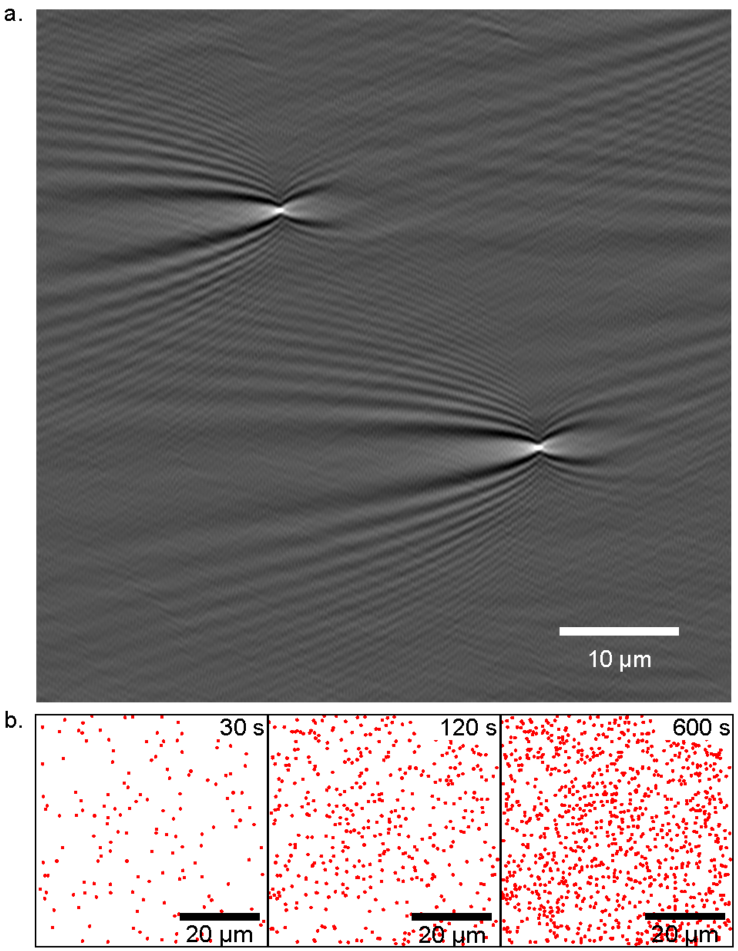
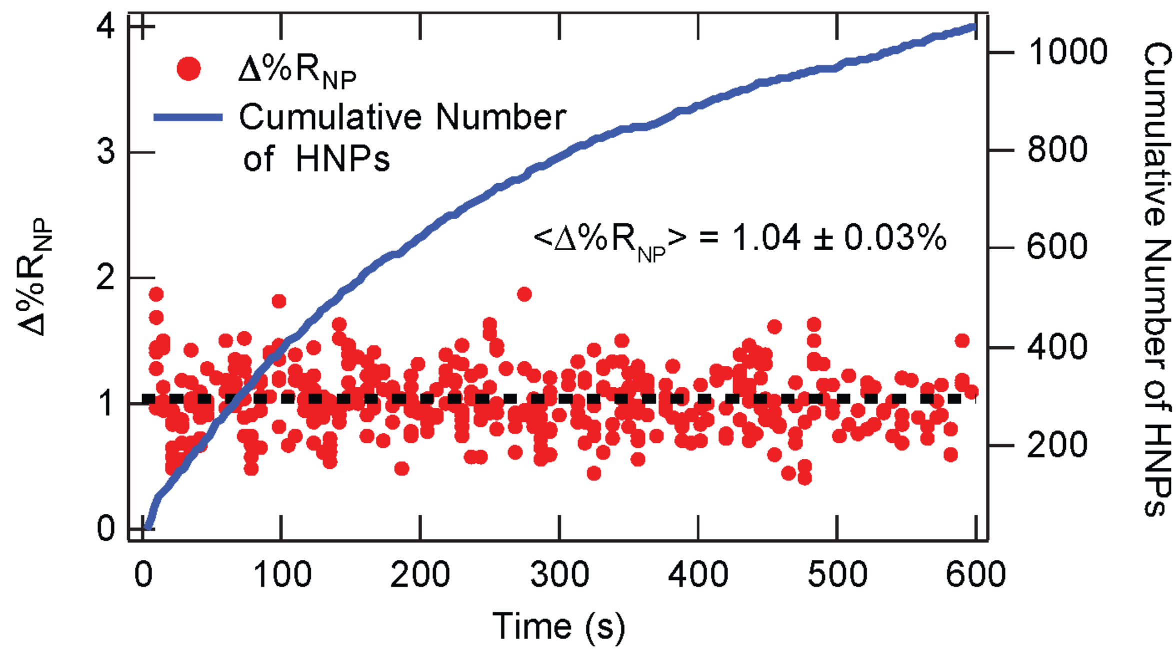
Single Nanoparticle SPRI 180 nm Hydrogel NPs





NIPam-based HNPs irreversibly adsorb onto C11-functionalized gold surfaces

2D maps of HNP adsorption after 30, 120 and 600 seconds

C11: undecanethiol

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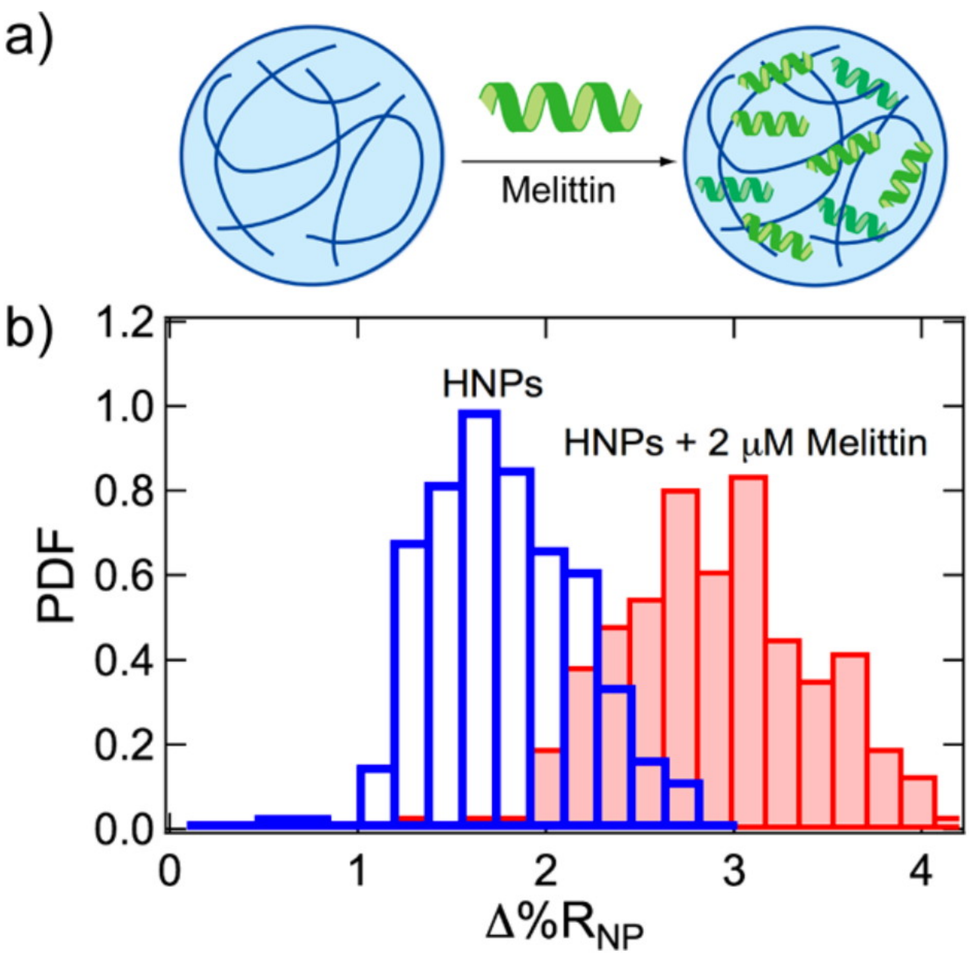
$<\Delta\%R_{NP}>$ for HNPs in the presence of Melittin

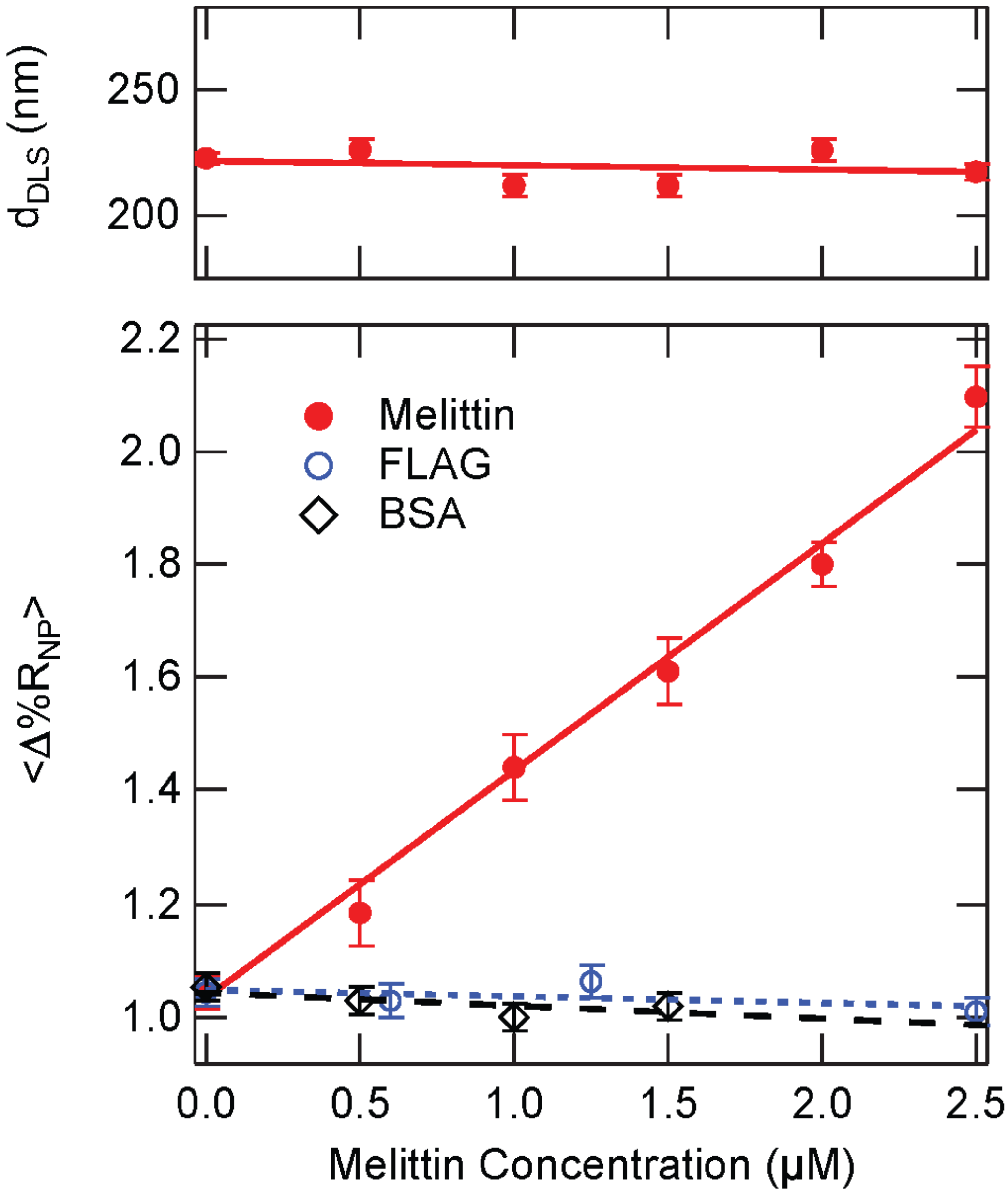


$<\Delta\%R_{NP}>$ for the HNPs increases with melittin concentration as the uptake of peptide increases the refractive index of the hydrogel nanoparticle.

Mike Cho

K. Cho et al. Analytical Chemistry, ***87*** *4973-4979 (2015).*

$\langle\Delta\%R_{NP}\rangle$ for HNPs in the presence of Melittin



$\langle\Delta\%R_{NP}\rangle$ for the HNPs increases with melittin concentration as the uptake of peptide increases the refractive index of the hydrogel nanoparticle.

Fluorescence measurements of the loss of melittin in solution are used to show that a 0.2% increase in $\langle\Delta\%R_{NP}\rangle$ corresponds to the uptake of 12,500 molecules per HNP on average.

Mike Cho

*K. Cho et al. Analytical Chemistry, **87** 4973-4979 (2015).*

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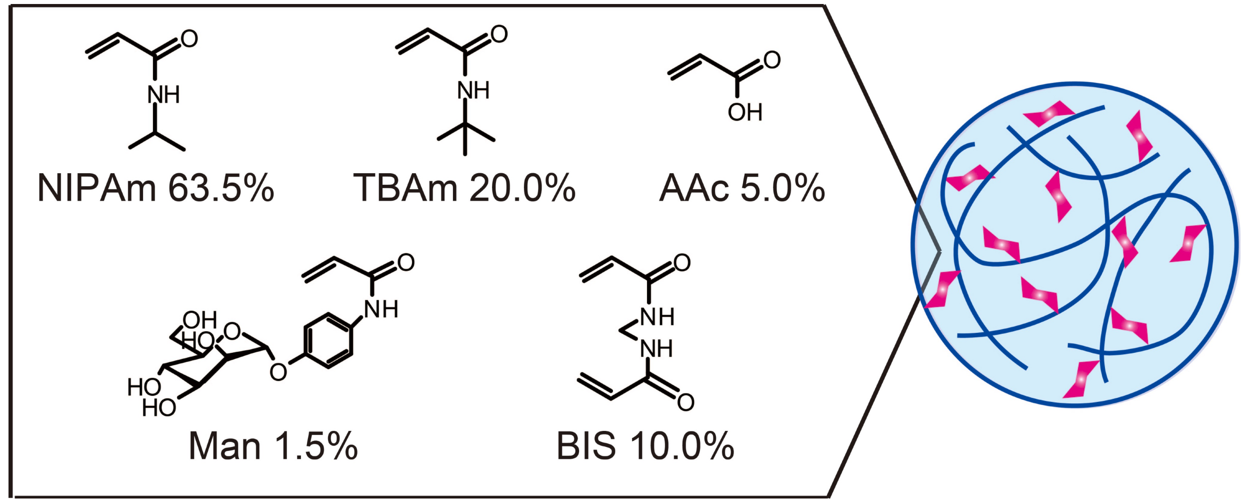
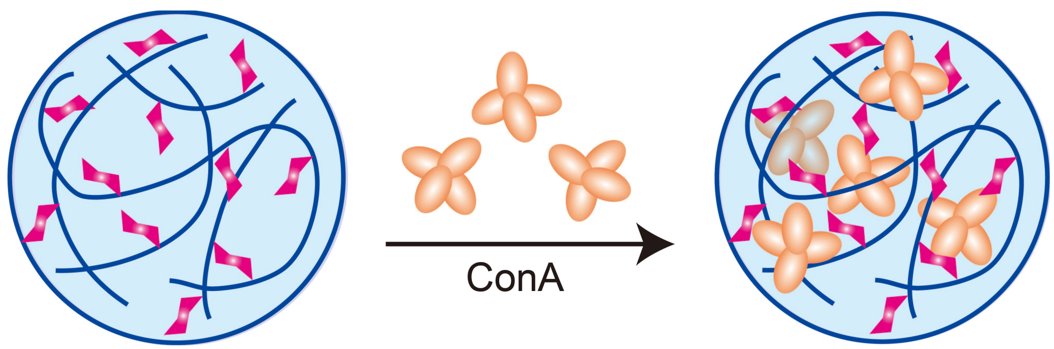
ConAuptake into HNPs

a)



b)



Yuhei

With Yuhei Terada and Prof. Yoshiko Miura (Kyushu Univ.)

A. M. Maley, Yuhei Terada et al., J. Phys. Chem. C, ***120*** *16843-16849 (2016).*

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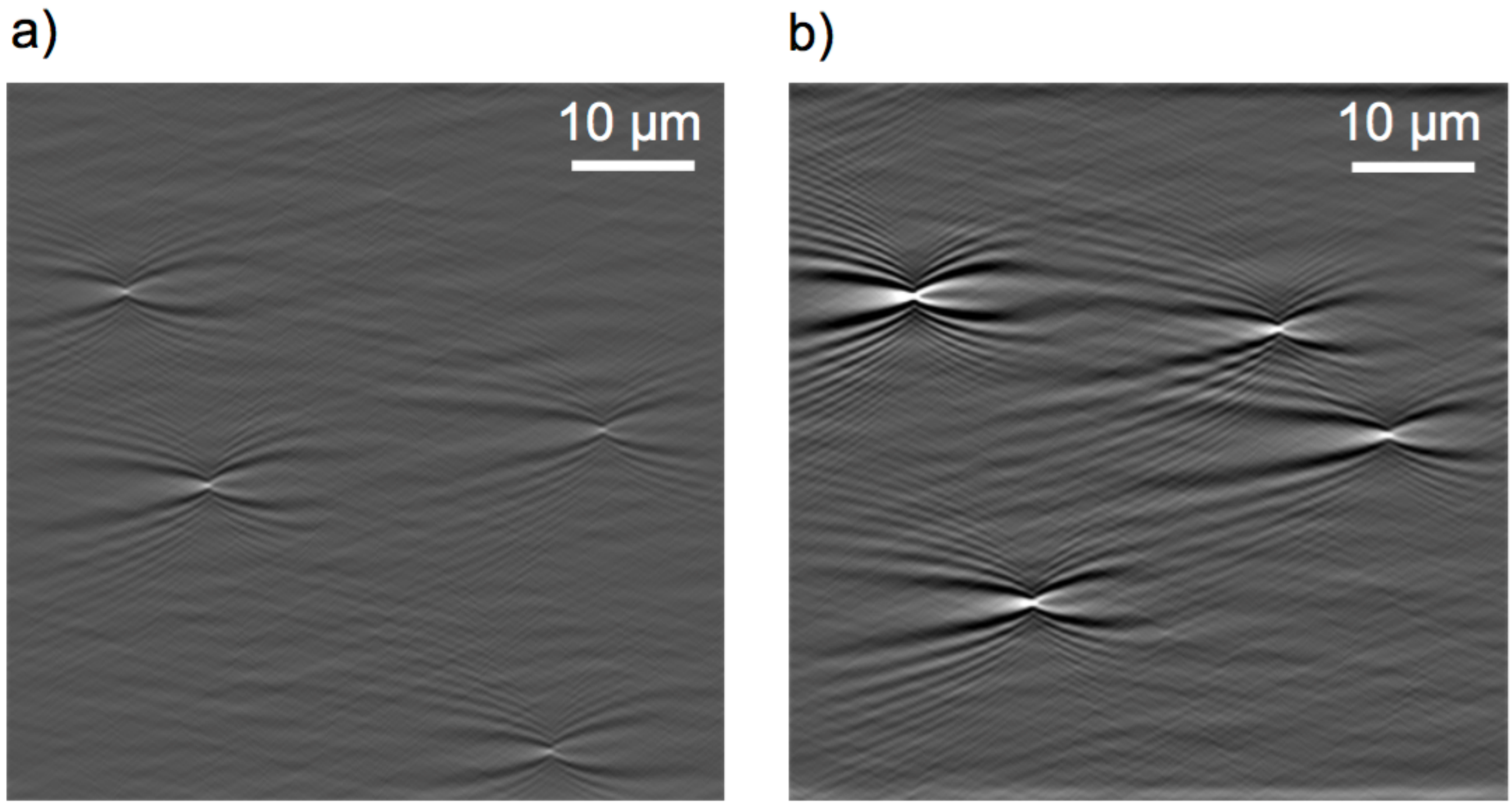
ConAuptake into HNPs



180 nm mHNPs | 180 nm mHNPs + 1µM ConA

Significant increase in the single nanoparticle response in the presence of ConA.

A. M. Maley, Yuhei Terada et al., J. Phys. Chem. C, ***120*** *16843-16849 (2016).*

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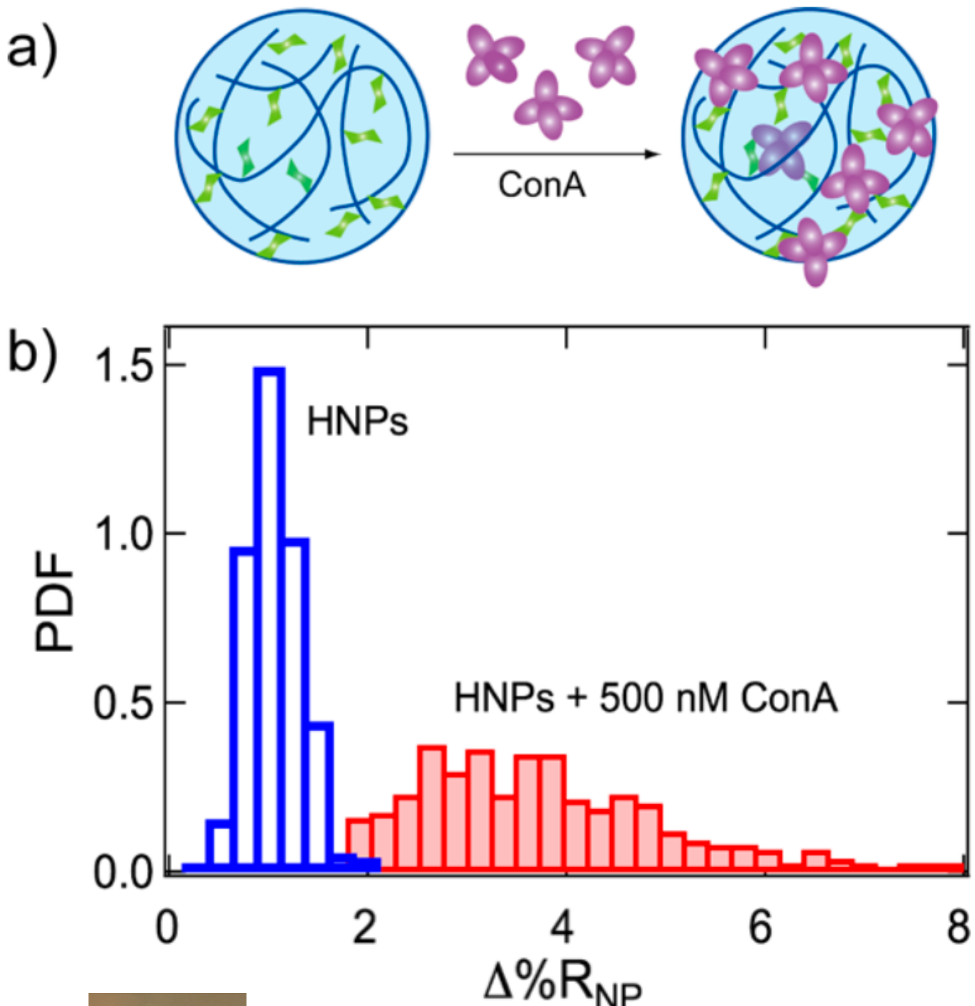
ConA uptake into HNPs



Both $\langle \Delta\%R_{NP} \rangle$ and the distribution width increases with ConA concentration as the uptake of ConA both increases the refractive index of the hydrogel nanoparticle and allows for inter-nanoparticle binding

Adam Maley

*A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, **11** 7447-7456 (2017).*

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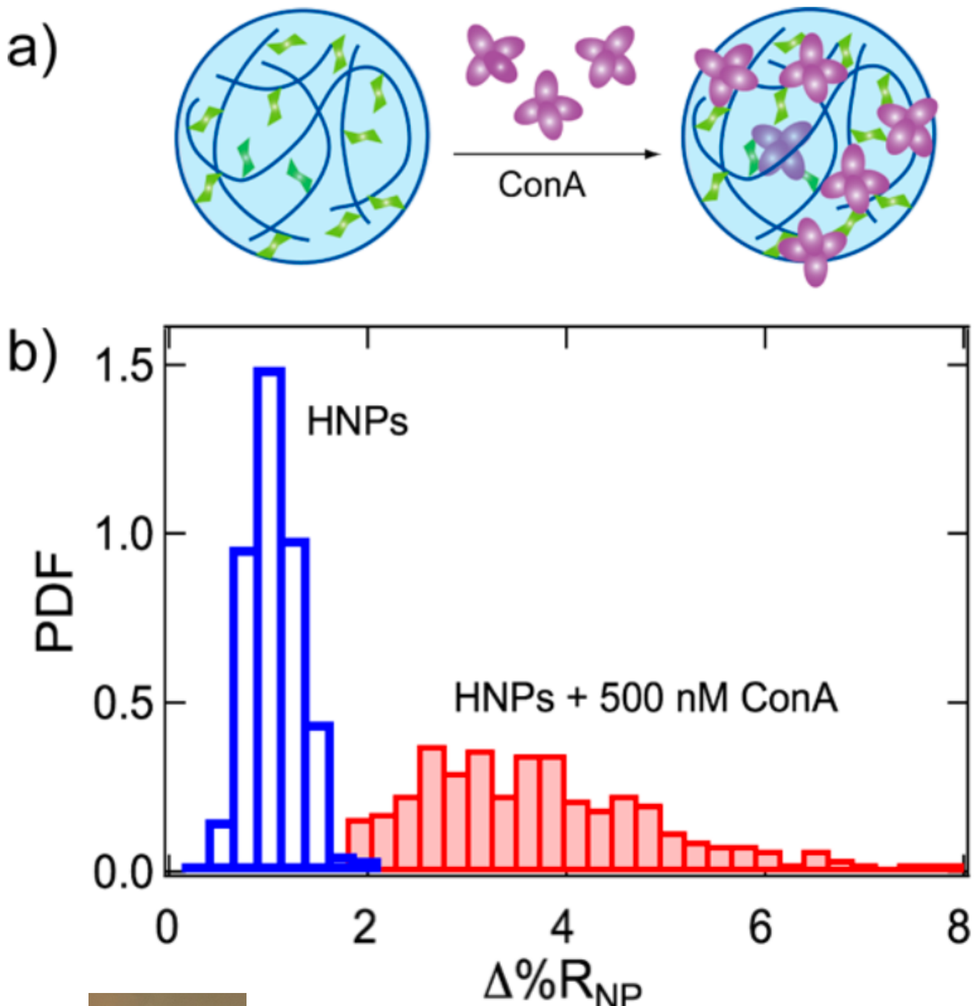
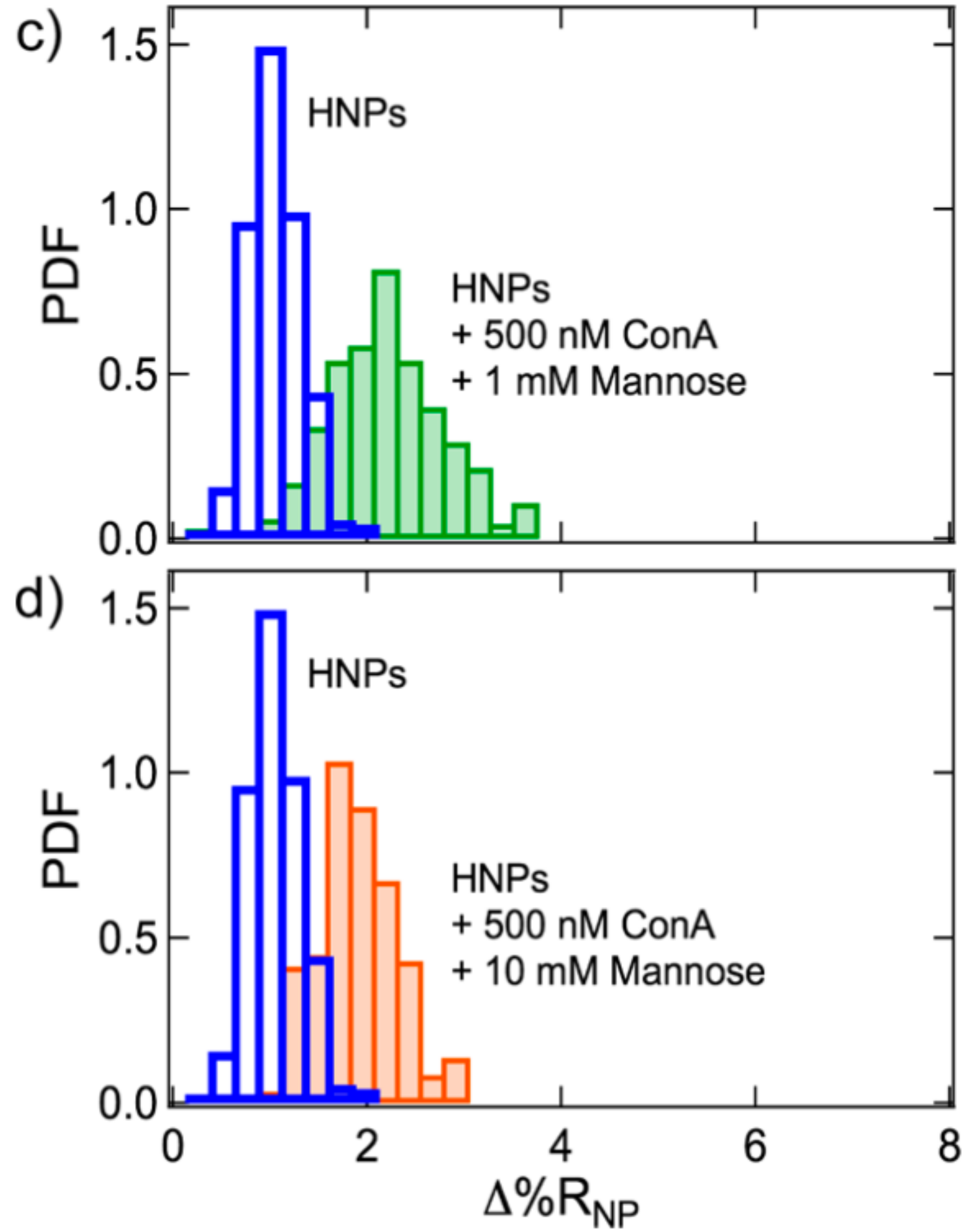
ConA uptake into HNPs



Mannose competition



Adam Maley

A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, ***11*** *7447-7456 (2017).*

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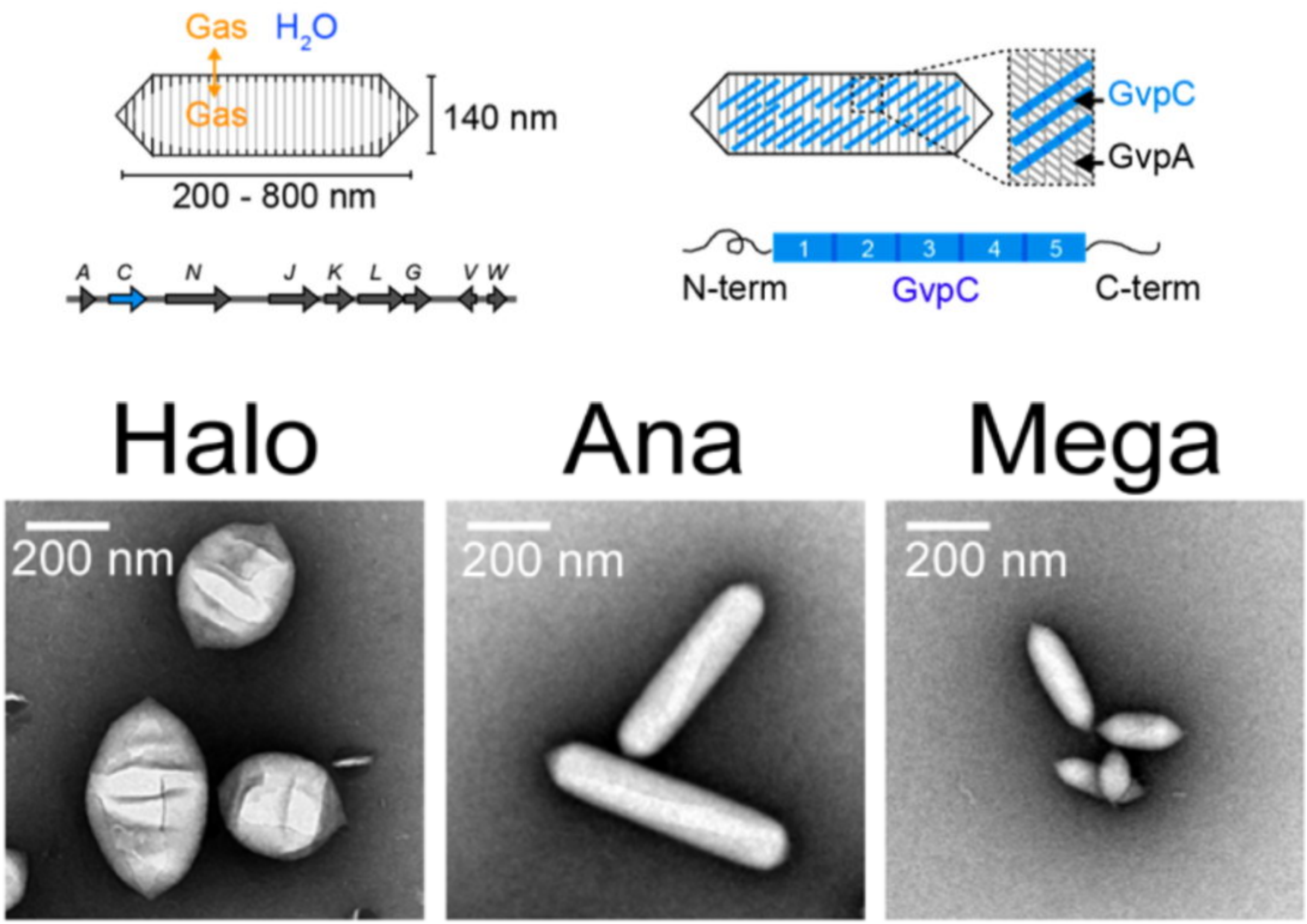
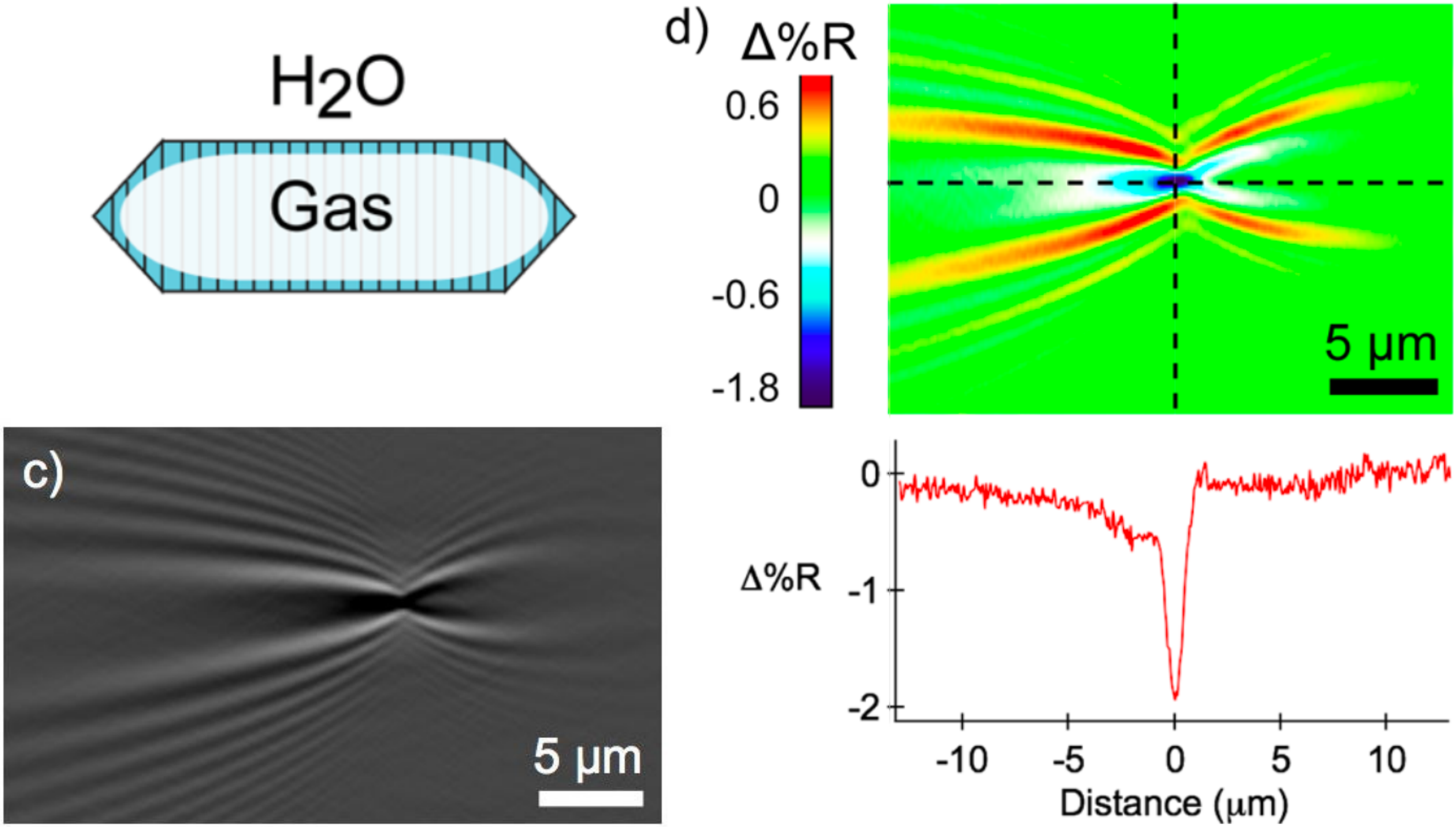
Biogenic Gas-Filled Protein Vesicles



With Prof. Mikhail G. Shapiro, Cal Tech.

*A. Lakshmanan et al., ACS Nano **10**, 7314−7322 (2016).*

Biogenic Gas-Filled Protein Vesicles



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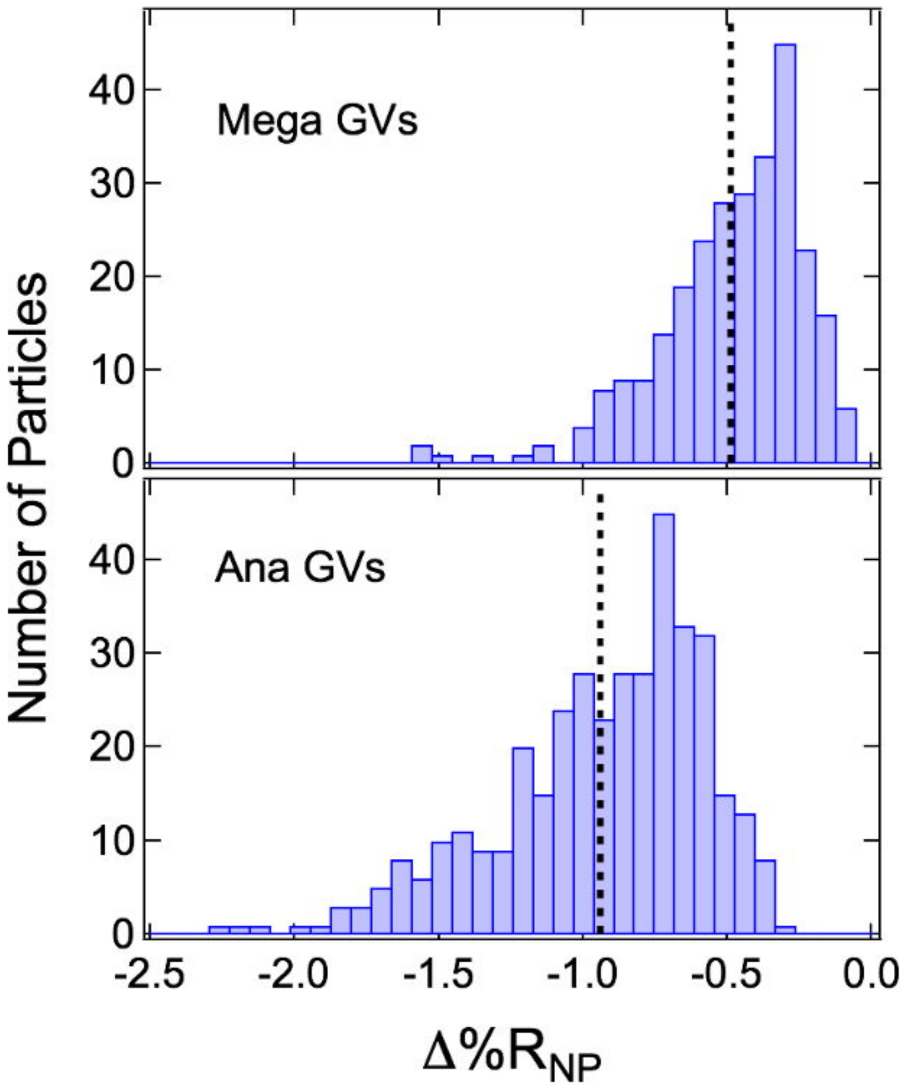
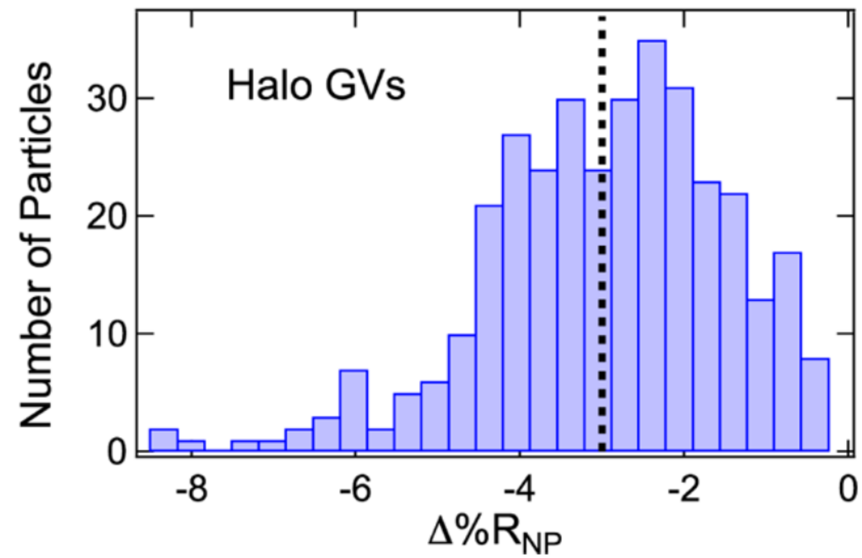
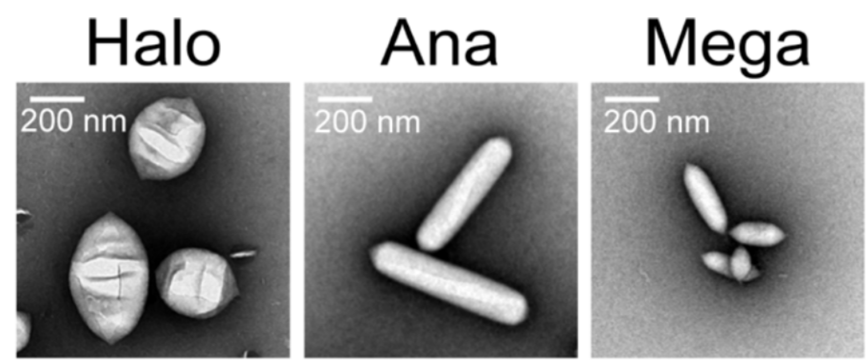


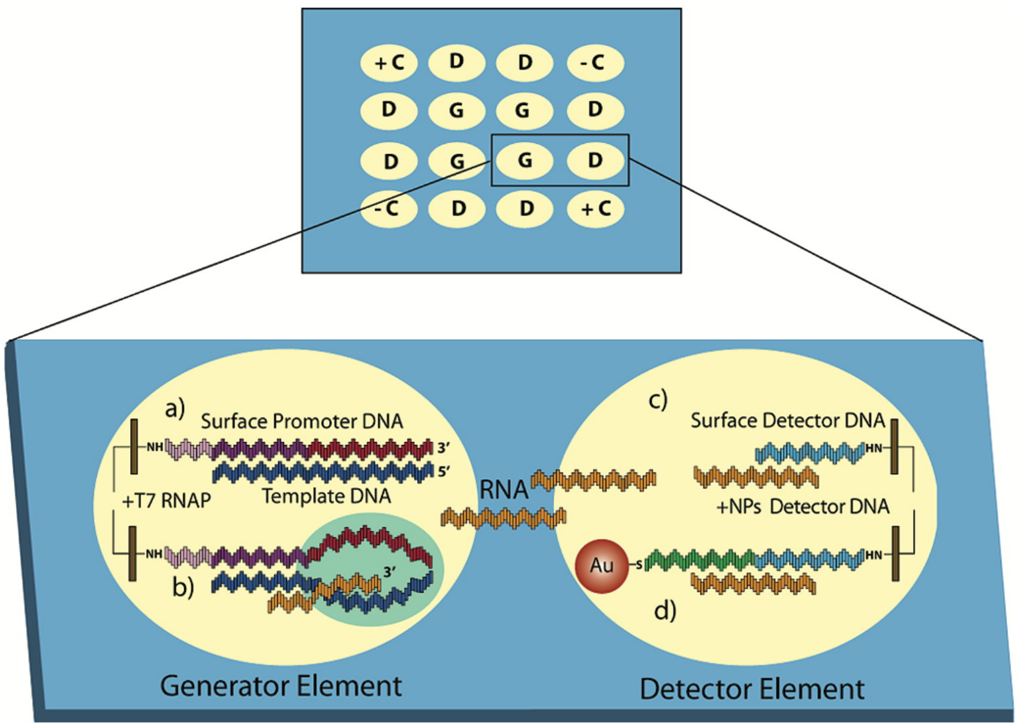


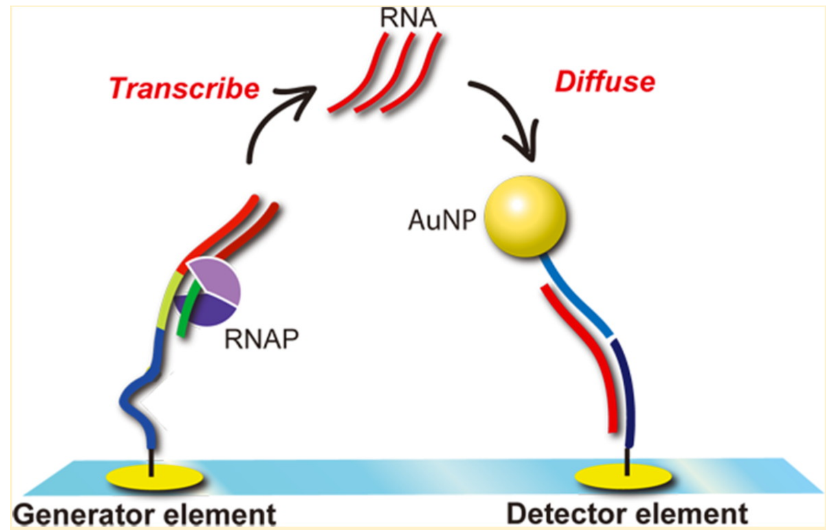


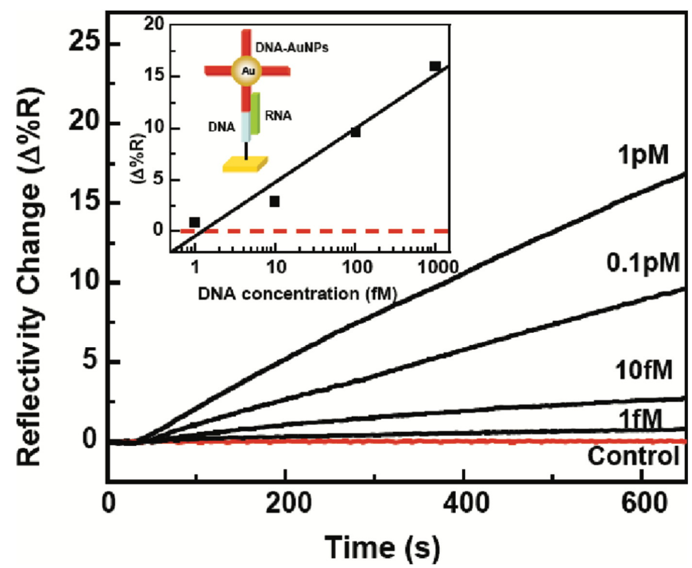
A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, ***11*** *7447-7456 (2017).*

Surface Polymerase Chemistries: RNA Polymerase Amplification

Dual Element Methodology







1 fM ssDNA detection

Iuliana Sendroiu

I. Sendroiu et al., J. Am. Chem. Soc., 133 4271-4273 (2011)

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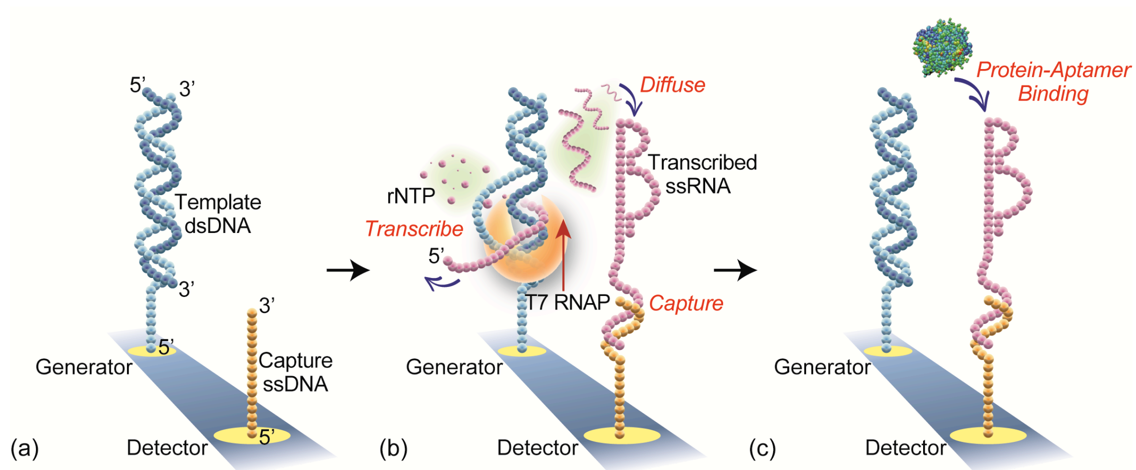
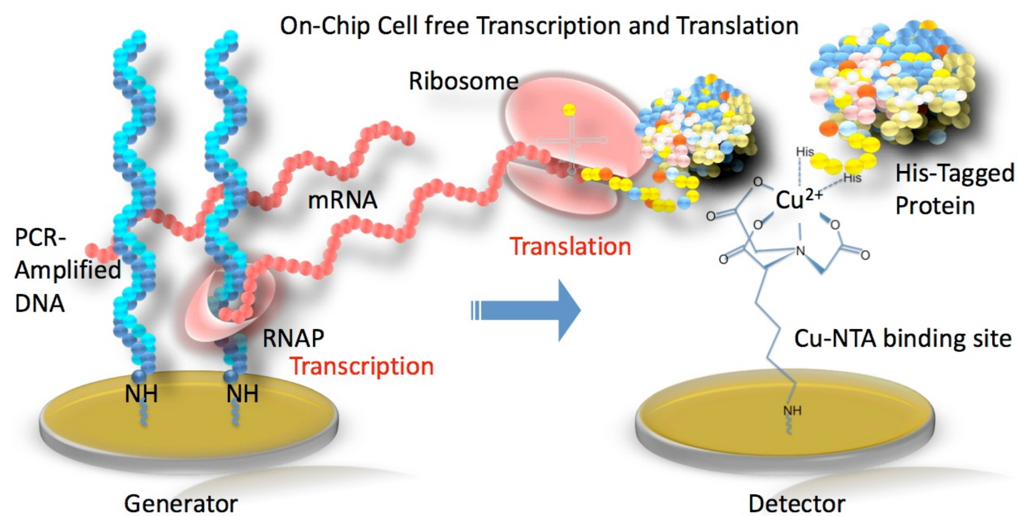
On-Chip Templated Biosynthesis of RNA and Protein Microarrays



On-Chip Synthesis of RNA aptamer microarrays

On-Chip Synthesis of Protein microarrays



Y. Chen, et al., Langmuir, ***28*** *8281-8285 (2012).*
T. H. Seefeld et al., J. Am. Chem. Soc., ***134*** *12358-12361 (2012).*

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NSF
UC-CRCC

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Prof. Ken Shea (UCI)
Prof. Reg Penner (UCI)
Prof. Andrej Luptak (UCI)
Prof. Donghyun Kim (Yonsei)

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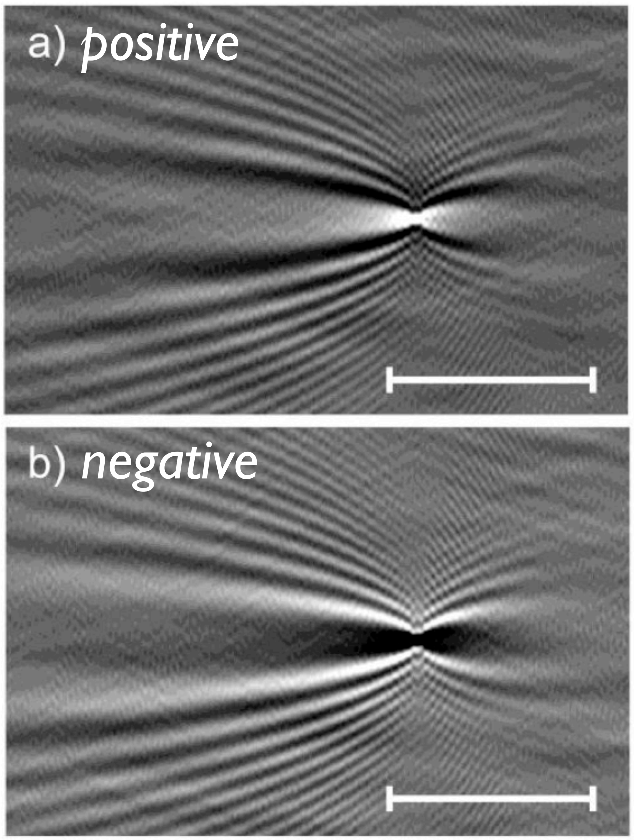
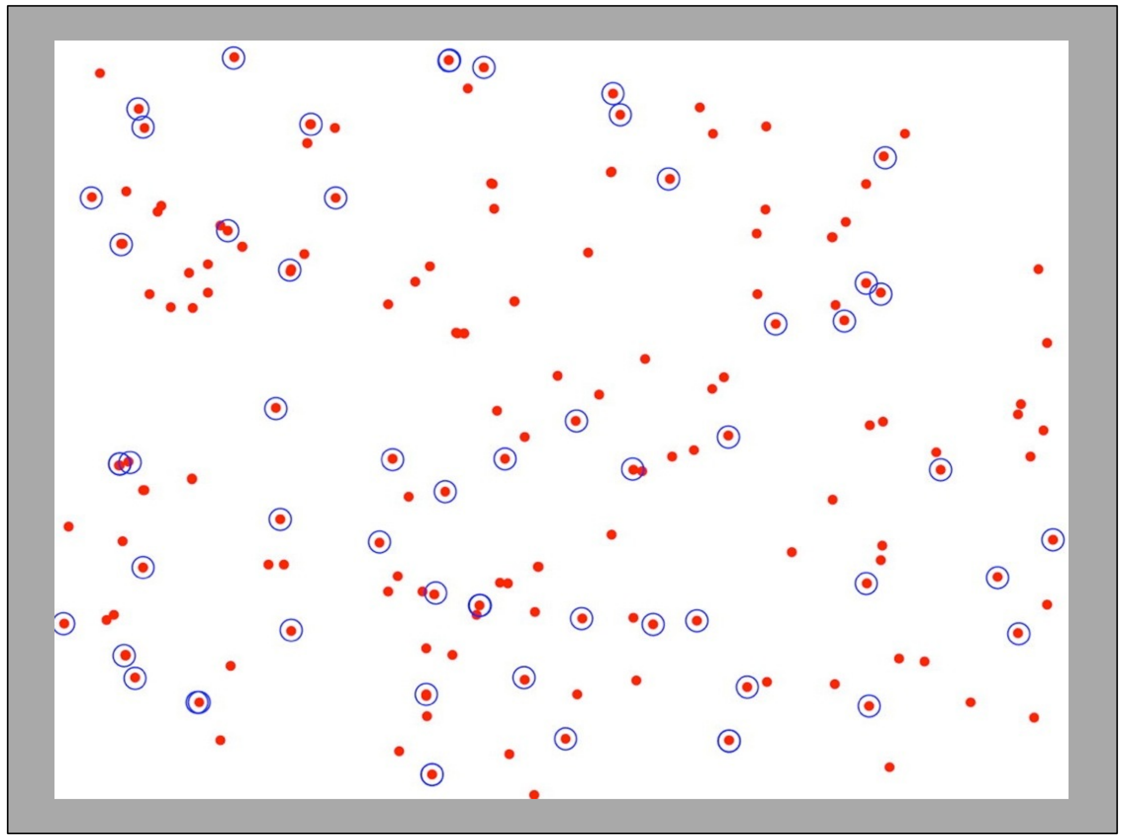
DNA nanoparticle adsorption onto single DNA sequences

How much adsorption of 40 nm DNA-modified Au Nanoparticles do we see onto DNA-modified surfaces with only a small number of binding sites?

two component complementary/non-complementary monolayer





0.01% complementary monolayer

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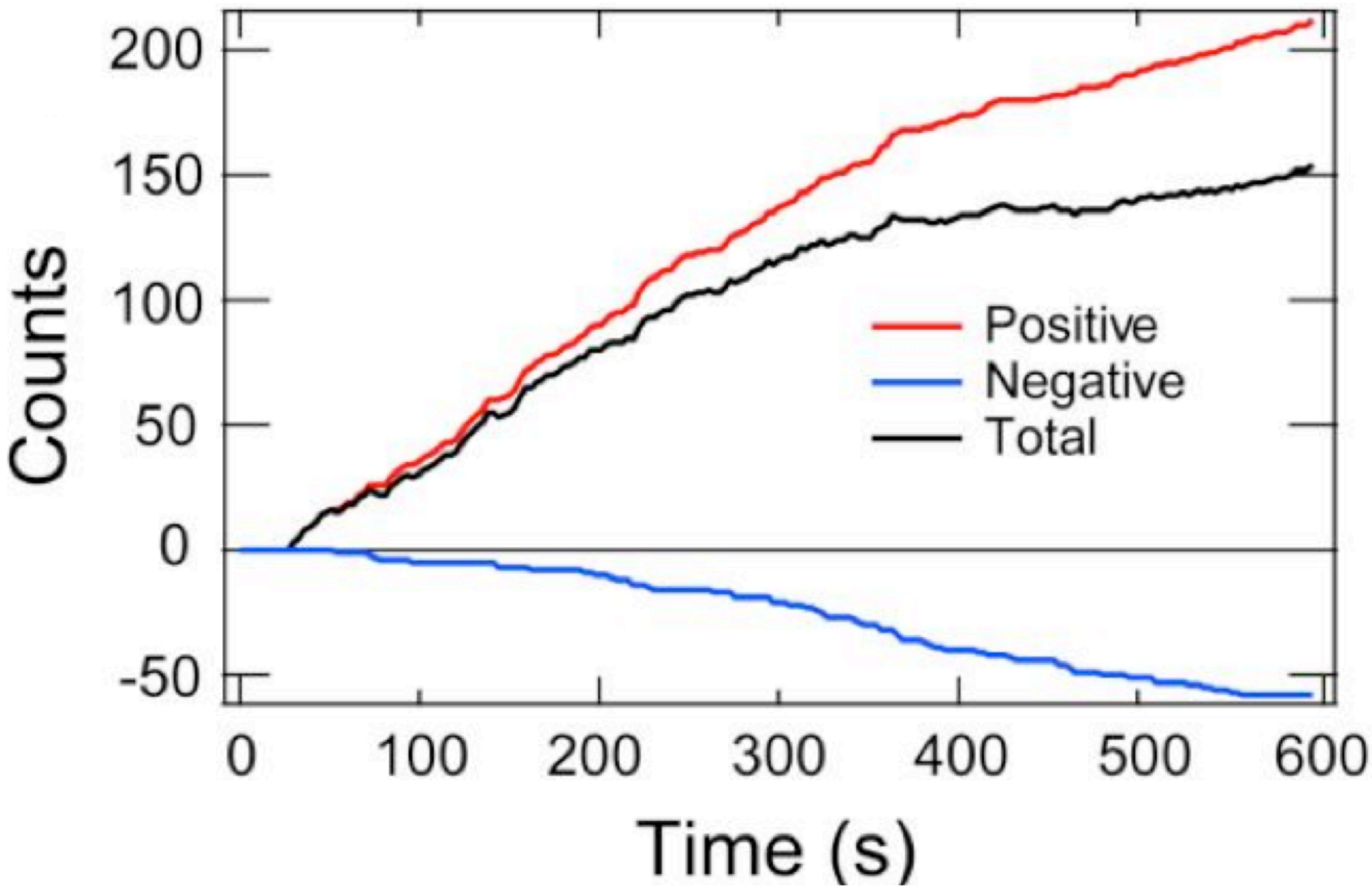

0.01% complementary monolayer

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Biogenic Gas-Filled Protein Vesicles

$$\mathrm{PDF} = \frac{1}{\Delta\%R_{\mathrm{NP}}\sigma\sqrt{2\pi}}\exp\left[-\frac{(\ln(\Delta\%R_{\mathrm{NP}}) - \mu)^2}{2\sigma^2}\right]$$

lognormal distribution function

$$g = \frac{m_3}{s^3} = \frac{\frac{1}{n}\sum(\Delta\%R_{\mathrm{NP}} - \langle\Delta\%R_{\mathrm{NP}}\rangle)^3}{s^3}$$

skewness

ACS Nano Article

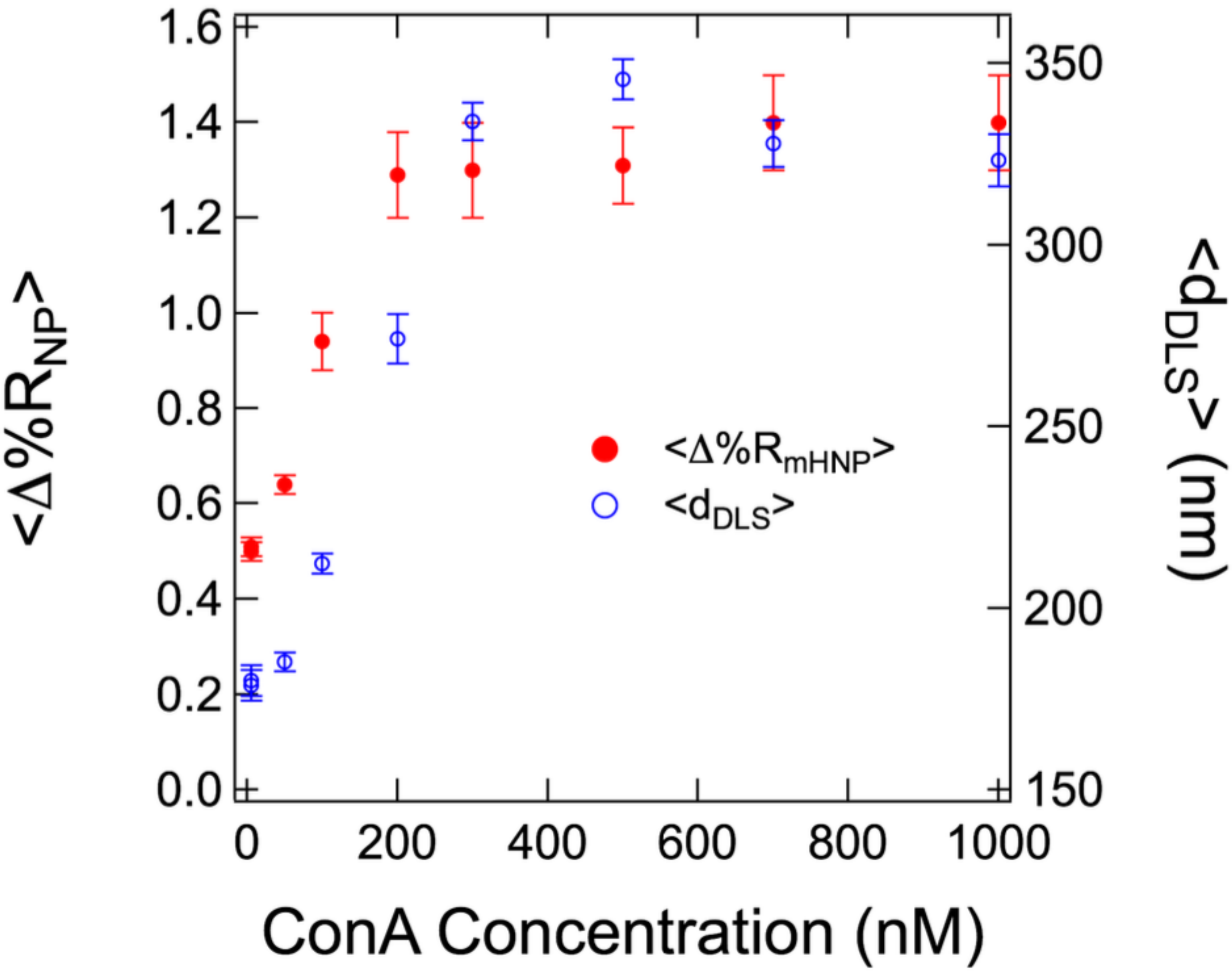
Table 1. Hydrodynamic Size Measurements from DLS for Polystyrene and Hydrogel Nanoparticles and Statistics from Single-Nanoparticle SPRI Measurements for Polystyrene and Hydrogel Nanoparticles and Gas Vesicles

nanoparticle	diameter (nm)	standard deviation (nm)	$\langle\Delta\% R_{\mathrm{NP}}\rangle$	standard deviation (s)	95% CI	skewness (g)	μ[a]	σ[b]	no. of NPs
PS (A)	85	25	0.34	0.10	0.01	0.59	−1.13	0.31	354
PS (B)	170	40	2.19	0.48	0.05	0.68	0.76	0.21	365
HNP	271	55	1.67	0.43	0.05	0.60	0.48	0.27	324
HNP + 2 μM melittin	272	65	2.79	0.52	0.08	0.02	1.01	0.20	172
HNP	272	50	0.90	0.27	0.03	0.55	−0.15	0.31	289
HNP + 500 nM ConA	357	75	3.6	1.3	0.2	0.79	1.22	0.37	307
HNP + 500 nM ConA + 1 mM Man	338	65	2.04	0.60	0.07	0.05	0.66	0.36	270
HNP + 500 nM ConA + 10 mM Man	320	55	1.74	0.41	0.05	0.30	0.53	0.24	241
Mega GV	–[c]	–	−0.49	0.26	0.03	−1.28	−0.84	0.52	274
Ana GV	–	–	−1.07	0.44	0.04	−1.53	−0.0083	0.38	395
Halo GV	–	–	−3.0	1.5	0.2	−0.74	0.95	0.58	345

[a]Log-normal PDF location parameter. [b]Log-normal PDF scale parameter. [c]Size measurements for GVs are reported in Table 2.

*A.M. Maley, G.J. Lu, M.G. Shapiro and R.M. Corn, ACS Nano, **11** 7447-7456 (2017).*

ConA uptake into HNPs



Average single nanoparticle response and DLS hydrodynamic diameter

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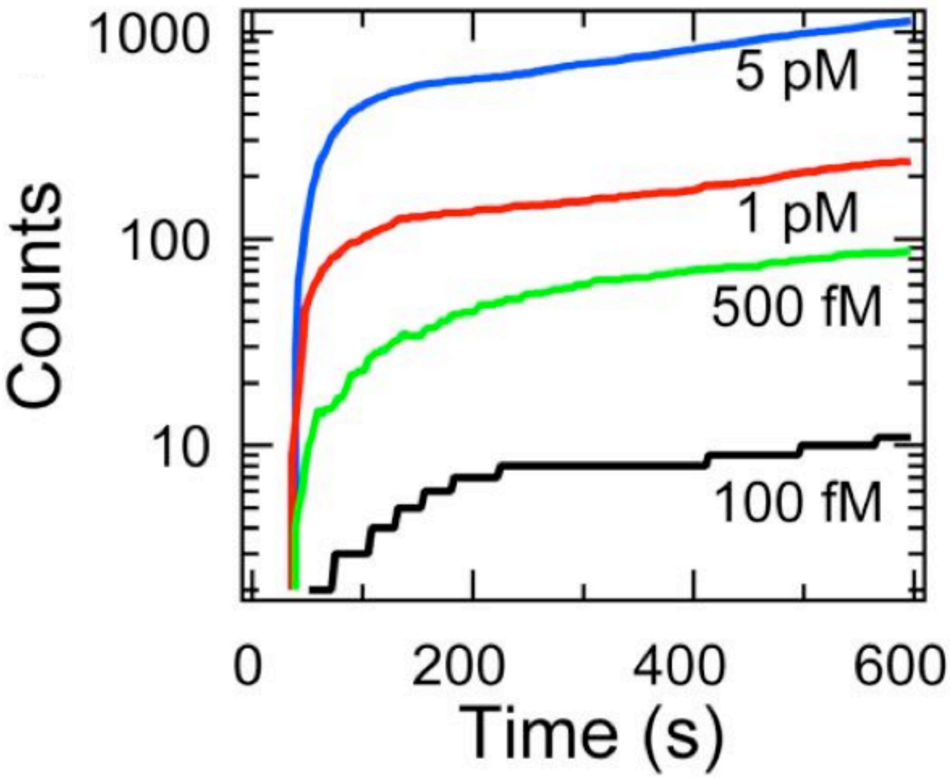
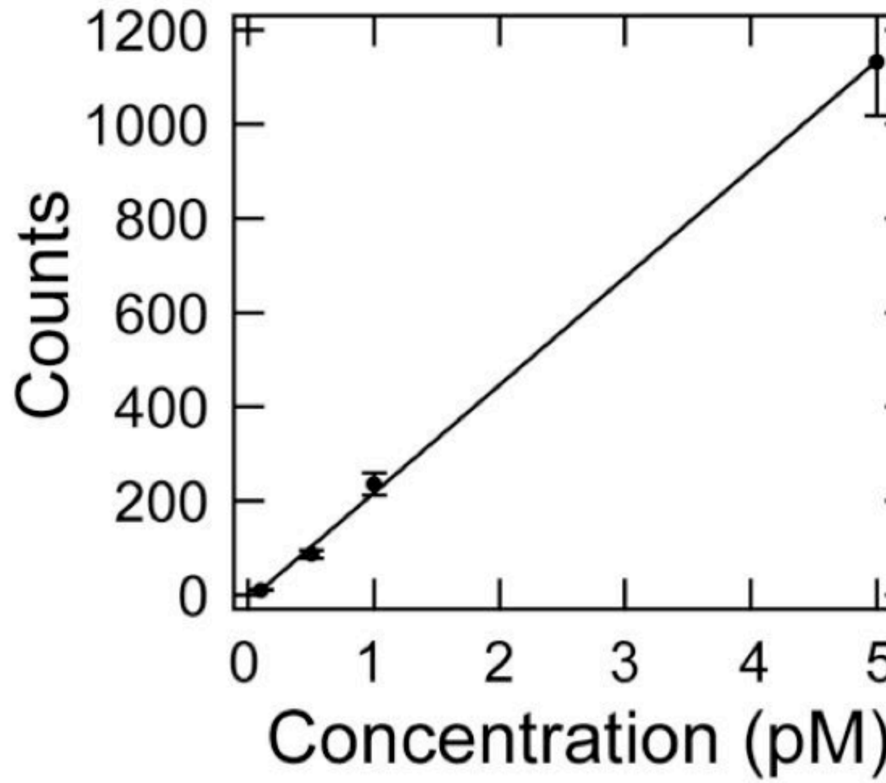
Bioaffinity Adsorption of 40 nm DNA-modified Au Nanoparticles





The number of binding events scale with Au NP concentration